

Response to Anonymous Reviewer #1

Interactive comment on “Real time analysis of insoluble particles in glacial ice using single particle mass spectrometry” by Matthew Osman et al.

Anonymous Referee #1

Received and published: 24 July 2017

Review to "Real time analysis of insoluble particles in glacial ice using single particle mass spectrometry" by Osman et al., AMTD 2017

Dear Anonymous Reviewer #1,

We thank you for your valuable feedback. We have reviewed all your comments/suggestions, and have attempted to address each to the best of our ability below. Please refer to italicized and indented portions for responses.

The manuscript by Osman and coworkers describes the application of a laser ablation single particle mass spectrometer on the analysis of particles in ice core samples. The authors describe the extraction of particles from the samples, the efficiency of particle transfer into the mass spectrometer and also attempt to perform a quantitative measurement of mass concentrations. The manuscript is well written and fits into the scope of AMT. Methods and results are clearly explained and presented. However, I have one major issue that is described below:

Major comments

While I do not have any concerns about the chemical analysis, which has been done by SPMS numerous times for atmospheric particles, I have serious problems with the mass concentration measurement:

C1) - The size dependent extraction efficiency curve was measured using PSL particles, a method that is called "external calibration" (referenced to Wendl et al., which is not an SPMS but an SP2 paper).

Since the SP2 remains the most common single particle, online methodology currently employed in ice core analyses, the experimental framework presented in Wendl et al. (2014) is viewed here to be a plausible surrogate for comparisons to our new SPMS-based single particle approach. Text has now been included in Sect. 2.3.4, making it explicit that the Wendl et al. (2014) "external calibration" approach is based on SP2 methods (Pg. 9 Lines 21-24):

"As defined by Wendl et al. (2014), an "external" calibration approach, as commonly employed in SP2-based measurements of refractory black carbon in glacial snow and ice, assumes that ϵ for a given monodisperse PSL standard scales with an unknown, morphologically/compositionally-heterogeneous and polydisperse, ice core sample."

Additionally, to provide a more direct comparison between SP2 and SPMS techniques, we have now incorporated a new section (3.1.1; Pg. 11-12 Lines 25-27, 1-23) entitled "Comparison of SPMS to SP2-based single particle methods in snow and ice".

C2) - The measured efficiencies are very low (highest values for 657 nm PSL particles are around 0.40%). This results in a very large correction factor needed to derive the mass concentration in

the sample.

While we acknowledge this potential fallback, we point out that the relatively low extraction efficiency is largely a result of differences inherent to SP2 vs. PALMS transmission and measurement capabilities, which have now been better contrasted/clarified in Section 3.1.1 (Pg. 12 Lines 12-22):

“More recent studies have achieved high nebulization efficiencies (~50%) up to ~2 μm using the CETAC Marin-5 pneumatic nebulizer (Teledyne CETAC Technologies, Omaha, NE, USA; Mori et al., 2016; Katich et al., 2017). In this context, however, it is noted that achieving too high of an extraction efficiency could be disadvantageous for SPMS, should the number of particles reaching the SPMS inlet exceed that instrument’s max transmission rate (~10 particles sec⁻¹ for PALMS; Cziczo et al., 2006) where the limit is data writing and laser repetition rate. This does not affect SP2, which can more rapidly measure the incandescence of carbonaceous material passing through a continuous laser (i.e., 2-3 orders of magnitude higher), though with the cost of i) not delivering information on internal mixing state or ii) aerodynamic size (as opposed to black-carbon volume equivalent diameter). More efficient nebulization at relevant SPMS sizes, coupled to more rapid excimer lasers and data writing, would increase the data acquisition rate.”

C3) - The efficiency is assumed to be independent of shape and composition. This is certainly an oversimplification.

We agree with the reviewer that this is indeed an important assumption underlying our methodology. We have now included text in 2.3.2 (Pg. 9 Lines 23-24) acknowledging the assumption more explicitly:

“... assumes that ε for a given monodisperse PSL standard scales with an unknown, morphologically and compositionally-heterogeneous and polydisperse, ice core sample.”

We have additionally included the following text at the end of Sect. 3.2.4 encouraging future studies to explore these assumptions in greater detail (Pg. 17-18 Lines 25-29, 1-7):

“While our results show potential exists for using SPMS to determine insoluble mass concentrations of particles in snow and ice, they also identify areas where more work is needed before SPMS can be used as a quantitative tool. These include: i) executing multiple extraction efficiency (eq. 1) calculations as a function of particle class (in addition to size), (...)”

C4) - Drifts in nebulizer efficiency have not been considered.

Since only one sample (DS14-05) was measured for quantitative purposes, transmission of the 8.8e6 particles/cc solution was not tested between sample(s). We did, however, test whether systematic trends in nebulization drift could occur over the hour-long measurement period. This test was done by directing a particle-laden airflow (nebulized from the monodisperse, 746 nm PSL liquid standard: $m_{\text{PSL}}(D_p = 746 \text{ nm}) = 8.8 \times 10^6 \text{ PSL particles cm}^{-3}$; Sect. 2.3.3), to an optical particle sizer (OPS; MesaLabs Bios DryCal 220), and performing continuous, one-second interval measurements over three separate ~1-hour long tests (i.e., the longest sample integration period in the manuscript). The nebulization efficiency (ϵ_{neb}) was calculated in this test as,

$$\epsilon_{\text{neb}}(D_p = 746 \text{ nm}) = \frac{n_{\text{OPS}}(D_p = 746 \text{ nm}) \cdot F_{\text{flow}}}{m_{\text{PSL}}(D_p = 746 \text{ nm}) \cdot V_{\text{neb}}}$$

where n_{OPS} is the PSL number concentration measured by the OPS, and the flow rates F_{neb} , V_{neb} and, F_{wet} are as described in the main text. The long-term drifts in nebulization, calculated as the linear percent change over the hour-long measurement interval, were determined in the three tests to be 22%, 9.2%, and -33 % ($\Delta\epsilon_{\text{neb}}/\Delta t = 0.18 \cdot 10^{-5} \text{ s}^{-1}$, $0.08 \cdot 10^{-5} \text{ s}^{-1}$, and $-0.30 \cdot 10^{-5} \text{ s}^{-1}$, respectively). Importantly, results of the three tests indicated that long-term drift direction was not systematic, as both negative and positive drift biases occurred over the one-hour nebulization periods (Fig. R1, shown below). It is thus reasonable to view the drift uncertainty as a simple spread about the hour-long mean of the three tests, in this case equating to $\epsilon_{\text{neb}} = 0.068 \pm 0.013$ (1 s.d.), or ~18% relative uncertainty.

It is equally important to note that in our study, calculation of particle mass-concentration (eq. 4) does not explicitly incorporate estimates of ϵ_{neb} , but rather estimates of the extraction efficiency, ϵ (eq. 5),

determined experimentally and independent of ϵ_{neb} . However, via eq. 6 (now included in the main text; see C5 below), ϵ is shown to be a function of ϵ_{neb} and transmission efficiency, ϵ_{trans} . Since past studies (e.g., Cziczo et al., 2006) have illustrated that PALMS transmission is relatively stable, we thus take the uncertainty interval calculated for ϵ (~30% relative uncertainty at $\epsilon(D_p = 746 \text{ nm})$; eq. 5) to implicitly encapsulate uncertainties in nebulization efficiency.

The above information has been included as supplementary material, and the following sentence added to Sect. 3.2.4 (Pg. 16 Lines 23-26):

“Note that while no systematic trends in nebulization drift were found over either hour-long measurement period, short term fluctuations in nebulization could occur; for the present experiment, such fluctuations are assumed to be encapsulated as uncertainty about the extraction efficiency parameterization (eq. 5; see Supplementary Material for details).”

Overall, we agree with the reviewer that future mass-concentration applications using SPMS – especially those where multiple successive samples are measured for mass concentration – should implement regular standardized checks for performance drift between samples. Text has also been added to 3.2.4 making this latter suggestion explicit (Pg. 17-18 Lines 25-29, 1-7; refer to C2 above).

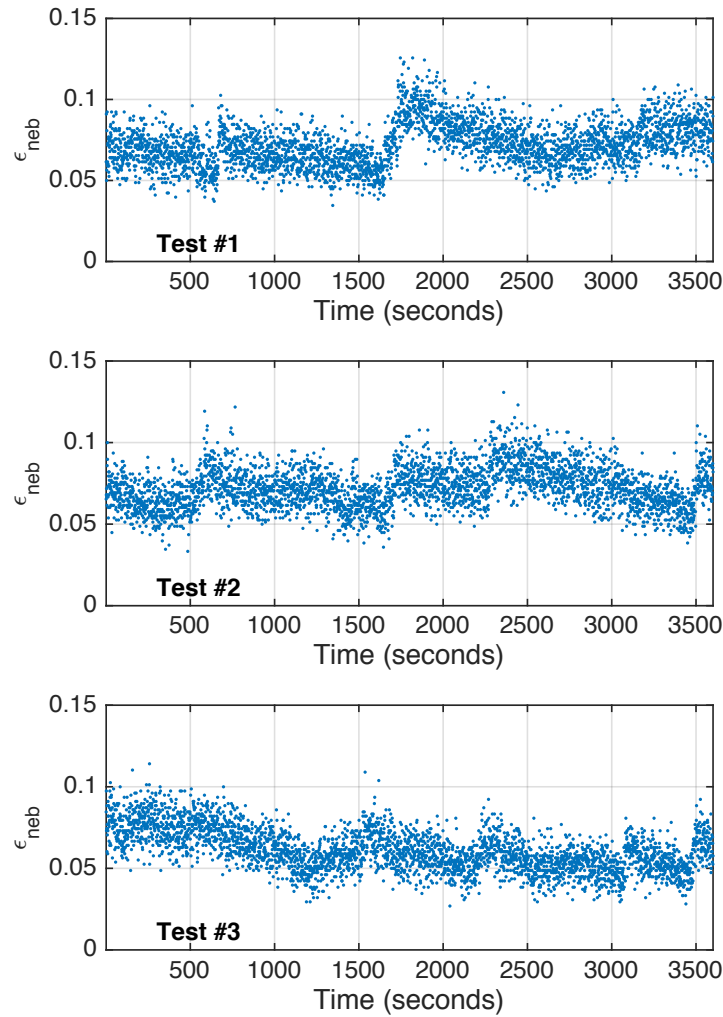


Fig. R1. Results of three separate nebulization drift tests.

C5) During the PSL calibration experiments, the authors used an OPS to monitor particle concentration and size to measure the transmission efficiency into the PALMS (which was found to be between 1 and – 16%). Why has this (or another method to measure size and number) not been done during the ice core sample analysis? By such, the transmission efficiency into the PALMS would have been measured using the real sample particles. Non-spherical shapes of the insoluble particles can certainly influence the transmission efficiency. Furthermore, methods exist to measure particle concentration in liquids. A comparison of the derived particle concentrations from the SPMS with such a reference measurement would have helped in validating the results. At least a determination of the mass concentration by filtering the solution and weighing the filter would have been possible, although here larger particle may dominate the total mass concentration. Without any comparison to an independent measurement of the same sample, the quantification of the results seems not reliable and overstretches the capabilities of an SPMS.

We thank the reviewer for his/her suggestions, each of which is valid. We note that PALMS transmission capabilities are well-characterized, as validated by this and past studies (details found in Cziczko et al. (2006; 2013), thus the largest uncertainty likely arises from nebulization effects, as addressed above. More generally, as to the reviewer's primary concerns concerning mass concentration measurements, we iterate that this study's primary goal was not to "oversell" the use of SPMS for conducting mass concentration measurements of particles in snow and ice, but rather was in attempting to illustrate the feasibility of – including highlighting current fallbacks inherent in and the potential for future improvements to – the method. We hope that future studies may improve upon this baseline study by implementing comparisons to alternate well-founded techniques for measuring particle concentration in liquid (e.g., Coulter Counter techniques, UHSAS). These sentiments and suggestions have now been noted at the end of 3.2.4 (Pg. 17-18 Lines 26-29, 1-8):

"While our results show potential exists for using SPMS to determine insoluble mass concentrations of particles in snow and ice, they also identify areas where more work is needed before SPMS can be used as a quantitative tool. These include:

(...)

iv) comparing SPMS-derived particle concentrations with results from alternate, well-founded high-precision instrumentation (e.g., an Ultra-High Sensitivity Aerosol Spectrometer (UHSAS; Droplet Measurement Technologies Inc., Boulder, CO), or Coulter Counter instrumentation)."

Minor comments:

Page 11, lines 1-3: How was determined that all water was evaporated?

