A dedicated robust instrument for water vapor generation at low humidity for use with a laser water isotope analyzer in cold and dry polar regions

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Abstract. Obtaining precise continuous measurements of water vapor isotopic composition in dry places (polar or high-altitude regions) is an important challenge. The current limitation is the strong influence of humidity on the measured water isotopic composition by laser spectroscopy instruments for low humidity levels (below 3000 ppmv). This problem is addressed by determining the relationships between humidity and measured δ¹⁸O and δD of known water standards. Here, we present the development of a robust field instrument able to generate water vapor, down to 70 ppmv, at very stable humidity levels (average 1σ lower than 10 ppmv). This instrument, operated by a Raspberry interface, can be coupled to a commercial laser spectroscopy instrument. We checked the stability of the system as well as its accuracy when expressing the measured isotopic composition of water vapor on the VSMOW–SLAP (Vienna Standard Mean Ocean Water – Standard Light Antarctic Precipitation) scale. It proved to be highly stable during autonomous operation over more than 1 year at the East Antarctic Concordia and Dumont d’Urville stations.

1 Introduction

The recent development of laser spectroscopy instruments now enables the continuous measurement of the isotopic composition of water vapor at many observation stations all around the world (Bailey et al., 2015; Bastrikov et al., 2014; Schmidt et al., 2010; Sodemann et al., 2017; Tremoy et al., 2011). In particular, the isotopic composition of water vapor has proven to be a very useful tool to document moist synoptic events at many locations (Bonne et al., 2014; Guilpart et al., 2017). In polar regions, the water vapor isotopic signal is not only useful for detecting the origin of moist air (Bréant et al., 2019; Kopec et al., 2014) but also for improving the interpretation of the isotopic composition of water in surface snow and ice core archives (Steen-Larsen et al., 2014). Indeed, exchanges occur after deposition between the surface snow and the water vapor, leading to modifications to the isotopic composition of the former and, hence, of the archived ice (Casado et al., 2016, 2018; Ritter et al., 2016).

Obtaining continuous and accurate measurements of the water vapor isotopic composition expressed on the VSMOW–SLAP (Vienna Standard Mean Ocean Water – Standard Light Antarctic Precipitation) scale measurements of the water vapor isotopic composition at Concordia station in central Antarctica is a key scientific challenge, as the deep ice core drilled there, EPICA Dome C, provides the oldest continuous water isotopic record expressed on the VSMOW–SLAP scale to date (Jouzel et al., 2007). Thus, it is a key reference for the study of past climate, and a correct interpretation of the isotopic record relies on the quantification of the transfer function between climate parameters and water isotopic composition in ice, which is itself influenced by exchanges with water vapor in the upper layers of the firn (Casado et al., 2018). Such knowledge is also of the uttermost impor-
tance for the interpretation of water isotope records from the new deep drilling project “Beyond EPICA – Oldest Ice” (https://www.beyondepica.eu/en/, last access: 5 April 2021), the aim of which is to drill a 1.5 million-year-old ice core at the Little Dome C site located 40 km from Concordia station and, hence, with similar low temperature and humidity conditions.

One of the main limitations of the current commercial instruments when deployed in polar regions is their relatively poor performance at low water vapor concentration. Generally, the precision of the measured isotopic ratios, $\delta^{18}$O and $\delta^D$, rapidly worsens when the mixing ratio decreases to humidity levels below 3000–5000 ppmv (parts per million by volume) (Bonne et al., 2014; Weng et al., 2020). However, in remote continental areas in Greenland and Antarctica, temperatures in winter can drop to very low values, leading to humidity levels down to 10 ppmv (Genthon et al., 2017). Arguably one of the most extreme experiments for continuous measurement of the water vapor isotopic composition was the deployment of a commercial Picarro L2130-i instrument at the East Antarctic French–Italian station of Concordia where the mean annual temperature is around $-54^\circ$C and the humidity barely exceeds 1000 ppmv during the warmest summer days (Casado et al., 2016). For such applications, there are two major impacts of low humidity on the raw isotopic signal: first, we generally observe an apparent increase in $\delta^{18}$O and $\delta^D$ with decreasing humidity level; second, the standard deviation associated with the continuous measurements of $\delta^{18}$O and $\delta^D$ of the water vapor increases. This can lead to overall uncertainties of several per mil for $\delta^{18}$O and tens of per mil for $\delta^D$. Therefore, it is of utmost importance to have a correct determination of the humidity dependency of the water vapor isotopic ratios.

Commercial instruments from Picarro Inc. are usually associated with a Picarro standards delivery module (SDM) designed to generate humidity at stable levels between 5000 and 30 000 ppmv. Using such a setup for humidity levels below 5000 ppmv leads to large uncertainties in the determination of the humidity influence on the water vapor isotopic composition (e.g., Guilpart et al., 2017). These uncertainties are due both to the instability of the water vapor generation using the SDM (in terms of water concentration – humidity and/or isotopic composition) and to the analytical noise in the spectroscopy measurements when the absorption signals are weak. An alternative commercial device is the LGR (Los Gatos Research) calibration system (Water Vapor Isotope Standard Source, WVISS), which uses a nebulizer to instantaneously evaporate microdroplets of liquid water from a standard reservoir into a large (1 L) vaporizing chamber (Dong and Baer, 2010). This system is very stable and well adapted for a humidity range between 2500 and 25 000 ppmv (Aemisegger et al., 2012).

Several homemade water vapor injection systems have been developed with the specific aim of achieving better generated humidity stability at low humidity levels. A first approach is to use a dew point generator that injects small amounts of water into dry air (Lee et al., 2005; Wang et al., 2009). This approach is time-consuming, as it takes a long time to reach equilibrium and relies on a very precise knowledge of the temperature to quantify the isotopic fractionation. A method using a piezoelectric microdroplet generator into a dry-air stream could generate water mixing ratios between 12 and 3500 ppmv (Iannone et al., 2009; Sturm and Knohl, 2009; Sayres et al., 2009). However, adjustment of the humidity level and long-term stability are difficult to obtain with such devices. Systems relying on the use of syringe pumps have also been built by Gkinis et al. (2010) and Tremoy et al. (2011): a small fraction of the input stream of liquid water is introduced into a hot oven where water is vaporized in the presence of a dry-air flow. These systems cover a humidity range between 2000 and 30 000 ppmv. Finally, bubbler systems, in which dry air flows through a large volume of water to create saturated vapor, are very robust but can only produce water vapor at high humidity levels (Ellehoj et al., 2013). The aforementioned devices are unfortunately not well suited for automatic long-term operation at low humidity levels. During the 2014–2015 summer field season at Concordia station in Antarctica, a homemade humidity generator specifically designed for low humidity levels (Landsberg, 2014) was deployed (Casado et al., 2016). The device used dual high-precision, low-volume, syringe pumps to generate stable humidity levels at two different isotopic compositions over the range from 100 to 800 ppmv (Casado et al., 2016). Unfortunately, we observed quite a large scattering among the isotopic values measured at similar humidity levels as well as a large discrepancy between the humidity dependency of the water isotopic ratios measured in the field and that measured in the laboratory. Upon return to the laboratory, these defaults were traced primarily to tiny leaks in the water supply lines to the syringes.

Therefore, we reengineered the prototype by Landsberg (2014) in order to develop a robust and autonomous device for stable low-level humidity generation for the purpose of precise humidity calibration of spectroscopic instruments. Such devices have now been operating with minimum manual intervention for more than 1 year at two polar stations in Antarctica, Dumont d’Urville and Concordia, coupled to Picarro laser spectroscopy instruments. Here, we detail the technical description of the instrument and show key performance characteristics, enabling, for instance, a discussion of small-amplitude signals such as the diurnal variability of the water vapor isotopic composition in remote dry sites in East Antarctica.

2 New vapor generator for low humidity levels

The low humidity level generator (LHLG) developed here relies on the same principle as the one developed by Landsberg (2014), i.e., a steady, undersaturated evaporation of a liquid

2.1 Physical principle

The LHLG is based on the undersaturated evaporation of a small droplet at the tip of a needle (Fig. 1). Liquid water is pushed through a needle around which dry air is flowing. Dry air is obtained from a bottle of high-purity synthetic air with pressure regulation through two manometers connected in series. The mass flux of water \( f_L \) is kept low compared with the air mass flow \( f_A \) so that the relative humidity (RH) of the downstream moist air flow remains low (RH < 0.1). Therefore, the air stays largely undersaturated, and its humidity is controlled only by the flow of liquid water in the needle and that of the dry air upstream of it. The mixing ratio (or humidity) of the air as classically provided by a Picarro instrument is given by

\[
MR = \frac{d_{H_2O} \times f_L \times R \times T_{st}}{f_A \times P_{st} \times M_{H_2O}},
\]

where \( d_{H_2O} = 1000 \text{ kg m}^{-3} \) is the density of water, \( R = 8.314 \text{ J mol}^{-1} \text{ K}^{-1} \) is the universal gas constant, \( T_{st} = 293.15 \text{ K} \) and \( P_{st} = 1013.25 \text{ hPa} \) are standard conditions of temperature and pressure, and \( M_{H_2O} = 18.10^{-3} \text{ kg mol}^{-1} \).