In general, PALMS will not provide spectra for particles that are water-saturated (i.e., "quenched"). However, for low-to-moderate degrees of water-saturation, PALMS can distinguish between wet and dry particles via water-cluster peaks ($m/z = 18$) that are prominent in the particle spectra (Murphy and Thompson, 1995). Determination of adequate water-evaporation could thus be experimentally determined by increasing the dry-to-wet flow ratio until evidence of water-saturation was eliminated in the particle spectra. The following text was added to Sect. 2.3 (Pg. 's 6-7 Lines 26, 1-2) clarifying this point:

"As PALMS generally does not provide spectra for water-saturated particles (Cziczko et al., 2006), the atomized particles were then adjoined with the dry flow, dropping the relative humidity and evaporating residual condensed water, resulting in dry residual particles entering the PALMS inlet."

Figures of mass spectra in appendix: What is the method to select which mass peaks are labeled. Apparently not always the largest peaks? Are peaks unknown/not identified, e.g. in Fig. A1, lowest panel, around m/z 90? Additionally, minor tick marks and/or grid lines help the reader to determine the m/z number of a peak in the mass spectrum.

In general, all major peaks corresponding to known ionic fragments are labeled. Specific details for this identification process can be found in prior studies, particularly references noted in Sect. 2.1 of the main text (e.g., Murphy and Thompson, 1997a,b; Cziczko et al., 2013). Many of the large peaks not

labeled correspond to organic particles, or, in this study's case, particles that have undergone severe post-aqueous processing. There presently remains ambiguity in organic fragment identification, as is the case for Fig. A1 for $m/z \sim 90$. The following text has been added (Pg. 20, Lines 15-17):

“All major peaks corresponding to known ionic fragments have been labeled (see Murphy and Thompson, 1997 a, b; and Cziczo et al., 2013).”

Grid lines and x-axis minor ticks have now been added to the (now Appendix 2's) mass-spectra plots.

References:

- Cziczo, D. J., Froyd, K. D., Hoose, C., Jensen, E. J., Diao, M., Zondlo, M. A., Smith, J. B., Twohy, C. H. and Murphy, D. M.: Clarifying the Dominant Sources and Mechanisms of Cirrus Cloud Formation, *Science*, 340, 1320–1324, 2013.
- Cziczo, D. J., Thomson, D. S., Thompson, T. L., DeMott, P. J., Murphy, D. M.: Particle analysis by laser mass spectrometry (PALMS) studies of ice nuclei and other low number density particles, *Int. J. Mass Spectrometry*, 258, 21-29. 2006.
- Murphy, D. M., and Thomson, D. S.: Chemical composition of single aerosol particles at Idaho Hill: Positive ion measurements, *J. Geophys. Res.*, 102, 6353-6368, doi:10.1029/96JD00858, 1997a.
- Murphy, D. M., and Thomson, D. S.: Chemical composition of single aerosol particles at Idaho Hill: Negative ion measurements, *J. Geophys. Res.*, 102, 6353-6368. doi:10.1029/96JD00859, 1997b.
- Murphy, D. M., and Thomson D. S.: Laser Ionization Mass Spectroscopy of Single Aerosol Particles, *Aerosol Science and Technology*, 22(3), 237-249, doi:10.1080/02786829408959743, 1995.

Response to Anonymous Reviewer #2

Interactive comment on “Real time analysis of insoluble particles in glacial ice using single particle mass spectrometry” by Matthew Osman et al.

Anonymous Referee #2

Received and published: 19 July 2017

Dear Anonymous Reviewer #2,

We thank you for your valuable feedback. We have reviewed all your comments/suggestions, and have attempted to address each to the best of our ability below. Please refer to italicized and indented portions for responses.

Referee Comments for Osman, M. et al., “Real time analysis of insoluble particles in glacial ice using single particle mass spectrometry”

General Comments

The authors implement a PALMS to successfully employ nebulizer+instrument techniques previously used by single particle soot photometers to make measurements of particle classification, size and concentration in ice core samples from Greenland. Though limited by a low particle transmission that is currently inherent to the PALMS instrument, the methodology still manages to result in new, interesting measurements that to my knowledge have not been realized before, making this a very worthy manuscript for publication after addressing some relatively minor concerns.

The results also provide substantial motivation to continue to push the transmission efficiency capabilities of the PALMS instrument, the results of which would make the methods used here a much more viable path towards measurement of particle concentrations and size distributions in ice cores. The presentation of the goals, setup, methodology and results are generally very clearly stated, with very few exceptions (notes below). Further, the manuscript seems to have been carefully prepared, as I struggled to find any typos, spelling errors or poor grammar. Below, I suggest some minor corrections to be addressed, including explanation of their monitoring of potential background contamination levels and system stability. Also, I suspect there may be an error in their calculation of nebulization efficiency (though it results in only a small change in the quoted number). Finally, I recommend more carefully explaining the differences in interpreting PALMS vs SP2 measurements of black carbon / soot.

Specific Comments

C1) Page 3, line 18: Also see Katich et al., 2017 (doi:10.1080/02786826.2017.1280597), which provides a lengthy closely-related discussion on aerosolizing particulate from snow and ice.

We thank you for pointing us to this recent study + discussion therein, which had been previously unknown to the authors; the reference for Katich et al., 2017 has now been included (Pg. 3 Line 19).

C2) Page 7, line 5,6: Did the authors intersperse regular measurements of ‘blanks’ (i.e. ultra-pure water) to quantify the average background level of particulate seen by PALMS when using a

‘clean’ nebulization system? There is mention of looking for background from the stainless steel band saw (not what I’m concerned about here) and of sonicating the parts between samples. But I wonder if there is a quantification of average background levels due to any residual particulate in the nebulizer lines? How does this compare to signal levels? Negligible?

We did not systematically implement measurements of blanks between samples as the reviewer describes here. Our methodology was to flush the flow line continuously with an inert N₂ gas-flow (for ~15 min) between samples to remove residual particulates prior to the next measurement. To answer this point we now state the following in Sect. 2.3 (Pg. 7 Lines 8-9):

“Between runs, the nebulizer and sampling beaker were cleaned and sonicated for 15 minutes using ultrapure (Milli-Q; 18.2 MΩ) water, and the flow-line flushed continuously with the inert carrier gas.”

We do acknowledge the utility of the reviewer’s suggestion to regularly implement blanks between measurements and have also included the following in 3.2.4 (Pg. 17-18 Lines 25-29, 1-7):

“While our results show potential exists for using SPMS to determine insoluble mass concentrations of particles in snow and ice, they also identify areas where more work is needed before SPMS can be used as a quantitative tool. These include: i) executing multiple extraction efficiency (eq. 1) calculations as a function of particle class (in addition to size), ii) incorporating regularized tests for drifts in SPMS extraction efficiency and employing “blank” tests between sample measurements in order to improve delineation to changes in background particulate levels, iii) achieving a greater number of particle measurements (either through improvements in particle extraction/PALMS transmission or longer sample integration times), and iv) comparing SPMS-derived particle concentrations with results from alternate, well-founded high-precision instrumentation (e.g., an Ultra-High Sensitivity Aerosol Spectrometer (UHSAS; Droplet Measurement Technologies Inc., Boulder, CO), or Coulter Counter instrumentation).”

C3) Page 8, line 19: Please clarify the phrase “rate of liquid nebulization”... same as rate at which liquid is fed to the nebulizer i.e. liquid uptake rate? Does this occur at a user controlled pump rate, or is it self-aspirating? If you have control over the pump rate, this could be another way to tweak the rate of particulate delivered to the PALMS inlet.

The “rate of liquid nebulization” is the average volumetric loss rate of the liquid sample during nebulization of the sample. This rate is indirectly user-controlled by setting the “wet” gas flow rate to the nebulizer (i.e., F_{wet}). However, while increasing the wet flow rate to the nebulizer could increase the rate of particulate delivered to the PALMS inlet, it was determined that water saturation (i.e., quenching) of the particles became problematic at much higher rates than that quoted in the manuscript (2 lpm). The following text has been added to Sect. 2.3.2 (Pg. 8, lines 23-24) to clarify:

“...V_{neb} is the rate of liquid nebulization (i.e., a prescribed volumetric loss rate of the sample, determined here using a scale; 4.4·10⁻⁶ ± 1.6·10⁻⁶ mL sec⁻¹)”

C4) Page 9, line 23: Agreed, the nebulization efficiency can drift over time, even substantially, depending on the solution being nebulized. Was this monitored by occasionally measuring transmission of the 8.8e6 particles/cc solution in between samples? If so, perhaps show a summary of nebulization stability in supplemental material?

Since only one sample (DSI4-05) was measured for quantitative purposes, transmission of the 8.8e6 particles/cc solution was not tested between sample(s). We did, however, test whether systematic trends in nebulization drift could occur over the hour-long measurement period. This test was done by directing a particle-laden airflow (nebulized from the monodisperse, 746 nm PSL liquid standard: m_{PSL}(D_p = 746 nm) = 8.8x10⁶ PSL particles cm⁻³; Sect. 2.3.3), to an optical particle sizer (OPS; MesaLabs Bios DryCal 220), and performing continuous, one-second interval measurements over three separate ~1-hour long tests (i.e., the longest sample integration period in the manuscript). The nebulization efficiency (ε_{neb}) was calculated in this test as,

$$\epsilon_{neb}(D_p = 746 \text{ nm}) = \frac{n_{OPS}(D_p=746 \text{ nm}) \cdot F_{flow}}{m_{PSL}(D_p=746 \text{ nm}) \cdot V_{neb}}$$

where n_{OPS} is the PSL number concentration measured by the OPS, and the flow rates F_{neb} , V_{neb} and F_{wet} are as described in the main text. The long-term drifts in nebulization, calculated as the linear percent change over the hour-long measurement interval, were determined in the three tests to be 22%, 9.2%, and -33 % ($\Delta\epsilon_{neb}/\Delta t = 0.18 \cdot 10^{-5} \text{ s}^{-1}$, $0.08 \cdot 10^{-5} \text{ s}^{-1}$, and $-0.30 \cdot 10^{-5} \text{ s}^{-1}$, respectively). Importantly, results of the three tests indicated that drift direction was not systematic, as both negative and positive drift biases occurred over the one-hour nebulization periods (Fig. R1, shown below). It is thus reasonable to view the drift uncertainty as a simple spread about the hour-long mean of the three tests, in this case equating to $\epsilon_{neb} = 0.068 \pm 0.013$ (1 s.d.), or ~18% relative uncertainty.

It is equally important to note that in our study, calculation of particle mass-concentration (eq. 4) does not explicitly incorporate estimates of ϵ_{neb} , but rather estimates of the extraction efficiency, ϵ (eq. 5), determined experimentally and independent of ϵ_{neb} . However, via eq. 6 (now included in the main text; see C5 below), ϵ is shown to be a function of ϵ_{neb} and transmission efficiency, ϵ_{trans} . Since past studies (e.g., Cziczo et al., 2006) have illustrated that PALMS transmission is relatively stable, we thus take the uncertainty interval calculated for ϵ (~30% relative uncertainty at $\epsilon(D_p = 746 \text{ nm})$; eq. 5) to implicitly encapsulate uncertainties in nebulization efficiency.

The above information has been included as supplementary material, and the following sentence added to Sect. 3.2.4 (Pg. 16 Lines 23-26):

“Note that while no systematic trends in nebulization drift were found over either hour-long measurement period, short term fluctuations in nebulization could occur; for the present experiment, such fluctuations are assumed to be encapsulated as uncertainty about the extraction efficiency parameterization (eq. 5; see Supplementary Material for details).”

Overall, we agree with the reviewer that future mass-concentration applications using SPMS – especially those where multiple successive samples are measured for mass concentration – should implement regular standardized checks for performance drift between samples. Text has also been added to 3.2.4 making this latter suggestion explicit (Pg. 17-18 Lines 25-29, 1-7; refer to C2 response).

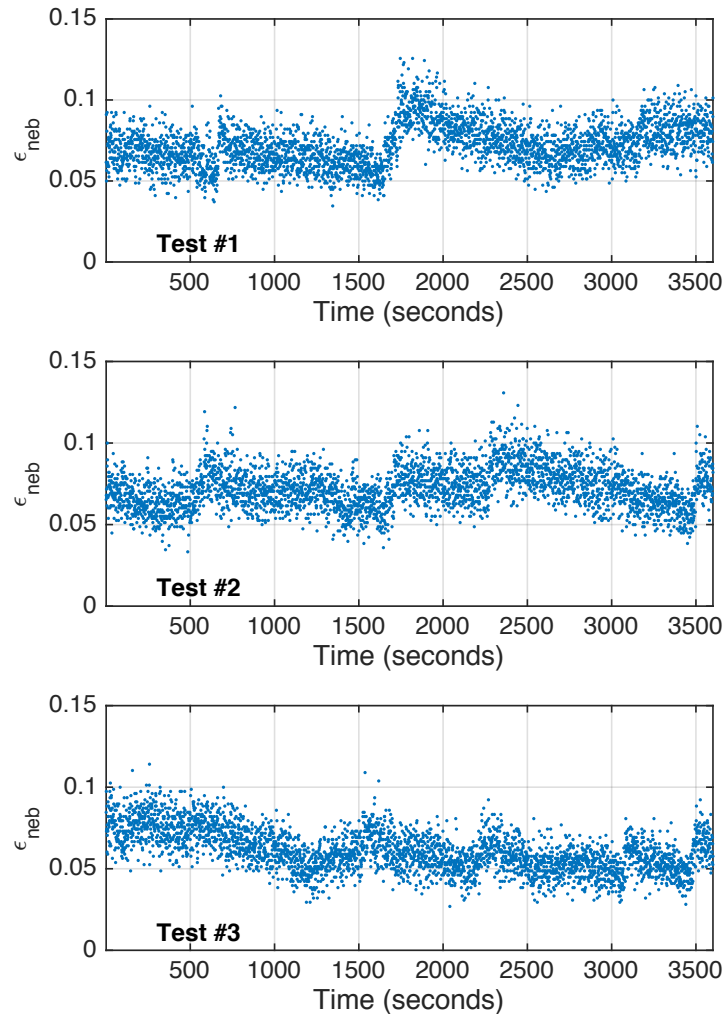


Fig. R1. Results of three separate nebulization drift tests.

C5) Page 11, line 6: The statement, “Scaling the efficiency curve by the ratio of excess-flow to the PALMS inlet flow”, I believe should read “Scaling the efficiency curve by the ratio of TOTAL-flow to the PALMS inlet flow.” Further, I don’t think I agree with the calculation of the nebulization efficiency, where it is achieved simply by making the correction due to particle loss from low PALMS sample flow. I would argue the following: If a nebulizer’s efficiency is defined as the ratio of ‘the rate of particles emerging in aerosol from the nebulizer’ (call it $R_{aerosol}$ = Naerosol/sec) to ‘the rate of particles introduced to the nebulizer’ (call it $R_{introduced}$, which is known from your known PSL concentration and liquid uptake rate), then to know Naerosol, you have to work backwards from the number of particles that PALMS sees.

****THIS DID NOT COPY / PASTE PROPERLY. PLEASE SEE ATTACHED PDF FOR EQUATIONS****

For convenience, we have copy and pasted the aforementioned attached .pdf below:

Page 11, line 6: The statement, “Scaling the efficiency curve by the ratio of excess-flow to the PALMS inlet flow”, I believe should read “Scaling the efficiency curve by the ratio of TOTAL-flow to the PALMS inlet flow.”

Further, I don't think I agree with the calculation of the nebulization efficiency, where it is achieved simply by making the correction due to particle loss from low PALMS sample flow. I would argue the following: If a nebulizer's efficiency is defined as the ratio of 'the rate of particles emerging in aerosol from the nebulizer' (call it $R_{aerosol} = N_{aerosol}/sec$) to 'the rate of particles introduced to the nebulizer' (call it $R_{introduced}$, which is known from your known PSL concentration and liquid uptake rate), then to know $N_{aerosol}$, you have to work backwards from the number of particles that PALMS sees.

$$\epsilon_{nebulization} = \frac{R_{aerosol}}{R_{introduced}} = \frac{N_{aerosol}/sec}{R_{introduced}} = \frac{N_{aerosol}/sec}{m_{psl} \cdot V_{neb}}$$

working backward from the number of particles need by PALMS...

$$(N_{aerosol}/sec) = \underbrace{\left(\frac{N_{particles\ detected\ by\ PALMS}}{sec} \right)}_{(1)} \cdot \underbrace{\left(\frac{N_{particles\ introduced\ to\ PALMS}}{N_{particles\ detected\ by\ PALMS}} \right)}_{(2)} \cdot \underbrace{\left(\frac{F_{wet} + F_{dry}}{F_{inlet}} \right)}_{(3)} \underbrace{\left(\frac{F_{flow}}{F_{dry}} \right)}_{(4)}$$

where

(1) = f_{PALMS}

(2) = ϵ_{trans}

(3) = the correction introduced in the text

(4) = needed to correct for the fact that only 2/5 of the flow into PALMS has gone through the nebulizer (i.e. F_{dry} is just dilution air)

thus...

$$\epsilon_{neb} = \epsilon_{PSL} \cdot \frac{1}{\epsilon_{trans}} \cdot \left(\frac{F_{wet} + F_{dry}}{F_{dry}} \right) = \frac{\epsilon_{PSL}}{\epsilon_{trans}} \cdot \left(\frac{F_{flow}}{F_{dry}} \right)$$

Plugging in some maximal numbers from figure 2, I get:

$$\epsilon_{neb(max)} = \left(\frac{0.004}{0.15} \right) \cdot \left(\frac{5}{2} \right) \cong 6.7\%$$

it's not far off the quoted 4%, but as a matter of correctness, should be changed (assuming I've not erred).

We thank the reviewer for sharing this concern, which was well articulated by the equations provided in his/her supplement. We agree with the reviewer's determination of the nebulization efficiency (void the incorporation of F_{dry} , which we believe should in fact be F_{wet}). However, we believe the reviewer's primary underlying concern (as it relates to miscalculation of the nebulization efficiency) was due primarily to a nomenclature error: we incorrectly stated “nebulization efficiency” as opposed to “extraction efficiency” in Sect 3.1. While 1) nebulization efficiency (i.e., $R_{aerosol}/R_{introduced}$, as defined by the reviewer) and 2) transmission efficiency can be viewed as two independent properties of the experimental set-up, both quantities are effectively encapsulated by 3) extraction efficiency (i.e., as per the reviewer $e_{psl} = e_{neb} \cdot e_{trans} \cdot [F_{wet}/F_{flow}]$); in this context, we believe our more-simplistic scaling correction remains valid.

To avoid nomenclature confusion and to render our SPMS results more directly comparable to past studies (e.g., Schwarz et al., 2012, Ohata et al., 2013, Wendl et al., 2013, Katich et al., 2017), our simplistic scaling has been removed, and a revised discussion now including nebulization efficiency has been added to Sect. 3.1 (Pg. 11 Lines 18-24), including the addition of a similar derivation to that

of the reviewer's to Appendix 1 (Pg., 19-20, Lines, 22-27, 1-10):

"A1. Calculating nebulization efficiency

Here, we derive the determination of nebulization efficiency (ϵ_{neb} ; Sect. 3.1). We define ϵ_{neb} as the flow-weighted ratio of the rate of successfully nebulized particles per unit time relative to the (liquid) number concentration of particles introduced to the nebulizer, such that

$$\epsilon_{neb} = \frac{f_{neb}(D_p)}{m_{PSL}(D_p) \cdot F_{neb}} \quad (A1)$$

where $m_{PSL}(D_p)$ and F_{neb} are as defined in eq. (1), and $f_{neb}(D_p)$ is the frequency of particles successfully nebulized (e.g., particles sec^{-1}) as a function of PSL diameter (D_p). In this case, $f_{neb}(D_p)$ is the quantity that must be solved for. We take

$$f_{neb}(D_p) = \frac{n_{PALMS}(D_p) \cdot F_{flow}}{\epsilon_{trans}(D_p)} \cdot \left[\frac{F_{flow}}{F_{wet}} \right] \quad (A2)$$

such that the scalar quantity $\left[\frac{F_{flow}}{F_{wet}} \right]$ acts as a correction for the flow balance of particles actually passing through the nebulizer (Figure 1) and $\epsilon_{trans}(D_p)$ corrects for the size-dependent particle transmission of PALMS. Note that $n_{PALMS}(D_p)$ and F_{flow} are as previously defined in eq. (1). Plugging eq. (A2) into (A1), and via relation to eq. (1),

$$\epsilon_{neb} = \frac{\epsilon(D_p)}{\epsilon_{trans}(D_p)} \cdot \left[\frac{F_{flow}}{F_{wet}} \right] \quad (A3)$$

as defined in eq. (6)."

C6) Page 16, line 6: Regarding the 0.8 g/cm³ density used for soot... What density are you referring to? Void free rBC? All recent literature that I know of uses either 2.0 g/cm³ (a bit of an old number) or 1.8 g/cm³, so maybe this is a typo, or maybe you have actually used the wrong number, or maybe you are referring to a density that isn't clarified. Please comment...

Assuming general compositional similarity between BC and soot, we used an effective soot density value of 0.8 g/cm³ following the experimental work of Moteki and Kondo (2010) and Kiselev et al. (2010), and as implemented by Schwarz et al. (2012). We have therefore retained 0.8 g/cm³, though have clarified that this is indeed an effective density value (Pg. 16, line 18-19).

C7) Figure 4: It is important to note somewhere that the soot size distributions here are not directly comparable to typical rBC size distributions shown in literature that are measured via incandescence, i.e. SP2 measurements. My understanding is that PALMS will only measure the size of the entire soot-containing particle (via a scattered-light signal), which includes any 'coating' that is combined with the BC particle, and is not a measurement of the 'core' refractory BC mass. On the other hand, SP2 measurements will separate the rBC core mass (or volume-equivalent-diameter, VED) from the coating associated with an individual rBC particle. This rBC core VED distribution is what is typically shown in literature. So one could not compare the soot distributions shown here to, say, Schwarz et al. 2013 ("Black Carbon Aerosol Size in Snow"). A slightly expanded discussion on the interpretation of SP2 vs PALMS measurements of soot/rBC is recommended.

We thank the reviewer for this valuable suggestion, and the following discussion was included at the end of Sect. 3.2.4 (Pg. 17-18 Lines 13-24):

"There are notable inherent differences between SPMS- and SP2-derived soot size distribution determinations, however. Namely, whereas SP2 can differentiate the volume-equivalent diameter of refractory soot-components in compositionally-heterogeneous particles (Schwarz et al., 2012), our SPMS approach presumes a particle to be comprised wholly of soot if its mass-spectrum is classified as such. Thus, depending upon the morphology, internal mixing state, and ionization potential of the analyzed soot particles, SPMS may be subject to positive size distribution biases (Fig. 4)."

Technical Corrections

Page 3, line 15: ‘Schwartz’ should be Schwarz.

Typo has now been corrected (Pg. 3, line 15).

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Real time analysis of insoluble particles in glacial ice using single particle mass spectrometry

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Abstract. Insoluble aerosol particles trapped in glacial ice provide insight into past climates, but analysis requires information on climatically-relevant particle properties, such as size, abundance, and internal mixing. We present a new analytical method using a time-of-flight single particle mass spectrometer (SPMS) to determine the composition and size of insoluble particles in glacial ice over an aerodynamic size range of $\sim 0.2 - 3.0 \mu\text{m}$ diameter. Using samples from two Greenland ice cores, we developed a procedure to nebulize insoluble particles suspended in melted ice, evaporate condensed liquid from those particles, and transport them to the SPMS for analysis. We further determined size-dependent extraction and instrument transmission efficiencies to investigate [the feasibility of determining](#) particle-class specific mass concentrations. We find SPMS can be used to provide constraints on the aerodynamic size, composition, and relative abundance of most insoluble particulate classes in ice core samples. We describe the importance of post-aqueous processing to particles, a process which occurs due to nebulization of aerosols from an aqueous suspension of originally soluble and insoluble aerosol components. This study represents an initial attempt to use SPMS as an emerging technique for the study of insoluble particulates in ice cores.

1. Introduction

Aerosol particles play a significant role in global climate, both directly, through regulation of regional and global radiation budgets and atmospheric photochemical cycles, and indirectly, through their ability to modulate cloud microphysical and precipitation processes. In both cases, the abundance,

size, morphology, and composition of the aerosols dictate their ability to affect climate (Seinfeld and Pandis 2006). As such, the ideal atmospheric particle measurement has been described as one that is able to “count each particle and report its size, chemical composition, and morphology rapidly” (Wexler and Johnston, 2011).

5 Inferences of past climates have been achieved using ice core records, whose chronologies provide direct and high-resolution records of past atmospheric composition (Legrand and Mayewski, 1997). A large number of methodologies have been developed to determine properties of aerosols in snow and ice. However, all have important limitations.