Physically, when the flux of water or air is changed, there is first a transient regime during which the radius of the droplet changes, modifying the evaporative surface and, therefore, the humidity of the outgoing air. Once a stationary regime is reached, the radius of the droplet is stabilized and the humidity is given by Eq. (1). In this regime, there is no accumulation of water molecules in the system; therefore the isotopic composition of the vapor produced is equal to the isotopic composition of the liquid water injected in the needle: \( R_V = R_L \) (note that because of the fractionation during the transition phase, the isotopic composition of the droplet \( R_D \) is different from \( R_L \) and \( R_V \); see Kerstel, 2020). When changing the flux of evaporating water, we modify the size of the evaporating surface and, therefore, the radius of the drop. The evolution of the radius of the drop can be obtained from the resolution of a nonlinear differential equation of the volume \( V \) of the drop:

\[
dV/dt = f_L - f_{evap},
\]

where \( f_{evap} = k_e \times S \) is the evaporation flux depending on \( k_e \) (the evaporation rate) and \( S \) (the surface area of the drop exposed to the dry air). A good approximation is to consider the shape of the drop as a fraction of a sphere of variable radius intercepted by the surface of a disk of constant radius (the syringe tip). By numerically solving differential Eq. (2), it is possible to faithfully simulate the behavior of the device under changing conditions (Kerstel, 2020). This numerical approach validates the theoretical explanation of the undersaturated evaporation of the droplet. Importantly, it is noted that in steady state, as is the case for our application, the isotopic composition of the generated humid air is identical to that of the injected water stream and, therefore, does not depend on the infusion rate nor the specific humidity.

2.2 Instrument conception

2.2.1 Technical realization

As the LHLG relies on operating in a stationary regime, it is important that the dry-air input and the water input are steady. Thus, the air and water fluxes as well as the air pressure in the evaporation chamber are regulated by electronic PID (proportional–integral–derivative) regulators. Temperature intervenes through its effect on fractionation and the evaporation rate (apart from a negligible effect on the flow controller stability), which could lead to a departure from steady-state operation. For these reasons, the temperature of the evaporation chambers was maintained at 20°C (within 1 °C over 24 h).

The dry-air flux is regulated by a high-precision mass flow controller (Vögtlin GSC-A9TS-DD22), which has an operating range from 6 to 600 sccm (standard cubic centimeters; SD cm3 min−1) and an accuracy of 3.3 sccm. The water flux is regulated by a high-precision syringe pump (Harvard Apparatus Pump 11 Pico Plus Elite Dual), which can produce a water flow down to 10.8 pL min−1 with an accuracy of 0.35 % using syringes with a volume ranging from 10 to 250 µL. We operate in the routine mode with a dry-air flow of 300 sccm and a water flow between 0.02 and 0.5 µL min−1 using mainly 50 or 100 µL syringes. The syringe pump is equipped with two syringes that provide two water flows into two evaporation chambers in parallel (Fig. 2). Each syringe is connected to a water reservoir and to an evaporation chamber by a double three-way liquid valve (Rheodyne MXX777603) switching from an “infuse” mode to a “withdraw” mode to refill the syringes. The water in the water reservoir is sampled every month to check its isotopic composition and is renewed when the level of water is below half the maximum level. A maximum evolution of the isotopic composition of the lab standard filling the water reservoirs has been observed.
as 0.05 ‰ and 0.5 ‰, respectively, for δ¹⁸O and δD over a 2-month period.

A major change to the instrument designed by Landsberg (2014) is the introduction of the double three-way valve with leak-tight connections and an internal volume of 1.9 µL. This modification is an important improvement as it enables automatic handling of the lab standards from a reservoir to the evaporation chamber with a robust connection, avoiding in particular potential air bubbles in the water flow. Indeed, the compressibility of air bubbles trapped in the water flow can lead to flow irregularities via the amplification of small nonlinearities in the progression of the syringe plunger. This would lead to non-steady-state operation, which, in turn, would create artifacts in the humidity and isotopic composition, reducing the performance of the calibration device (see Kerstel, 2020). In addition, the three-way valve provides the opportunity for a “withdrawn” mode in which the syringes draw lab-standard water from a reservoir. When equipped with 100 µL syringes, the instrument can operate for several hours up to 1 day between refills. With the addition of the auto-refill option and the effective suppression of bubbles, the instrument can be used unattended for many months, as required for an Antarctic winter field campaign.

The evaporation chambers are stainless-steel cylinders equipped with specific connectors (Swagelok Ultra-Torr SS-4CD-TW-25) holding silicon rubber septa through which needles are inserted toward the middle of the chamber. The pressure in both chambers is regulated by a pressure controller (Bronkhorst P-702CV-1K1A-AAD-22-V) with a precision of 3 mbar in a range from 0 to 1000 mbar. This pressurization of the two chambers combined with the relatively high flow (higher than required by the infrared spectrometers) enables a steady state to be maintained irrespective of whether or not the infrared spectrometer is connected and also increases the time efficiency of calibration procedures. The spectrometer is not sensitive to the inlet pressure, and the precision of the pressure controller is not an essential aspect. On the contrary, the precision of the flow controller is key for the precision of the humidity level produced by the instrument: it is 1 % for the air flow, which is comparable to the precision of the measurement of the humidity level with the optical spectrometer. When the instrument is connected to the infrared spectrometer, the excess humid air flow is exhausted to the room through the pressure pump, and the spectrometer only pumps what is required (Fig. 2).

The control of the instrument is ensured by a Raspberry Pi that can be interfaced to a Picarro water analyzer (L2130-i in our case) in sequencer mode (see below). The hardware was designed to meet the specifications dictated by field conditions: (1) all components are fixed in a transportable case (except the dry-air bottle), isolated from vibration by an anti-vibration foam; (2) a panel of connectors (e.g., HDMI, USB and Ethernet) ensures the accessibility to the instrument when it is closed; (3) the electrical and electronic parts (e.g., power supply and Raspberry Pi) are separated from the rest of the instrument (e.g., sensors and gauges). Both the electrical and electronic parts are fully and easily accessible in case of failure.

2.2.2 Software details

The control software has been developed using open-source Python libraries and homemade drivers, including a user interface displaying the state of relevant components and the value of the different sensors. The software (HumGen) can be downloaded online (https://github.com/ojsd/humgen, last access: 4 April 2021; https://zenodo.org/record/4003465#YF82sz8o-Uk, last access: 4 April 2021).

The LHLG can operate in eight different states, with each state representing a specific setup for each element (valve position, syringe pump infusion rate, dry-air flow rate and pressure). These eight states can be divided into three categories:
a routine mode, an expert mode and a humidity-isotopes calibration mode. The simple mode is composed of six predefined states referring to the classic isotopic calibration in everyday routine operation (Table 1): (1) measurement of the outside air water vapor isotopic composition; (2) drying of the cavities; (3) “humidity boost”, in order to more quickly reach the desired humidity level in the cavities; (4) injection of Standard A in the corresponding evaporation chamber at a set humidity level; (5) injection of Standard B in the corresponding evaporation chamber; (6) refill of the syringes. The expert mode is useful to adjust each parameter manually: flow rates on controllers FCA and FCB, opening of electrovalves A and B, mode (infuse or withdraw) and infused rate for the syringe pump, pressure regulation, state of the double three-way valve, activation of the pressure pump at the exhaust, and opening of external electrovalves from the dry-air tank and to the inlet (Fig. 2). The humidity-isotopes calibration mode produces a scale of increasing humidity steps in the evaporation chambers (e.g., from 100 to 1000 ppmv, through steps of 100 ppmv for 50 min for each standard). The details of the sequence (standard type, humidity level and duration of each step) is defined in a text file by the operator from the Raspberry interface, with the Raspberry itself being connected to Ethernet for remote access.

The Picarro L2130-i analyzer has an external valve sequencer, which is able to turn up to six electrovalves on or off and create loop sequences with defined durations for each step of the sequence. This tool can be diverted from its original purpose by using it as a six-digit code: each of the humidity generator states is associated with a code. When the Picarro valve sequencer matches one of the state codes, this state is triggered on the humidity generator. This eases both the operator’s activities and the data post-treatment, because the current valve status – thus the calibration instrument state – is saved in the analyzer output data file, in the “Valve-Mask” column. The Raspberry inside the LHLG reads the valve sequencer state code using the Picarro’s remote control interface (a RS232 serial connection through one of the rear-facing DB9 connectors).

A set of tools has been developed to quickly check daily calibration. In the field, analyzer and LHLG data are archived daily and sent to the laboratory, i.e., at Laboratoire des Sciences du Climat et de l’Environnement (LSCE), Gif-sur-Yvette. They are checked semi-automatically once a week to warn maintenance personnel in the event of a malfunction.

3 Performance of the instrument

The stability of the instrument has been tested over a large range of parameters. We show an example in Table 2. We modified the air flow associated with Standard A (the same results can be obtained with Standard B) between 200 and 400 sccm with an air flow on channel B of half the value of channel A. The infusion rate was varied between 0.03 and 0.14 µL min\(^{-1}\) in order to produce humidity levels of 400 and 800 ppmv. The 1σ standard deviations observed over 10 min plateaus are comparable to the standard deviation obtained when the air flow is set to 300 sccm.

For routine measurement, the air flow and infusion rate have been adjusted to optimize the stability of the generated vapor while minimizing the dry-air consumption. Thus, the LHLG is able to generate stable levels of humidity (drift lower than 20 ppmv over 1 h and 1σ below 10 ppmv over 10 min) from 70 to 2400 ppmv following the optimal set points shown in Table 3.

3.1 No fractionation during water vaporization in the cavity

We checked that there was no fractionation of the water during its transfer from the bottles to the syringe pump and then from the syringe to the moist air generated in the vaporization chamber through the following tests.