10 So-called “offline” particle retrieval methods, requiring a precursor step to first concentrate particles prior to analysis, have been widely employed for ice core studies. Offline studies typically use a pre-existing analytical instrument and are therefore of low expense, requiring only that the sample be transported to the instrument. When paired with quantitative analytical techniques, such as Raman (Sakurai et al., 2011) and sublimation energy dispersive (SED) X-ray spectroscopy (Drab et al. 2002, Iizuka et al., 2009, Oyabu et al. 2015), as well as optical techniques, such as transmission and scanning
15 electron microscopy (Murr et al. 2004; Ellis et al. 2015), offline methods can achieve determinations of particle size, composition, and morphology. There are limitations of offline techniques, however, including the time and resources required for sample recovery and transport, and these limitations often render such methods insufficient for acquiring contiguous, high-resolution particle records from ice
20 cores. Further, offline methods can increase the chance of sample contamination and/or particle losses occurring during handling, transport, and storage (Ohata et al., 2011).

“Online” instrumentation, in contrast, allows for rapid, real time delineations of ice core
particulate properties, necessary for high-resolution measurements of particle mass-concentrations and
(or) size-distributions along an ice core. For example, Coulter Counter (CC) or laser particle detection
(LPD) based instrumentation is typically used for determining insoluble particle (“dust”) size
25 distributions in glacial ice (Hamilton and Langway, 1967; Knipperz and Stuut, 2014). However, neither CC nor LPD techniques provide associated information on particle composition or morphology; broad assumptions of both parameters are thus required to infer ice core dust concentrations from these instruments. Alternate online techniques, such as inductively coupled plasma mass spectrometry (ICP-

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MS) and ion chromatography (IC), are also routinely used to determine elemental (e.g., Cd, Ce, Zn, Pb) and soluble ion (e.g., Na^+ , SO_4^{2-} , NH_4^+) concentrations in ice cores, providing independent indices of past dustiness (e.g., Rhodes et al. 2011), and anthropogenic particle emissions (e.g., Osterberg et al. 2008, McConnell et al. 2008). Albeit analytically precise, ICP-MS and IC rely on techniques that
5 analyze only a portion of aerosols in an aqueous suspension/solution, and not the properties of the progenitor aerosols themselves, i.e., particulate morphology or size.

The recent advent of a commercially available intracavity laser-induced incandescence photometer, the Single Particle Soot Photometer (SP2; Droplet Measurement Technologies, Inc. Boulder, CO) has allowed for a new class of online measurements where both concentrations and size
10 distributions of (refractory) carbonaceous particles in snow and ice can be recovered on a per-particle basis (McConnell et al. 2007; Kaspari et al. 2011; Ginot et al. 2014; Sun et al. 2014). McConnell et al. (2007) were the first to measure past black carbon deposition continuously along a Greenlandic ice core using SP2 instrumentation. More recent SP2 studies have highlighted the necessity of quantifying instrumental errors and systematic uncertainties when extracting trace amounts of highly heterogeneous
15 material from snow and ice ([Schwarz et al. 2012](#); [Ohata et al. 2013](#); [Wendl et al. 2014](#)). Specifically, these studies have shown that correction factors, primarily a result of the efficacy achieved in nebulizing insoluble particles from a melted ice core matrix and aerodynamically delivering them to the SP2 for analysis, can be large and vary significantly between laboratory set-ups ([Lack et al., 2014](#), [Katich et al., 2017](#)).

To date no single online method has proven capable of determining the size and chemical composition for all particle-types situated in glacial ice. Here we expand upon the advantages of the SP2-based methodology by using a time-of-flight (ToF) single particle mass spectrometer (SPMS), allowing for analyses of small aerosol masses (~fg) and differentiation of internally mixed components – from both refractory (e.g. black carbon, mineral dust) to volatile (e.g. sulfuric acid, organic
25 compounds) – on a particle-by-particle basis. This technique leverages a state of the art instrument used for atmospheric studies (Murphy, 2005; Cziczo et al. 2006) to provide real-time (i.e., online) measurements, thereby minimizing contamination, losses during handling, and sample preparation requirements. We describe the development and application of the SPMS method to resolve the size,

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abundance, and composition of a host of insoluble particle-types including, but not limited to, mineral/metallic, biomass burning, soot and organic particles contained within ice cores. [Using ice core samples from two sites in west-central Greenland we test the feasibility of SPMS as a new online tool for particulate measurements in glacial ice, and evaluate the qualitative and quantitative utility of this method.](#) We conclude with suggestions for future application and improvements to the method.

2. Methodology

2.1 Particle Analysis by Laser Mass Spectrometry (PALMS) and particle classification

The PALMS instrument has previously been used for *in situ* measurements of airborne particles in both laboratory settings, as well as airborne and ground-based field campaigns. The PALMS instrument has been described previously (Murphy and Thomson, 1995; Thomson et al., 2000; Cziczko et al., 2006, Cziczko et al., 2013). In brief, accumulation- to coarse-mode ($\sim 0.2 - 3.0 \mu\text{m}$ aerodynamic diameter) particles entering the PALMS inlet are brought into a vacuum and passed through two continuous Nd-YAG lasers causing a scattering of green-wavelength light ($\lambda \approx 532 \text{ nm}$). The time difference between (and the magnitude of) scatter signals provide a measure of the particle diameter (aerodynamic and geometric, respectively). The scattering additionally triggers a [193 nm](#) excimer laser pulse ($\sim 10 \text{ ns}$ pulse duration; 10^9 W cm^{-2}), which desorbs and ionizes the particles. The resultant ions are then passed to a ToF mass spectrometer, providing a mass spectrum of individual particles in real time. PALMS produces mass spectra for >95% of particles detected by the visible laser (Cziczko et al., 2006).

Since PALMS has a single ToF mass spectrometer, spectra are limited to either positive or negative ions during a given sampling period (Murphy and Thompson et al. 1997a,b). The classification of a particle in either the positive or negative ion mode depends on that particle's chemical makeup, e.g., a mineral dust particle identified in the positive ion mode is typically associated with metal-oxide (Na^+ , K^+ , Al^+ , Fe^+ , FeO^+) markers. An empirical algorithm (employed in the *Interactive Data Language, IDL*), tuned to previously acquired laboratory and field data for ambient atmospheric particles (Cziczko et al., 2013), was used to automate particle classification based on key markers of each particle's mass spectrum. Particle classifications derived using this algorithm are described in Tables 1

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and 2, and include mineral dust, biomass burning particles, soot (elemental carbon), organic and/or sulfate/organic material, heavy oil/combustion products, and sea salt. Due to deviations of particles found in ice core samples from those in an airborne state (Murphy and Thomson, 1997a,b; Cziczo et al., 2013) additional algorithms were developed to distinguish the following classes: (1) calcium-rich (2) “processed” (i.e., containing a broad mixture of both refractory and volatile components), (3) phosphorous-rich biological and (4) phosphorous-rich inorganic particles (Zawadowicz et al., 2016). Additional information on the processing of atmospheric particles trapped in ice is provided in Sect. 2.3.1, and the methodology used for elimination of contamination particles (a result of ice processing) is given in Sect. 2.2.2.

10 2.2 Samples

2.2.1 Ice core samples

The ice core samples used in this study are from two distinct locations in west-central Greenland. DS14 is situated on the Disko Island ice cap (69°39'N, 52°44'W) at an elevation of ~1250 m asl. ~20 km from the coast, and represents a “coastal” site. GW14 is located on the Greenland ice sheet (69°12'N, 44°31'W) at an elevation of ~2250 m, ~240 km from the coast and represents an “inland” site. Average accumulation rates, calculated from age-depth and density profiles, are high at both locations: ~0.5-0.6 m water equivalent (w.eq.) yr⁻¹ and ~0.3-0.4 m w.eq. yr⁻¹ at DS14 and GW14, respectively, allowing for seasonally-resolved age-depth profiles (accuracy for GW14 is estimated to be within 2-3 months; accuracy at the higher melt intensity DS14 site is estimated to be within ± 1 years) using seasonal maxima in water-isotopic and soluble ion chemistry measurements. Samples from both sites are from the modern (last few years) era.

Core processing involved discretization of samples via stainless steel band saw cuts followed by standard sample preparation procedures for firn samples (Osterberg et al., 2006), which included manually shaving the outer 4-5 mm of firn/ice from each sample using a pre-cleaned ceramic ZrO microtome blade under a laminar flow clean bench. All samples remained chilled at -20°C (diurnal variation <± 5°C) from collection until ~2 hours prior to PALMS analysis, minimizing potential volatile losses (Ohata et al., 2013, Wendl et al., 2014). Four samples were analyzed from GW14 (GW14-01 to -

04), sampled discretely at 20 cm intervals between depths of 4-5 m. Each sample corresponded to ~2-3 months over winter 2004 – fall 2005. Six samples were analyzed from DS14 (DS14-01 to -06) sampled discretely at 12.5 cm intervals between depths of ~2-3m, corresponding to the year ~2011 (± 1 year). For the purposes of this study, all samples from each site are presented as compiled results in Sect. 3.2, except DS14-05 (see discussion in Sect. 3.2.3) and DS14-06 (see discussion in Sect. 3.2.2).

2.2.2 Blank tests and insoluble artifacts

To delineate ZrO ceramic or metal (stainless steel) artifacts derived from core processing, band saw cuts were made on frozen ultrapure water controls (18.2 M Ω) and processed as described in Sect. 2.2.1 to infer baseline levels of metal and ZrO contamination. Stainless steel has been observed with PALMS previously and the signature of this material is therefore well known (Murphy et al., 2010). Stainless steel contamination was identified as Fe⁺, Al⁺ and Mo⁺ with occasional Sn⁺ and Ti⁺ and/or TiO⁺ without the presence of alternate mineral/metallic markers (Table 1). Blank tests indicated negligible contamination of stainless steel from core processing when band saws were used ($\ll 1$ sec⁻¹). Contamination artifacts of ZrO-rich particles from the microtome ceramic blade were more common. Due to the low probability of natural stainless steel and ZrO-enrichment (Thomson et al., 1997a,b), these particles were assumed to be indicative of contamination and eliminated from the data.

2.3 Nebulization

Nebulization was used to aerosolize particles after melting the ice samples. The condensed-phase water was then evaporated in the low-humidity nebulization flow, thereby releasing particles. In order to quantitatively analyze the production of aerosol, the liquid solution was nebulized and transported to PALMS at known flow rates. Figure 1a shows a schematic of the experimental setup. Melted sample was placed into a glass container attached to a custom Collision-type atomizer. Dry, inert carrier gas (N₂) was separated into two flows using a rotameter, creating “wet” and “dry” flows, 2.0 L min⁻¹ and 3.0 L min⁻¹ ($\pm \sim 0.1$ L min⁻¹), respectively. The former flow was directed to the atomizer, dispersing the liquid sample into a mist with insoluble particles contained within a fraction of the droplets. [As PALMS generally does not provide spectra for water-saturated particles \(Murphy and](#)

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Thompson, 1995), the atomized particles were then adjoined with the dry flow, dropping the relative humidity and evaporating residual condensed water, resulting in dry residual particles entering the PALMS inlet. The PALMS inlet accommodates a flow of 0.44 L min⁻¹ so the balance of the total flow is filtered and returned to the atmosphere. Samples GW14-01 to GW14-04 and DS14-01 to DS14-04 were analyzed with PALMS in positive and negative ion mode alternately for 10 minutes resulting in a frequency of ~1-2 particles sec⁻¹. Sample DS14-05 was analyzed for one hour while DS14-06 was used for additional solubility tests (Sect. 2.3.1). Between runs, the nebulizer and sampling beaker were cleaned and sonicated for 15 minutes using ultrapure (Milli-Q; 18.2 MΩ) water, and the flow-line flushed continuously with the inert carrier gas.

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2.3.1 Soluble artifacts

An important aspect of analyzing particles re-aerosolized from a liquid suspension, as described in the last section, is the homogenous dilution of soluble components of the original aerosol in the liquid water matrix. Following evaporation of the condensed water during nebulization and transport (Fig 1a), this material is formed from two distinct processes during nebulization and transport to PALMS. First, nebulized droplets not containing an insoluble residual particle evaporate to form small particles comprised solely of the solute ions in the original droplet. Second, droplets containing insoluble residual particles are “coated” with the soluble ions following evaporation of the droplet’s condensed liquid. This may be visualized as a slurry of insoluble mineral dust particles in a water matrix containing a dissolved salt. Nebulized droplets of the slurry may or may not contain a mineral dust particle with the frequency dependent on the concentration of the dust; however, all droplets will contain salt ions with the amount dependent on the concentration in the solution. In this simple system, nebulization would produce either pure salt particles or mineral dust particles coated by salt. We refer to the latter as *post-aqueous coating*. This process is illustrated in Figure 1b.