First, the isotopic compositions of three different lab standards calibrated against VSMOW at LSCE (\(\text{H}_2\text{O}–\text{CO}_2\) equilibrium followed by isotope ratio mass spectrometry for \(\delta^{18}\text{O}\); cavity ring-down spectroscopy for \(\delta\text{D}\); calibrated every 3 years using VSMOW and SLAP provided by the International Atomic Energy Agency) were compared, after their generation by the present LHLG and by the commercial SDM, both at a humidity of 2000 ppmv over 50 min time spans. The measured \(\delta^{18}\text{O}\) and \(\delta\text{D}\) values agreed to within 0.5%e and 2%e, respectively, for the three lab-standard waters calibrated against VSMOW: EPB (\(\delta^{18}\text{O} = −6.24\%e; \delta\text{D} = −43.6\%e\)), NEEM (\(\delta^{18}\text{O} = −33.50\%e; \delta\text{D} = −257.2\%e\)) and FP5 (\(\delta^{18}\text{O} = −48.33\%e; \delta\text{D} = −383.5\%e\)). Second, the measured isotopic composition of the same standard (FP5) generated at different humidity levels between 1000 and 2400 ppmv by the SDM and the LHLG show the same \(\delta^{18}\text{O}\) (\(\delta\text{D}\)) evolution with humidity within the respective uncertainties (Fig. S1 in the Supplement).

3.2 Stability of the water vapor delivery and associated water isotopic composition

A proper approach to quantify the stability of our system is to use the Allan variance, which is defined as

\[
\sigma_y^2(t) = \frac{1}{2} \left( (y_{n+1} - y_n)^2 \right).
\]

where \(y_n\) are the successive measurements over a period \(t\).

An Allan variance plot as a function of averaging time is indeed useful to determine the optimal time over which the sample humidity and the isotopic composition should be averaged to obtain a precise determination (low standard deviation) and avoid drift. Figure 3 displays the Allan deviation (square root of the Allan variance) in \(\delta^{18}\text{O}\), \(\delta\text{D}\) and humidity obtained by running a long plateau of Standard A or Standard B in the “infuse” mode over 4 h for different humidity

levels. The humidity variance always stays below 10 ppmv over the 4 h test, and the δ^{18}O and δD Allan deviations display minimum values below 1 ‰ and 7 ‰, respectively. The minimum value for the δ^{18}O and δD Allan deviation is generally obtained for about 15 min of measurement. While the Allan deviation of δ^{18}O and δD is dependent on the analyzer used, we observe that the Allan deviation at 1000 s (17 min) for δ^{18}O and δD also depends to some extent on the humidity level: the lowest levels are obtained for humidity levels of 770–1080 ppmv, and the highest levels are obtained for humidity level of 170 ppmv.

In the routine mode (Fig. 4), we perform plateaus of 30 to 50 min (50 min when the instrument is unattended as the time to reach the plateau varies between a few minutes and 30 min). We then select the last 10 min before the following switch of the instrument to measure the average level of humidity and the isotopic ratios, δ^{18}O and δD. We also calculate the associated standard deviations and reject the values if the humidity standard deviation exceeds 30 ppmv over these last 10 min. In Fig. 4, one observes that the standard deviations for humidities generated in the routine mode are actually much lower. The corresponding standard deviations for the isotopic ratios (δ^{18}O and δD; see values indicated in Fig. 4) increase with decreasing humidity, reflecting the decrease in the molecular absorption signal recorded by the L2130-i laser analyzers. This has an obvious impact on the determination of the relationship between humidity and water vapor isotopic composition.

The performance of the present LHLG can be compared to the performance of the SDM (see Supplement Figs. S1

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**Table 1.** Typical routine sequence of measurements and calibration for two standards A and B at 1000 ppmv for a measurement site located at sea level. No mixing occurs between standards A and B during steps “Standard A” and “Standard B” (see Supplement). Note that the humidity-isotopes mode and the expert mode can also be included in the valve sequencer but are not used in a daily calibration routine.

<table>
<thead>
<tr>
<th>States (min)</th>
<th>Flow FCA (sccm)</th>
<th>Flow FCB (sccm)</th>
<th>Valve A</th>
<th>Valve B</th>
<th>Syringe pump (µL min(^{-1}))</th>
<th>Inlet valve</th>
<th>Dry-air valve</th>
<th>Pressure controller for exhaust (mbar)</th>
<th>Pressure pump for exhaust (mbar)</th>
<th>Double three-way valve</th>
</tr>
</thead>
<tbody>
<tr>
<td>Outside air (1100)</td>
<td>0</td>
<td>0</td>
<td>Closed</td>
<td>Closed</td>
<td>0</td>
<td>Open</td>
<td>Closed</td>
<td>Off</td>
<td>Off</td>
<td>To chamber</td>
</tr>
<tr>
<td>Drying (20)</td>
<td>400</td>
<td>400</td>
<td>Open</td>
<td>Open</td>
<td>0</td>
<td>Closed</td>
<td>Open</td>
<td>Off</td>
<td>Off</td>
<td>To chamber</td>
</tr>
<tr>
<td>Boost (0.7)</td>
<td>300</td>
<td>300</td>
<td>Open</td>
<td>Open</td>
<td>Infuse at 2.5</td>
<td>Closed</td>
<td>Open</td>
<td>905</td>
<td>On</td>
<td>To chamber</td>
</tr>
<tr>
<td>Standard A (50)</td>
<td>300</td>
<td>150</td>
<td>Open</td>
<td>Closed</td>
<td>Infuse at 0.25</td>
<td>Closed</td>
<td>Open</td>
<td>905</td>
<td>On</td>
<td>To chamber</td>
</tr>
<tr>
<td>Standard B (50)</td>
<td>150</td>
<td>300</td>
<td>Closed</td>
<td>Open</td>
<td>Infuse at 0.25</td>
<td>Closed</td>
<td>Open</td>
<td>905</td>
<td>On</td>
<td>To chamber</td>
</tr>
<tr>
<td>Reset (1)</td>
<td>Closed</td>
<td>Closed</td>
<td>Closed</td>
<td>Closed</td>
<td>Withdraw max speed</td>
<td>Open</td>
<td>Closed</td>
<td>Off</td>
<td>Off</td>
<td>From standard</td>
</tr>
</tbody>
</table>
Table 2. Evolution and stability of humidity and δ¹⁸O (same water used for the different tests) for different syringe infusion rates and dry-air flows.

<table>
<thead>
<tr>
<th>Air flow (sccm)</th>
<th>Infusion rate (µL min⁻¹)</th>
<th>Humidity (ppmv)</th>
<th>10 min 1σ standard deviation for humidity (ppmv)</th>
<th>δ¹⁸O (‰)</th>
<th>10 min 1σ standard deviation for δ¹⁸O (‰)</th>
</tr>
</thead>
<tbody>
<tr>
<td>200</td>
<td>0.07</td>
<td>808</td>
<td>1 -7.88</td>
<td>0.89</td>
<td></td>
</tr>
<tr>
<td>300</td>
<td>0.11</td>
<td>851</td>
<td>2 -7.73</td>
<td>0.85</td>
<td></td>
</tr>
<tr>
<td>400</td>
<td>0.14</td>
<td>818</td>
<td>2 -7.95</td>
<td>0.90</td>
<td></td>
</tr>
<tr>
<td>200</td>
<td>0.03</td>
<td>374</td>
<td>1 -8.45</td>
<td>1.92</td>
<td></td>
</tr>
<tr>
<td>300</td>
<td>0.05</td>
<td>411</td>
<td>2 -9.16</td>
<td>1.64</td>
<td></td>
</tr>
<tr>
<td>400</td>
<td>0.07</td>
<td>415</td>
<td>3 -9.05</td>
<td>1.59</td>
<td></td>
</tr>
</tbody>
</table>

Figure 4. Records of δ¹⁸O, δD and humidity over three humidity plateaus (72 ppmv on the left, 425 and 335 ppmv on the right) obtained with the LHLG. The gray rectangles indicate the period (10 min) over which the average values are kept for calibrating the data generated by the L2130-i analyzer.

Table 3. Set points for the water infusion rate and dry-air flow at a temperature of 20°C.

<table>
<thead>
<tr>
<th>Humidity (ppmv)</th>
<th>Infusion rate (µL min⁻¹)</th>
<th>Dry-air flow (sccm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>80</td>
<td>0.01</td>
<td>300</td>
</tr>
<tr>
<td>160</td>
<td>0.02</td>
<td>300</td>
</tr>
<tr>
<td>320</td>
<td>0.04</td>
<td>300</td>
</tr>
<tr>
<td>800</td>
<td>0.1</td>
<td>300</td>
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<tr>
<td>1200</td>
<td>0.15</td>
<td>300</td>
</tr>
<tr>
<td>1600</td>
<td>0.2</td>
<td>300</td>
</tr>
<tr>
<td>2400</td>
<td>0.3</td>
<td>300</td>
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</tbody>
</table>

and S2). First, as seen in Fig. S2, a comparison has been performed at a humidity level of 800 ppmv, for which we have numerous daily calibrations performed with a SDM from a 4.5-year field deployment in Svalbard (Leroy-Dos Santos et al., 2020). The best SDM performance displays a standard deviation 1σ of 31 ppmv, which is significantly worse than the performance of the LHLG (standard deviation 1σ lower than 10 ppmv on average and down to 2 ppmv for 30 % of the generated humidity plateaus). Second, as seen from Fig. S1, while we measure the same influence of humidity on measured δ¹⁸O and δD either with the SDM or with the LHLG, the 1σ values on humidity levels are much larger for the SDM than for the LHLG.