Residual particles composed purely of soluble material after removal of condensed phase water were not typically observed. This is likely because the amount of soluble material in ~μm diameter droplets produced by the nebulizer is not high enough to produce particles above the lower PALMS detection limit of ~0.2 μm (i.e., the concentration is <1% by volume with respect to water). To quantify

the degree of post-aqueous processing, three experiments were performed: 1) sample DS14-06 was analyzed with PALMS in the negative ion mode to determine background particulate levels, 2) the sample was then filtered through 0.02μm inorganic membrane filters (Whatman Anotop 10) to remove all particulates in the PALMS size range, and 3) the filtered sample was then doped with 746 nm diameter polystyrene-latex spheres (“PSLs”; Polyscience, Inc, Warrington, PA, USA) to provide a single core of a specific chemical composition on which soluble material would coat. For experiment 2) PALMS exhibited a transmission of $\ll 1 \text{ sec}^{-1}$ (i.e., less than 1% the rate without removing insoluble particles from the ice), evidence that droplet residuals were not of sufficient size for detection. Quantification of the post-aqueous coating is discussed further Sect. 3.2.2.

10 2.3.2 Calibrating particle extraction efficiency

A particle extraction efficiency curve was developed to quantitatively determine the number of particles analyzed by PALMS versus the initial number concentration of particles in solution. The extraction efficiency, ϵ , represents the multiplicative probability that a particle of diameter D_p will be (1) nebulized, (2) transported to the PALMS inlet, (3) detected and (4) ablated and ionized with the excimer laser. Both ϵ and PALMS transmission have previously been shown to be a function of aerodynamic particle diameter (Cziczo et al., 2006) so a size-dependent extraction efficiency curve in the particle diameter, $D_p = \sim 0.2 - 3.0 \text{ μm}$ was determined here with monodisperse PSL particles of known number concentrations via the following:

$$\epsilon_{PSL}(D_p) = \frac{n_{PALMS}(D_p) \cdot F_{flow}}{m_{PSL}(D_p) \cdot V_{neb}}, \quad (1)$$

20 where,

$$n_{PALMS}(D_p) = \frac{f_{PALMS}(D_p)}{F_{inlet}}$$

where F_{flow} is the gas flow rate (monitored continuously using a *MesaLabs Bios DryCal 220*; STP cc sec^{-1}); V_{neb} is the rate of liquid nebulization (i.e., a prescribed volumetric loss rate of the sample, determined [here using](#) a scale; $4.4 \cdot 10^{-4} \pm 1.6 \cdot 10^{-6} \text{ mL sec}^{-1}$); $m_{PSL}(D_p)$ is the liquid number concentration (particles mL^{-1}); $n_{PALMS}(D_p)$ is the number concentration of particles analyzed by

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PALMS per unit volume air (particles cc^{-1}), [in turn](#) defined as the frequency of PSL [spheres](#) analyzed by PALMS, $f_{PALMS}(D_p)$ (particles sec^{-1}), normalized by the PALMS inlet flow rate, F_{inlet} (STP cc sec^{-1}). The size dependent [extraction](#) efficiency was assumed to be independent of particle composition and aerodynamic shape, i.e., $\epsilon_{PSL}(D_p) = \epsilon_{particle}(D_p) = \epsilon(D_p)$.

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2.3.3 Polystyrene Latex Sphere (PSL) standards

The determination of $\epsilon(D_p)$ from eq. (1) required particles of known size. Nine PSL sizes were used: 244, 341, 505, 652, 746, 953, 1500, 1990, and 3000 nm diameter, all NIST certified at 1% solid weight percent by volume (w/v) (Polyscience, Inc., Warrington, PA, USA). All PSLs were spherical and near unit density (1.05 g cm^{-3}). The manufacture's weight percentage claim (1% w/v) was assumed sufficient for calculations (Wendl et al., 2014). Using this mass loading fraction, the 746 nm PSL was diluted with ultrapure water (Milli-Q; 18.2 M Ω) until a PALMS transmission response of 3-5 sec^{-1} was reached. This corresponded to a number concentration of $\sim 8.8 \cdot 10^6$ particles cm^{-3} . This standard number concentration was then used as the reference dilution ratio for the remaining PSL standards.

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2.3.4 Mass concentration determination

A mass concentration can be determined with a statistically representative number of particles analyzed by PALMS. Here we define a statistically representative number of particles as the minimum number required to develop a particle size distribution such that outliers, particularly large particles, do not apply erroneous weighting to subsequent mass determinations. In the context of incandescence-based single particle methods (i.e., SP2), Schwarz et al. (2012) have suggested 10,000 particles per sample.

As defined by Wendl et al. (2014), an “external” calibration approach, [as commonly employed in SP2-based measurements of refractory black carbon in glacial snow and ice](#), assumes that ϵ for a given monodisperse PSL standard scales with an unknown polydisperse, [as well as morphologically and compositionally-heterogeneous](#), ice core sample. By rearrangement of eq. (1):

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$$m_{sample}(D_p) = \frac{n_{PALMS}(D_p) \cdot F_{flow}}{\epsilon(D_p) \cdot V_{neb}} \quad (2)$$

Incorporating continuous measurements of the composite flow balance, eq. (2) indicates that determination of the number concentration, $m_{sample}(D_p)$, of a sample containing particulates is feasible using PALMS. Note the external calibration approach does not account for drift in [extraction](#) efficiency via eq. (1), which [may accumulate](#) uncertainty over time. This suggests recalibration of ϵ should be executed periodically based on performance drift.

The normalized logarithmic size distribution of retrieved particles, i.e., those measured by PALMS, for the i^{th} particle class, can be described by:

$$N_{sample}^i(D_p) = \int_{LB(D_p)}^{UB(D_p)} \frac{F_{flow}}{\epsilon(D_p) \cdot V_{neb}} \cdot \frac{dn_{PALMS}(\log D_p)}{d \log D_p} d \log D_p \quad (3)$$

where the efficiency corrected size distribution of particles is integrated under the size-dependent upper to lower [extraction](#) efficiency bounds $UB(D_p)$ and $LB(D_p)$, as determined in 2.3.2. Assuming a spherical shape of all particles, the integrated mass, $M_{sample}^i(D_p)$, of a lognormal size distribution can be calculated by scaling the lognormal volume distribution (Seinfeld and Pandis, 2006) by a particle density representative of that particle class (ρ^i):

$$M_{sample}^i(D_p) = \rho^i \cdot \frac{\pi}{6} D_p^3 \int_{LB(D_p)}^{UB(D_p)} \frac{F_{flow}}{\epsilon(D_p) \cdot V_{neb}} \cdot \frac{dn_{PALMS}(\log D_p)}{d \log D_p} d \log D_p = \rho^i \cdot \frac{\pi}{6} \cdot D_p^3 N_{sample}^i(D_p) \quad (4)$$

Due to the heterogeneous chemical makeup of individual particles, even within a single particle-class, a class-representative ρ^i is subject to some uncertainty. Past studies have illustrated that PALMS can obtain error inclusive, objective estimates of ρ^i (Murphy et al., 2004), provided a large number of particle spectra per class (nominally, [\$>10^4\$](#)).

3 Results and discussion

3.1 Size-dependent extraction, [nebulization](#), and transmission [efficiencies](#)

The size dependent extraction efficiency curve determined from the experimental data is illustrated in Figure 2. The 657 nm diameter particle size showed the greatest efficiency, determined via eq. (1) to be $4.0 \cdot 10^{-3}$ (0.40%). At the min and max PSL sizes, 244 and 3000 nm, respectively, the

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extraction efficiency decreases to $8.6 \cdot 10^{-5}$ and $8.3 \cdot 10^{-6}$ respectively (~ 1 particle out of $\sim 10^4$ or 10^5). A nonlinear least squares approach was used to fit a lognormal distribution to the nine PSL sizes, giving:

$$\varepsilon(D_p) = \frac{1}{2\pi^{1/2} \log(0.57)} \exp\left(-\frac{[\log(D_p) - \log(0.66)]^2}{2 \log(0.57)^2}\right) \quad (5)$$

with a goodness-of-fit statistic (mean-square error) of $1.80 \cdot 10^{-7}$. The computed upper and lower 95% confidence intervals, calculated using an assumption of normally distributed model-observation residuals, encapsulate all data points (Figure 2).

Most particle losses are assumed to occur via “dumping” of excess flow (Figure 1a). Since the PALMS aerodynamic inlet allows a maximum inlet flow rate of 0.44 liters per minute, an inferred $\sim 91\%$ of nebulized particles are lost to this flow. To better delineate the origin of the remaining particle losses, the PALMS transmission efficiency (ε_{trans}) was computed by splitting a monodisperse, particle-laden PSL flow between PALMS and an optical particle sizer (OPS; MesaLabs Bios DryCal 220). A ε_{trans} -curve was calculated as the flow-weighted ratio of the airborne number concentration of particles successfully ionized by PALMS (Figure 1) to that simultaneously measured by the OPS. The experimentally derived ε_{trans} showed a maximum of $\sim 15\text{-}16\%$ efficiency over the particle size range $\sim 300\text{-}1000$ nm (Figure 2), dropping to $\sim 2\%$ at ~ 250 nm and $\sim 1\%$ transmission at 3000 nm. The maximum ε_{trans} determined here was $\sim 5\text{-}6\%$ higher than found by Cziczko et al. (2006). The differences are attributed to adjustments made to the PALMS aerodynamic inlet and sizing lasers since that study.

Provided information on $\varepsilon(D_p)$ and $\varepsilon_{trans}(D_p)$, the experimental nebulization efficiency (ε_{neb} : defined as the number of particles successfully nebulized relative to the number concentration of particles introduced to the nebulizer) can be determined by taking the ratio of $\varepsilon(D_p)$ to $\varepsilon_{trans}(D_p)$ scaled to that of the composite-to-wet flow rate (see Appendix 1 for derivation), such that

$$\varepsilon_{neb}(D_p) = \frac{\varepsilon(D_p)}{\varepsilon_{trans}(D_p)} \cdot \frac{F_{flow}}{F_{wet}} \quad (6)$$

By eq. (6), a nebulization efficiency of $5.1\% (\pm 1\sigma = 0.8\%)$ was found for the 341-1990 nm particle size-range, and $<1\%$ nebulization efficiency at smaller/larger particle sizes.

3.1.1 Comparison of SPMS to SP2-based single particle methods in snow and ice

The SP2 represents the most common online, single particle method currently used for

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measuring particles in glacial snow and ice, providing a benchmark for methodological comparisons to SPMS. The methodology presented here was constructed to allow for a broader size distribution of aerosols sent to PALMS compared to SP2 nebulization techniques used in [most](#) prior ice core studies (McConnell et al., 2007; Ohata et al., 2011, 2013; Schwarz et al., 2012; Wendl et al., 2014) but with [much](#) lower efficiency. For example, Ohata et al. (2011) achieved a max efficiency of ~12%, comparable to Schwarz et al. (2012) and Wendl et al. (2014). However, the SP2 achieves optimal transmission in the low-mid accumulation mode particle size (<400-500 nm; Schwarz et al., 2012), which have lower impaction probabilities (i.e., wall losses during transmission). As such, these studies have commonly incorporated a U5000AT+ ultrasonic nebulizer (Cetac Technologies, Inc., Omaha, NE, USA), which reaches highest nebulization efficiency in the band 100 > D_p > 500 nm (Schwarz et al., 2012); such a range would prove too restrictive for SPMS applications.

[More recent studies have achieved high nebulization efficiencies \(~50%\) up to ~2 \$\mu\$ m using the CETAC Marin-5 pneumatic nebulizer \(Teledyne CETAC Technologies, Omaha, NE, USA; Mori et al., 2016; Katich et al., 2017\). In this context, however, it](#) is noted that achieving too high of an extraction efficiency could be disadvantageous for SPMS, should the number of particles reaching the SPMS inlet exceed that instrument's max transmission rate (~10 particles sec⁻¹ for PALMS; Cziczo et al., 2006) where the limit is data writing and laser repetition rate. This does not affect SP2, which can more rapidly measure the incandescence of carbonaceous material passing through a continuous laser (i.e., 2-3 orders of magnitude higher), though with [the](#) cost of [i](#) not delivering information on [internal](#) mixing state, [or ii](#) aerodynamic size (as opposed to black-carbon volume equivalent diameter). More efficient nebulization at relevant SPMS sizes, coupled to more rapid excimer lasers and data writing, would increase the data acquisition rate.