3.3 Determination of the influence of humidity on water vapor isotopic composition

Contrary to the commercial SDM, which hardly produces stable and reproducible humidity levels below 500 ppmv, the LHLG was able to produce stable 10 min humidity plateaus daily over the range from 70 to 2400 ppmv with an associated standard deviation of the order of 10 ppmv over more than 1 year at the Concordia and Dumont d’Urville stations (installation in December 2018). The stability of the LHLG allows a robust quantification of the L2130-i analyzer drift thanks to a daily measurement of the same water isotopic standard reference (see Table S2 which shows no measurable drift over a 3-week period). It also permits the characterization of the measurement nonlinearities observed at low humidity (Fig. 5). The more than 1-year-long Concordia and Dumont d’Urville datasets showed that the humidity dependence of δ¹⁸O and δD did not vary measurably. The uncertainty of the obtained calibration curve can be attributed entirely to the L2130-i δ¹⁸O and δD measurements.
In other words, the uncertainty bars on the horizontal (x) axis in Fig. 5, associated with the LHLG, are negligible.

Our data show a result already observed in Weng et al. (2020): while the dependency of $\delta^{18}O$ and $\delta D$ on humidity is similar for low $\delta^{18}O$ and $\delta D$ lab standards (NEEM and FP5), we observe a different behavior for the $\delta D$ vs. humidity relationship for the high $\delta^{18}O$ and $\delta D$ lab standard (EPB). This result strengthens the recommendation of Weng et al. (2020) to use two water standards in the range of the measured water vapor isotopic composition to best calibrate our final data. In our case, our applications were in Antarctica; thus, we used our two lowest lab standards (NEEM and FP5). For the two standards and for this particular Picarro L2130-i (results are expected to depend on the instrument), the same dependency of isotopic composition vs. humidity is observed. We express this dependency as the relationship between the difference in $\delta D$ or $\delta^{18}O$ between the measured value at the given humidity and the value of the same standard measured at a humidity of 2000 ppmv. The experimental data for NEEM and FP5 from Fig. 5 are fitted through polynomial functions with respect to humidity $h$ (in ppmv):

$$\delta^{18}O - \delta^{18}O_{\text{ref}} = 3.97 \times 10^{-18} \times h^6 - 3.586315 \times 10^{-14} \times h^5 + 1.2843645994 \times 10^{-10} \times h^4 - 2.3087753445094 \times 10^{-7} \times h^3 + 2.1857285350473100 \times 10^{-4} \times h^2 - 0.10603325432255400000 \times h + 23.7; \quad (4)$$

$$\delta D - \delta D_{\text{ref}} = 6.859 \times 10^{-17} \times h^6 - 6.0047709 \times 10^{-13} \times h^5 + 2.0790331349 \times 10^{-9} \times h^4 - 3.61319302207374 \times 10^{-6} \times h^3 + 3.30716141498371 \times 10^{-3} \times h^2 - 1.53651645114701 \times h + 313. \quad (5)$$

These curves are valid only for a given Picarro analyzer and for humidity higher than 70 ppmv and lower than 2000 ppmv. Outside of this calibration range, the extrapolation of the polynomial function may lead to anomalous corrections.

After this correction, the measured values corrected from humidity dependence are corrected using the comparison of the measured values of the two standards at 2000 ppmv to their VSMOW-calibrated values as explained in Sect. 3.5 below.

### 3.4 Accuracy of the system and calibration on the VSMOW–SLAP scale

The accuracy of the system has been addressed performing a two-standard calibration and measuring a third standard treated as an unknown. We used two lab standards calibrated vs. VSMOW on the VSMOW–SLAP scale with large $\delta^{18}O$ and $\delta D$ differences (EPB and FP5) as well as the lab standard NEEM, which was independently calibrated against VSMOW. The three lab standards were vaporized at 800 ppmv and measured by the same L2130-i analyzer.

We used the measured and true values of EPB and FP5 to estimate the $\delta^{18}O$ value of the NEEM standard from its measured value (Table 4). Using the linear relationship obtained from VSMOW-calibrated EPB and FP5 $\delta^{18}O$ vs. measured EPB and FP5 $\delta^{18}O$ values (Fig. S3), following the recommendations of the National Institute of Standards and Tech-

**Table 4.** Comparison of measured vs. VSMOW-calibrated $\delta^{18}O$ values for three standards measured with a Picarro analyzer after the generation of water vapor using the LHLG.

<table>
<thead>
<tr>
<th>Standard</th>
<th>VSMOW-calibrated value</th>
<th>Measured value at 800 ppmv</th>
<th>Measured value humidity corrected from dependence (Eq. 1)</th>
</tr>
</thead>
<tbody>
<tr>
<td>EPB</td>
<td>-6.24 ‰</td>
<td>-8.27 ‰</td>
<td>-10.78 ‰</td>
</tr>
<tr>
<td>NEEM</td>
<td>-33.5 ‰</td>
<td>-34.48 ‰</td>
<td>-36.99 ‰</td>
</tr>
<tr>
<td>FP5</td>
<td>-48.33 ‰</td>
<td>-49.02 ‰</td>
<td>-51.53 ‰</td>
</tr>
</tbody>
</table>
Figure 6. \( \delta^{18}O, \delta D \) and humidity records over December 2018 and the beginning of January 2019. Raw isotopic values are shown in gray. Corrected isotopic values at an hourly resolution are shown in black after the correction of the influence of humidity on the water isotopic ratios and the adjustment of \( \delta^{18}O \) and \( \delta D \) values on the VSMOW–SLAP scale using relationships between measured lab-standard values and known VSMOW-calibrated lab-standard values.