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3.2 Method application: Greenlandic ice core particulates

3.2.1 Ice core particulate chemical classifications

4931 spectra (2362 positive and 2569 negative) were analyzed from DS14 (DS14-01 to DS14-05), and 553 spectra (233 positive and 320 negative) from GW14 (GW14-01 to GW14-04). The main discrepancy between the number of particles measured between the two sites is not particle loading, but

due to 60 minutes of sampling time for DS14-05 (as opposed to 10 minutes for all other samples) in the positive and negative ion modes. Categorization is broadly divided into natural and anthropogenic sources, as previously noted in modern-era particles from Greenland (Drab et al., 2002, VanCuren et al., 2012), but with some overlap. For example, biomass burning and mineral dust now come from both source-types. Representative particle spectra for all classes, in both the positive and negative ion modes, are provided in [Appendix 2](#).

In positive ion mode, mineral dust is distinguished using primary alkaline markers, Na^+ , K^+ , as well as Al^+ , Fe^+ and other metals and their oxides. A Ca-rich particle sub-class of the mineral/metallic class was delineated by the additional presence of calcium and its oxides (e.g., Ca^+ , CaO^+ , CaOH^+ and Ca_2O^+ and/or CaKO^+). This mineral sub-class may represent a terrestrial carbonate dust source (Murphy and Thomson, 1997b) or marine-derived CaCO_3 or calcium-sulfates. We suggest the latter possibility is consistent with the high prevalence of Ca-rich particles at the coastal DS14 site (~30% of positive particles) relative to the inland GW14 site (~8%; Figure 3). A fraction of the mineral/metallic-rich particles were mixed with organic components (e.g., carbon cluster isomers, C_n^+ , hydrocarbons, organo-nitrogen markers). This particle sub-class was delineated as “processed”, as it is likely affected by post-aqueous processing; at this time we cannot fully decouple this artifact from an original atmospheric association (see also Sect. 3.2.2. below).

The biomass-burning category contains signatures of carbon-cluster isomers, elevated K^+ and/or potassium sulfates (K_3SO_4^+). These particles were the second most abundant positive-ion mode type at DS14 (31%) with lower representation at GW14. In the negative ion mode, sulfate-organic rich spectra made up the largest proportion of particles at DS14 (>66%), but comprised just over one third of particles at GW14 (Figure 3). Inspection of the organic/sulfate class at both sites indicated that a substantial proportion of particles also contained elevated PO_2^- and PO_3^- . Past studies have noted the challenge in identifying the origin of P-rich particles, with both mineral and biological origin possible (Creamean et al., 2014). The classification scheme by Zawadowicz et al. (2016) was adopted here to identify the P-rich particle subset as either of biological or inorganic origin (Table 2), with the latter suggestive of mineral dust or fly ash particles. The biological particles were found to be largest contributor of particles analyzed at DS14 in negative ion mode, 39%. These particles were also found to

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have the largest size of all the particle classes at both sites, with a median geometric diameter of 705 nm. This is consistent with a local marine biogenic source (Figure 4). Additionally, we note that biomass burning and biogenically-sourced particles can appear similar in the positive ion mode, especially if post-aqueous processing is involved (Zawadowicz et al., 2016). Due to similarities in particle size (median particle diameter within ~10 nm; Figure 4) and relative abundance with the biological class, we interpret the positive ion mode biomass-burning particles to be of equivalent marine-biological origin at the DS14 site.

Soot is distinguished in both the positive and negative ion modes by the presence of elemental carbon (e.g., C^+ , C_2^+ , C_3^+ , etc., or C^- , C_2^- , C_3^- , etc.) with organic peaks. Soot was common at both sites in the negative ion-mode, showing an abundance (composite between positive and negative spectrum) of ~18% and 28% at DS14 and GW14, respectively. Both abundances are considerably higher than previously reported by Drab et al. (2002) in a firm sequence at Summit, Greenland, who estimated <5% of particles were soot or combustion-based although the authors noted the difficulty in optically evaluating these particles, due their tendency to aggregate. A currently unknown feature of the data is the high negative polarity soot abundance at the GW14 site that appears to correspond to positive polarity mineral or metallic particles. This feature may be indicative of an anthropogenic particle influence. The heavy oil combustion class, distinguished by elevated vanadium markers (V^+ and VO^+), was of low abundance at both sites, representing <0.1% of all particles measured (not shown in Figure 3).

3.2.2 Post-aqueous processing

The inclusion of sea salt in both the positive and negative ion modes suggests an artifact arising from post-aqueous processing. This class was observed at both GW14 and DS14 but at low abundance (<1-2%). Sea salt aerosol would have dissolved upon melting of the ice samples prior to analyses. This classification may therefore represent evaporation of an uncommonly large droplet or a coating where the underlying particle composition is not observed. In addition, a non-negligible percentage of the positive-ion mode particles from both sites were not readily characterized, with “unclassified” particles comprising 8% and 6% of particles at DS14 and GW14, respectively. Manual inspection of spectra from this class indicates particles containing key markers from numerous classes (Cziczo et al., 2013),

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thereby leading to confusion of the classification method. This may represent particles undergoing severe post-aqueous processing or coagulation within the solution or after nebulization and led to the work discussed in more detail in the next section.

We note that a single-particle mass spectrometer similar to PALMS was recently used to measure chemical composition of insoluble particles in precipitation samples collected in Sierra Nevada (Creamean, et al. 2013, 2014, 2016). In those studies, post-aqueous processing was not investigated in detail and single particle classifications were not adjusted based on the possibility of processing.

3.2.3 Solubility tests

Atmospheric particles are not chemically homogenous and often contain both soluble and insoluble components (Murphy, 2005). Several particle classes presented here, specifically the sea salt and organic/sulfate classes and, to a lesser extent, the P-rich and Ca-rich classes, were inferred to contain an appreciable proportion of soluble components, though of indeterminate origin (i.e., of atmospheric or post-aqueous processing origin). To test the degree of post-aqueous coating to these four particle classes, four negative ionic species were chosen as class-representative markers: the ions $\text{PO}_2^- + \text{PO}_3^-$ ($m/z = 63 + 79$) for P-rich biological particles, $\text{SO}_3^- + \text{HSO}_4^-$ ($m/z = 80 + 97$) for sulfate-rich/organic particles, $^{35}\text{Cl}^- + ^{37}\text{Cl}^-$ ($m/z = 35 + 37$) for sea salts and Cl^- -rich mineral dust, and $\text{CN}^- + \text{CNO}^- + \text{NO}_2^-$ ($m/z = 28 + 42 + 48$) for soluble organics. The peak areas of all four ionic species were calculated in particulates analyzed in an initial sample run of DS14-06, considered the control experiment. The sample was twice filtered using a $0.02\text{-}\mu\text{m}$ sterile mesh filter to remove insoluble particles. The sample was then doped with 746 nm PSL particles to act as a carrier of any soluble material. A blank test was performed using DI water doped with the same 746 nm PSL particles.

For all four species the observed peak areas decreased for the filtered (insoluble particle-free) experiments to the DI and filtered particle tests (Figure 5). Wilcoxon rank sum tests were used to determine the difference in the median peak area of the two experiments (i.e., the DS14-06 sample run versus the DI and the PSL-doped DS14-06 sample run). In both the ($\text{PO}_2^- + \text{PO}_3^-$) and ($\text{CN}^- + \text{CNO}^- + \text{NO}_2^-$) tests the two experiments' median peak areas were significantly different ($p < 0.05$), indicating that negligible post-aqueous coating of soluble phosphorous ($\text{PO}_2^- + \text{PO}_3^-$) or soluble organics ($\text{CN}^- +$

CNO⁻ + NO₂⁻) were found on the PSLs. These results suggest that P-rich particulates (the biological and P-rich inorganic classes) and organo-nitrogen particulates measured in PALMS are likely comprised predominantly of the original insoluble particle. This was not the case for the chlorine and sulfate-containing particles, indicating these materials were found in the solution. Since sea salt and sulfate-rich/organic particles are soluble and a significant source of atmospheric chlorine and sulfate (Murphy et al., 1998), this suggests that when using this or similar nebulization methodology, these particle classes (1) cannot be directly retrieved, and (2) cause coating of insoluble particles. More generally, the relatively small particle sizes of the organic/sulfur-rich particles at both GW14 and DS14 (Fig. 4) suggest that post-aqueous coating could be increasingly important for smaller sized particles.

3.2.4 Feasibility of PALMS for particle concentration determinations

We found the number of particles measured during the 10 minute sample runs described above were insufficient to quantitatively discern mass concentration within the ice. To test the outcome of longer sample runs, sample DS14-05 was run for an hour in both the positive and negative ion modes. This resulted in multiple classes containing >500 particles, the threshold number of particles employed here for class-dependent mass concentration measurements, but still an order of magnitude smaller than the number suggested by Schwarz et al. (2012) for laser-induced incandescent methods. Organic/sulfur-rich particles were not considered due to the uncertainty regarding post-aqueous processing (see prior section). Representative class densities were taken as 0.8 g cm⁻³ for soot (denoting the effective density of black-carbon; Moteki and Kondo, 2010; Kiselev et al., 2010; Schwarz et al., 2012), 2.7 g cm⁻³ for mineral dust/metallic particles (Murphy et al., 2008), and 1.3 g cm⁻³ for biomass burning and biological particles (Li et al., 2016). To calculate mass concentrations, particle size distributions for each class were integrated over the order of magnitude size interval $D_p = 0.25$ nm to 2.5 μ m using eq. (4) and parameterized using the experimentally derived extraction efficiency provided in eq. (5). Note that while no systematic trends in nebulization drift were found over either hour-long measurement period, short term fluctuations in nebulization could occur; for the present experiment, such fluctuations are assumed to be encapsulated as uncertainty about the extraction efficiency parameterization (eq. 5; see Supplementary Material for details). Under these assumptions, the mass concentration results for DS14-

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05 are shown in Table 3; the mineral/metallic particle class (including the Ca-rich and processed particles) had the highest concentration of particles (consistent with the high relative abundance in DS14-05), estimated at $10.1 \pm 5.7 \text{ ng g}^{-1}$, while the biomass-burning/biological, P-rich biological, and the soot particle classes were estimated at 5.6 ± 2.5 , 7.0 ± 2.6 , and $1.6 \pm 0.7 \text{ ng g}^{-1}$, respectively.

5 Direct comparison of these concentrations to previously published values is complicated by differences in the particle size range retrieved by various analytical techniques as well as site-to-site differences. For example, Coulter counter techniques can extend the size-range of insoluble particle retrieval up to $50 \mu\text{m}$ (Delmonte et al. 2002) where a few particles near the upper limit would dominate the mass. Where comparisons are possible, the concentrations reported here are in line with what has
10 been previously reported. For example, mineral/metallic concentrations of $10.1 (\pm 5.7) \text{ ng g}^{-1}$ for DS14-05 are within an order-of-magnitude of that reported by Bory et al. (2002) for recent snowfall at the higher-elevation, inland NGRIP site in Greenland.

A more direct comparison of SP2 vs. SPMS methodologies is available utilizing black carbon measurements made recently on DS15, a firm core collected in 2015 adjacent to the DS14 site. Using the
15 SP2 method (following the methodology of McConnell et al., 2007; see also Wendl et al., 2014), a mean black carbon concentration in the upper 5 m of DS15 was found to be $0.78 \pm 0.97 \text{ ppb}$ (J. McConnell, *pers. comm.*), within range of the soot concentrations ($1.6 \pm 0.7 \text{ ng g}^{-1}$) calculated using PALMS on the selected core segment of DS14. There are notable inherent differences between SPMS-
20 and SP2-derived soot size distribution determinations, however. Namely, whereas SP2 can differentiate the volume-equivalent diameter of refractory soot-components in compositionally-heterogeneous particles (Schwarz et al., 2012), our SPMS approach presumes a particle to be comprised wholly of soot if its mass-spectrum is classified as such. Thus, depending on the varying morphologies, internal mixing states, and ionization potentials of the analyzed soot particles, SPMS may be subject to positive size distribution biases (Fig. 4).

25 While our results show potential exists for using SPMS to determine insoluble mass concentrations of particles in snow and ice, they also identify areas where more work is needed before SPMS can be used as a quantitative tool. These include: i) executing multiple extraction efficiency calculations as a function of particle class (in addition to size), ii) incorporating regularized tests for

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drifts in SPMS extraction efficiency and (or) regularly employing “blank” tests between sample measurements in order to improve delineation to changes in background particulate levels, iii) achieving a greater number of particle measurements, either through improvements in particle extraction/PALMS transmission or longer sample integration times, and iv) comparing SPMS-derived particle concentrations with results from alternate, well-founded high-precision instrumentation (e.g., an Ultra-High Sensitivity Aerosol Spectrometer (UHSAS; Droplet Measurement Technologies Inc., Boulder, CO), or Coulter Counter techniques).

4. Conclusions

In this study we develop and apply a new methodology utilizing SPMS to characterize particulates trapped in snow and ice at the single particle level. We show that a single instrument, in this case PALMS, can be used to discern the aerodynamic size, composition, and concentration of insoluble ice core particulates. This online method reduces preparation time and resources required for filter-based particle retrieval methods. Based on compositional differences in particles found in two Greenlandic firn cores, we define 8 distinct particle classes in the positive ion mode and 6 in the negative ion mode (Tables 1 and 2; Appendix 2). These classes are a combination of common atmospheric types previously described in the literature and types relatively rare in the atmosphere but common in the ice samples. Differences in the relative abundances of classes found between the two sites are consistent with the sites’ unique geography and climatology, notably, marine versus high altitude/inland locations. Furthermore, this study demonstrates the feasibility of using PALMS to infer a sample’s mass concentration using an external calibration of a size-dependent extraction efficiency which parameterizes the nebulization, delivery, and ablation/ionization of particles from concentration slurries (Wendl et al., 2014).

We find that two classes, organic/sulfate and sea-salt particulates, appear to be artifacts of post-aqueous processing, highlighting the need for further evaluation of these particle classes in future studies. More broadly, this phenomenon is a critical feature of particles analyzed from a liquid suspension, which has not been fully appreciated in previous studies (e.g. Ault et al., 2011; Creamean et al., 2014): completely soluble aerosols or soluble fractions of particles are homogenized into the melt

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solution from which insoluble components are extracted. These components are then present as small, solute-only, particles and as a surface coating on insoluble particles. The former were typically smaller than the PALMS threshold size but this need not be the case in future studies using different instruments. While the concentration of soluble material can be reduced by dilution, at this time there is no solution to quantifying the original soluble aerosol, finding the original association of soluble with insoluble components, or completely removing this material from the analysis step.

We group the future application of SPMS to ice core studies into qualitative and quantitative uses. We show that SPMS can be readily used for qualitative analysis, as is done in atmospheric studies (Murphy, 2005); future application may include particle classification and source-allocation using mass-spectral compositional markers. Quantitatively, we show that mass concentration may be feasible but with long sample periods required for statistically-relevant conclusions at current instrument rates. Measurement frequency during an hour-long period registered $\sim 1\text{-}2$ particle sec^{-1} . Instrumental improvements, or e.g. sample concentrations, leading to ~ 10 sec^{-1} could reduce the required sample by a similar factor. This suggests that further instrument improvements in extraction efficiency, including nebulization as well as data/laser repetition rate, would correspondingly reduce sampling time.

Data Availability

All data used in the generation of this manuscript is archived at MIT and the Harvard Dataverse and is available upon request.

Appendix

A1. Calculating nebulization efficiency

Here, we derive the determination of nebulization efficiency (ϵ_{neb} ; Sect. 3.1). We define ϵ_{neb} as the flow-weighted ratio of the rate of successfully nebulized particles per unit time relative to the (liquid) number concentration of particles introduced to the nebulizer, such that

$$\epsilon_{neb} = \frac{f_{neb}(D_p)}{m_{PSL}(D_p) \cdot F_{neb}} \quad (A1)$$

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where $m_{PSL}(D_p)$ and F_{neb} are as defined in eq. (1), and $f_{neb}(D_p)$ is the frequency of particles successfully nebulized (e.g., particles sec^{-1}) as a function of PSL diameter (D_p). In this case, $f_{neb}(D_p)$ is the quantity that must be solved for. We take

$$f_{neb}(D_p) = \frac{n_{PALMS}(D_p) \cdot F_{flow}}{\varepsilon_{trans}(D_p)} \cdot \left[\frac{F_{flow}}{F_{wet}} \right] \quad (A2)$$

such that the scalar quantity $\left[\frac{F_{flow}}{F_{wet}} \right]$ acts as a correction for the flow balance of particles actually passing through the nebulizer (Figure 1) and $\varepsilon_{trans}(D_p)$ corrects for the size-dependent particle transmission of PALMS. Note that $n_{PALMS}(D_p)$ and F_{flow} are as previously defined in eq. (1). Plugging eq. (A2) into (A1), and via relation to eq. (1),

$$\varepsilon_{neb} = \frac{\varepsilon(D_p)}{\varepsilon_{trans}(D_p)} \cdot \left[\frac{F_{flow}}{F_{wet}} \right] \quad (A3)$$

as defined in eq. (6).

A2. PALMS mass spectra

In this section, we provide exemplary particle spectra for each class, including the traditional particulate classes using the algorithm described in Cziczo et al. (2013), as well as the additional particle classes used in this study, in both the positive and negative ion mode. All major peaks corresponding to known ionic fragments have been labeled (see e.g., Murphy and Thompson, 1997 a, b; and Cziczo et al., 2013 for details).

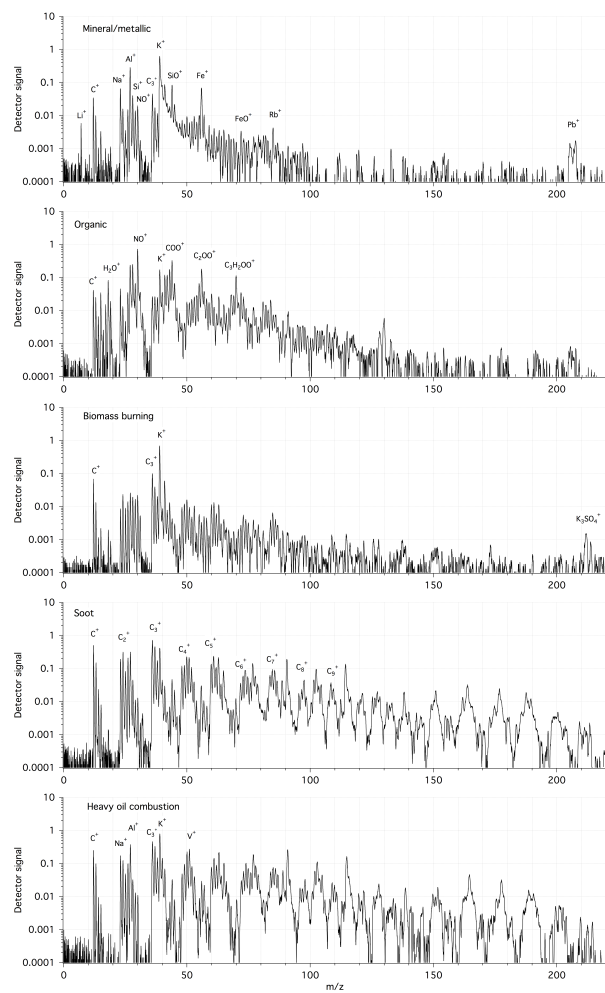
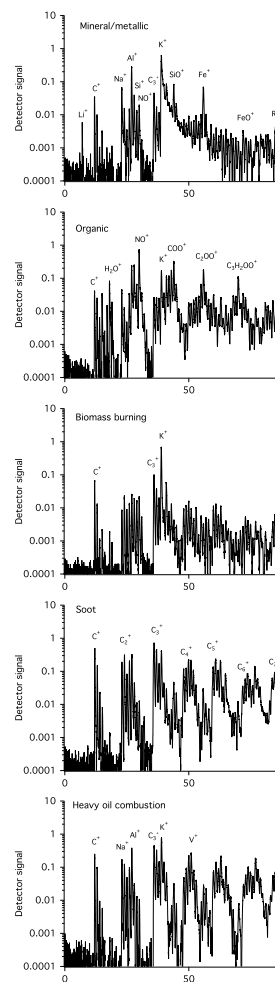


Fig. A1. Exemplary spectra taken from the DS14 and GW14 samples for the traditional positive ion mode particle classes using the classification algorithm described in Cziczo et al. (2013). From top to bottom: mineral/metallic, organic, biomass burning, soot, and heavy oil combustion/vanadium-rich.

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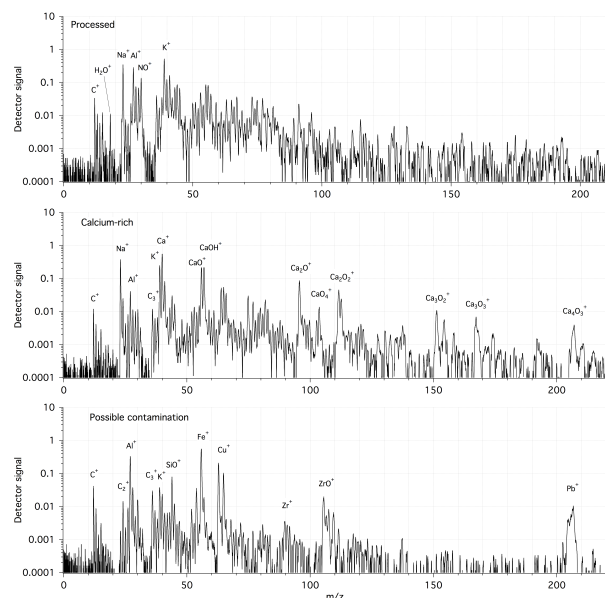
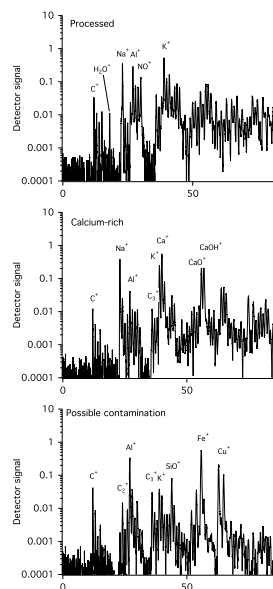


Figure A2. Exemplary spectra taken from the DS14 and GW14 samples for three positive ion mode classes commonly observed in the ice core data. From top to bottom: processed, Ca-rich, and contamination/heavy-metal. These classes were originally categorized in the mineral/metallic positive ion category using the algorithm of Cziczo et al., (2013).

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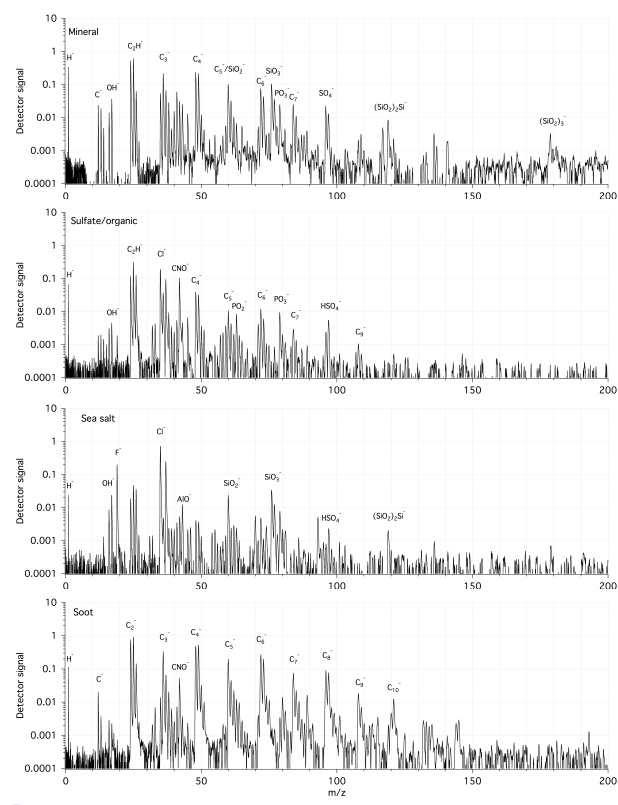
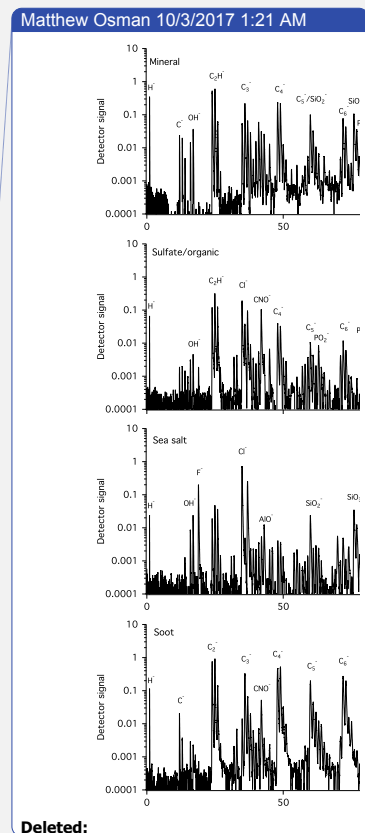


Fig. A3. Exemplary spectra taken from the DS14 and GW14 samples for the traditional negative ion mode particle classes using the classification algorithm described in Cziczo et al. (2013). From top to bottom: mineral, sulfate/organic, sea salt, and soot.