Figure 7. Focus on diurnal variability of \( \delta^{18}O, \delta D \) and humidity recorded at Concordia. Gray curves show the raw measurements, and black curves show the corrected records. The red (blue) bars indicate the calculated uncertainty due to the isotopic ratio vs. humidity dependence (Fig. 5) on the corrected \( \delta^{18}O \) and \( \delta D \) values during periods with maximum (minimum) humidity. The red (blue) rectangles indicate a half-day with maximum (minimum) humidity.

4 Application

The main application of this device is the interpretation of water isotopic profiles at dry sites, in particular in polar regions. As shown in Fig. 5, the influence of humidity on the measurement of the water vapor isotopic composition with the L2130-i analyzer is large when humidity is be-
looking at the diurnal variability in the raw \( \delta \) corrected and corrected data is particularly important in the midity correction which generally shifts the curves to lower \( \delta \) values during a period with a higher humidity level – hence when the isotope ratio–humidity correction is smaller than the signal changes by practically 180° over some periods. Whereas the raw isotope signal peaks during the night, the corrected record shows higher isotope ratios during daytime. The diurnal variability recorded on both raw and corrected isotopic values during a period with a higher humidity level – hence when the isotope ratio–humidity correction is smaller (around day 355 in Fig. 6) – also shows that the \( \delta^{18} \)O (\( \delta \)D) diurnal cycles are indeed in-phase with the humidity cycle. This result confirms the correlation between humidity cycles and \( \delta^{18} \)O and \( \delta \)D of the water vapor at the daily scale at Concordia, as reported by Casado et al. (2016). Thus, we conclude that the anticorrelation of \( \delta^{18} \)O and \( \delta \)D with humidity in the raw data (highlighted in blue in Fig. 7) during periods of low humidity is an artifact due to the influence of the humidity level on the vapor isotopic measurements by the L2130-i analyzer.

5 Conclusions

We have developed an autonomous instrument for low humidity generation (70 to 2400 ppmv) with controlled water vapor isotopic composition that is specifically aimed at carrying out continuous measurements of the water vapor isotopic composition using a laser-based spectrometer in regions characterized by very low humidity, such as polar regions. If needed, an interface permits the user to conveniently connect the new LHLG to the valve sequencer port of commercial Picarro instruments. After more than 1 year of routine operation at two Antarctic sites (Dumont d’Urville and Concordia), this instrument has proven to be very reliable and robust. It consistently generates stable humidity levels with a \( \sigma \) variability lower than 10 ppmv over more than 10 min. Moreover, its performance is significantly better than that of the Picarro SDM at low humidity.

We used this instrument for the calibration of our water isotopic data with a special focus on accurately quantifying the influence of humidity on the measured isotopic composition of the water vapor. This effect is huge at low humidity. We showed that this has an important impact on the interpretation of the diurnal cycles of \( \delta^{18} \)O and \( \delta \)D in the water vapor at Concordia station at humidity below 1500 ppmv. We were able to confirm that, at this site, the diurnal \( \delta^{18} \)O and \( \delta \)D variability is actually correlated with humidity variability, which would not have been possible without the new LHLG instrument.

Finally, the development of such an instrument is an important step forward to a better understanding of the transfer function between climate parameters and the isotopic composition of deep ice cores from the remote East Antarctic plateau, especially in the context of the new program “Beyond EPICA – Oldest Ice”. It should be completed by ongoing development of laser spectrometers better adapted to low humidity levels, such as those based on the technique of optical feedback cavity-enhanced absorption spectroscopy (OFCEAS) (Casado et al., 2016; Landsberg, 2014; Landsberg et al., 2014).

Code availability. The software (HumGen) can be downloaded online from https://zenodo.org/record/4003465#.YHMSmD-xWUn, (Jossoud and Leroy Dos Santos, 2020, last access: 6 April 2021).

Data availability. The data used in this paper are provided in the Supplement.
Supplement. The supplement related to this article is available online at: https://doi.org/10.5194/amt-14-2907-2021-supplement.

Author contributions. CLDS, MC, FP and EK designed and built the instrument. OJ realized the software interface development. CLDS, MC and AL installed the instrument in Antarctica and tested it extensively. EK, SK, MF, AL and EF tested the instrument in the laboratory. AL wrote the paper with input from all co-authors.

Competing interests. The authors declare that they have no conflict of interest.

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