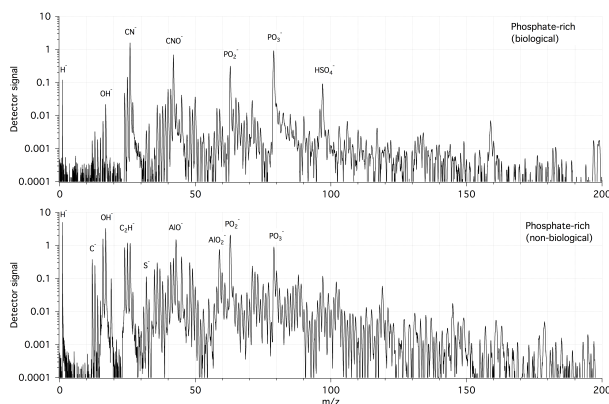


Figure A4. Exemplary spectra taken from the DS14 and GW14 samples for two negative ion mode classes commonly observed in the ice core data. From top to bottom: biological and P-rich inorganic. Both classes were originally categorized in the sulfate/organic negative-ion category using the algorithm of Cziczko et al., (2013).

Author Contribution

M.O. helped design the experiment, prepared the ice core samples, analyzed the samples, contributed to the data analyses, and wrote the manuscript. M.Z. helped design the experiment and contributed to the data analyses. S.B.D. collected the DS14 and GW14 ice cores, assisted with ice core sample preparation. D.J.C. helped design the experiment and supervised the laboratory work. All authors participated in writing and editing the manuscript, and contributed to the interpretation of the results.

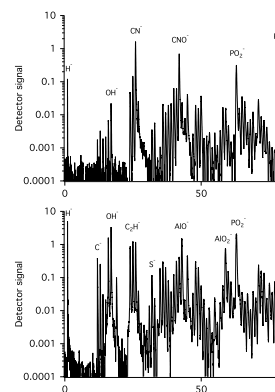
Competing interests

The authors declare that they have no competing conflicts of interest.

Acknowledgements

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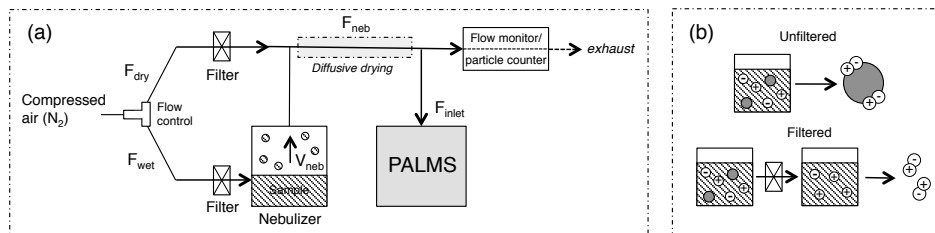
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- 5 Figure 1. (a) Schematic of the experimental setup. Inert air (N_2) is split into dry and wet flows, with the latter, V_{neb} , sent to nebulize the melted ice. The low relative humidity of the combined flow evaporates condensed water and sends dried particles to PALMS. Air and liquid flows (F and V) were continuously monitored. Parker IDN-4G filters were used to remove particles from the dry nitrogen gas. (b) Idealized representation of the two types of particles produced by a slurry of insoluble particles in a solution. In the top case an insoluble particle is coated with soluble ionic material initially dissolved in the condensed liquid. In the bottom case an insoluble particle-free solution produces residual particles of the dissolved material (see text for details).
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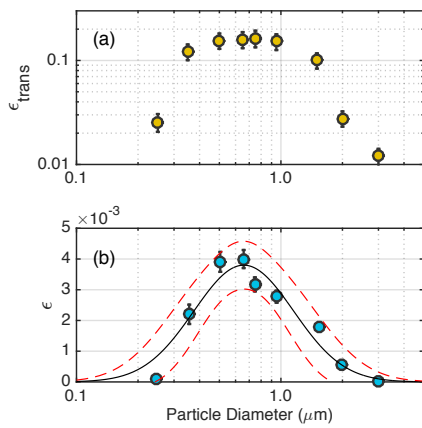


Figure 2. (a) PALMS transmission efficiency, ϵ_{trans} , as a function of aerodynamic particle size. (b) The system extraction efficiency, ϵ , as a function of particle size. The black line is the least squares lognormal regression with 95% confidence limits as red-dashed lines. Error bars represent the propagated standard error for all parameters in eq. (1).

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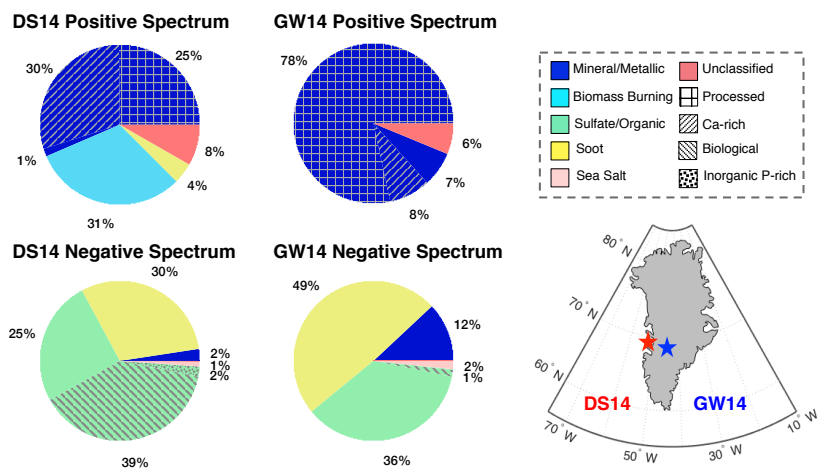


Figure 3. Particle class abundances at the DS14 and GW14 sites, and their relative locations in Greenland (lower left). The Ca-rich and processed particle sub-classes discussed in the text are associated here with the mineral/metallic classes. Likewise, and biological and inorganic P-rich sub-classes are associated with the sulfate/organic class. Particle classes below the 1% abundance level are not shown and contaminant particles from ice core processing were eliminated before data analysis. See Sect. 3.2 for additional details.

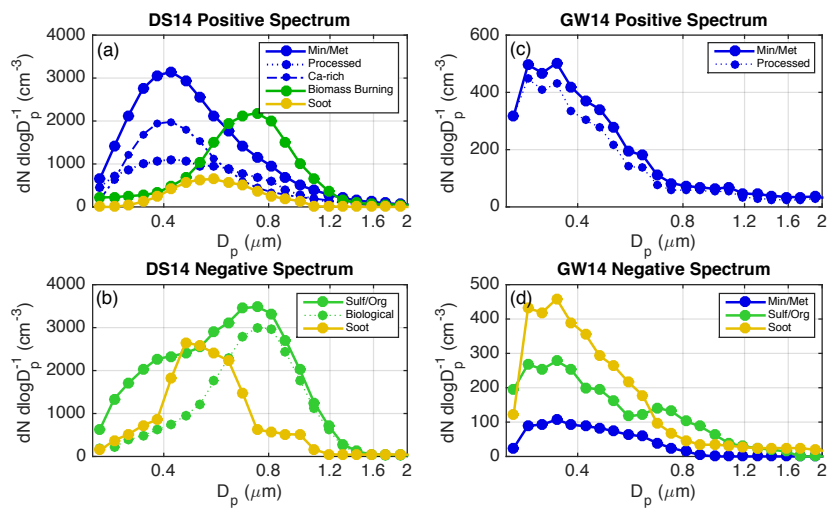


Figure 4: Log-normalized 3-pt smoothed size distributions for particle classes containing >100 particles at DS14 (a-b) and GW14 (c-d) in positive (a-c) and negative (b-d) ion mode. Due to the PALMS lower size limit the x-axis origin is 0.25 μm .

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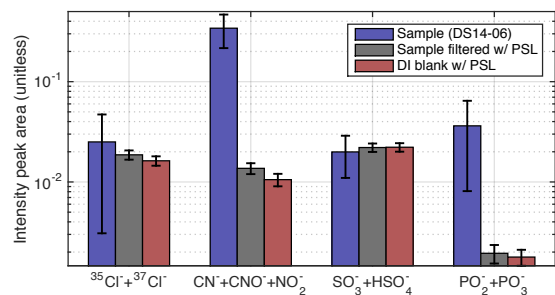


Figure 5. Median normalized peak intensity for $\text{PO}_2^- + \text{PO}_3^-$ ($m/z = 63 + 79$), $\text{SO}_3^- + \text{HSO}_4^-$ ($m/z = 80 + 97$), $^{35}\text{Cl}^- + ^{37}\text{Cl}^-$ ($m/z = 35 + 37$), and $\text{CN}^- + \text{CNO}^- + \text{NO}_2^-$ ($m/z = 28 + 42 + 48$) for particles in DS14-06 filtering experiments: unfiltered (control; blue), filter doped with PSL particles (grey), and DI-water blank doped with PSL particles (red). Error bars represent one median absolute deviation, estimated using a bootstrap (resampling) approach ($n = 1000$). Note the post-aqueous coating in these four quantities exhibits no significant difference between the DI blank and the filtered samples.

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Tables

Table 1. Positive spectra classification.

Particle Class	Primary Identifier
Mineral/metallic	Na ⁺ , Al ⁺ , K ⁺ , Fe ⁺ present, possible Si ⁺ and/or SiO ⁺
Calcium-rich	Ca ⁺ , CaO ⁺ , CaOH ⁺ and Ca ₂ O ⁺ and/or CaKO ⁺ present
Processed	Combination of Na ⁺ , Al ⁺ , K ⁺ , Fe ⁺ and abundant organics, sulfates and nitrates
Organic	Organics, NO ⁺ , NO ₂ ⁺ , no other characteristic markers
Biomass burning/biological	High K ⁺ , sulfates and organics without other mineral/metallic markers
Soot	C ⁺ , C ₂ ⁺ , C ₃ ⁺ , C ₄ ⁺ , etc. V ⁺ and VO ⁺
Sea salt	Na ⁺ , K ⁺ , Na ₂ Cl ⁺
Heavy oil combustion	V ⁺ and VO ⁺
Heavy metal (contamination)	High Fe ⁺ , possibly Al ⁺ , Mo ⁺ and/or Sn ⁺ without other mineral/metallic markers (stainless steel) Ti ⁺ and TiO ⁺ without other mineral/metallic markers (processing saw) Zr ⁺ and ZrO ⁺ (processing saw)

5

Table 2. Negative spectra classification.

Particle Class	Primary Identifier
Biological ¹	PO ₂ ⁻ , PO ₃ ⁻ , CN ⁻ , CNO ⁻
P-rich (inorganic) ¹	PO ₂ ⁻ , PO ₃ ⁻ , CN ⁻ , CNO ⁻
Mineral	SiO ₂ ⁻ , SiO ₃ ⁻ , possible SiO ₂ AlO ₂ ⁻
Sea salt	F ⁻ , Cl ⁻ no other characteristic markers
Soot	C ⁻ , C ₂ ⁻ , C ₃ ⁻ , C ₄ ⁻ , etc.
Sulfate/organic	CN ⁻ , CNO ⁻ , other organics, HSO ₄ ⁻ no other characteristic markers

¹Classes differentiated by ratios of PO₃⁻ to PO₂⁻ and CN⁻ to CNO⁻, as described in Zawadowicz et al. (2016).

10 Table 3. DS14-05 particle relative abundance (RA) and calculated mass concentration values. The values of ρ are inferred based on prior studies (see Sect. 3.2.3).

Particle Class	Particle count (RA, %)	ρ (g cm ⁻³)	Concentration (ng g ⁻¹) ($\pm 1\sigma$)
<i>Positive spectrum</i>			
Mineral/metallic ¹	1229 (54.8%)	2.7	10.1 (± 5.7)
Biomass burning/biological	721 (32.1%)	1.3	5.6 (± 2.5)
<i>Negative Spectrum</i>			
P-rich (biological)	970 (40.3%)	1.3	7.0 (± 2.6)
Soot	706 (29.3%)	0.8	1.6 (± 0.7)

¹Includes pure mineral/metallic particles, as well as the subsidiary Ca-rich and Processed particle classes.

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