This document includes:

i. Point-by-point responses to Referee #1 and Referee #2.
ii. The revised manuscript (including changes, as requested by Referee #1 and #2, which are highlighted in yellow. Grey highlights are for general edits.
iii. Manuscript supplement.

Manuscript format description:

Black text shows the original referee comment, blue text shows the authors response, and red text shows quoted manuscript text. Changes to the manuscript text are shown as *italicized and underlined*. We used bracketed comment numbers for referee comments (e.g., [R1.1]) and author's responses (e.g., [A1.1]). Line numbers refer to the discussion/review manuscript.

Response to referee comments and suggestions on amt-2018-390 by Könemann et al.

Anonymous Referee #1

Received: 25 November 2018

General comment:

The paper describes an instrument that should be much superior to the WIBS or UVAPS in characterizing fluorescent aerosol. The 16-channel fluorescence spectra should provide far more information for characterizing aerosol than existing WIBS instruments, while still measuring very large numbers of spectra per day. This instrument appears to be able to significantly expand our understanding of bioaerosols and other fluorescent particles in the atmosphere. The paper is clearly written. It should be published. I suggest the authors think about the following.

Author response: We want to thank Referee #1 for his/her positive and constructive assessment.

Specific/technical comment:

[R1.1] The title and abstract say, "a new instrument for . . ." Then line 144 is: "Introduced here is a new instrument for the detection and characterization of individual particles; the Spectral Intensity

Bioaerosol Sensor (SIBS, Droplet Measurement Technologies)." Then later the text says, "The SIBS was originally designed and marketed to record time-resolved fluorescence lifetime." If it has already been marketed, the use of "new" seems possibly inaccurate. How long has it been marketed? I suggest dropping "new" from the title and the text. There is no need for it. Also, a book chapter by Huffman (one of the authors of the present paper) and Santarpia, "Online Techniques for Quantification and Characterization of Biological Aerosols," in Microbiology of Aerosols eds., Anne-Marie Delort and P Amato (2017) mentions both types of SIBS (the breakdown spectroscopy SIBS and the spectral intensity SIBS). That chapter was published over a year ago and was presumably written many months before that.

[A1.1] We agree with Referee#1 and took out the word "new" from the title and abstract. It is true that the SIBS was briefly introduced within the book chapter "Online Techniques for Quantification and Characterization of Biological Aerosols" (Huffman and Santarpia, 2017). This reference is based on the same unit as discussed in amt-2018-390 and referenced by a conference poster, because no other citation was available at that time. Information stated in this book chapter was based on unpublished and non-peer-reviewed data, available because we had already been working together with Alex Huffman in 2015 with respect to the earliest version of the SIBS. Since then, the instrument underwent many modifications (hardware and software) and revisions for which the SIBS unit from 2015 and the unit in its current state are not comparable anymore.

[R1.2] A new and noteworthy part of this paper (maybe the most new and noteworthy part) is that the instrument is commercially available. Instruments that could do the key parts of what is done here (two fluorescence spectra each with a different excitation wavelength is measured for each particle) have been around for some years, e.g., Huang, Pan et al., and Pan et al. But routine measurements were far from feasible by others. I suggest stating in the abstract and introduction that the instrument is built by DMT and commercially available. I suspect more people will read it if they know they could buy one. Many instruments described in papers, especially new instruments, can only be used by the researchers that built and know how to use them.

[A1.2] As suggested by Referee#1, we added a reference to DMT within the abstract and conclusions for clarification. The linkage between the SIBS and DMT is already given, within the introduction, in:

(P5, L155-156): "Introduced here is a new instrument for the detection and characterization of individual particles; the Spectral Intensity Bioaerosol Sensor (SIBS, Droplet Measurement Technologies)."

[R1.3] RE: "originally designed and marketed to record time-resolved fluorescence lifetimes". Are spectra required for measuring fluorescence lifetimes? Was the SIBS designed and marketed to measure spectra at two excitation wavelengths? I think what is meant is: It was designed and marketed to measure time- and spectrally-resolved fluorescence lifetimes.

[A1.3] Correct. The SIBS was originally designed to measure time- and spectrally-resolved fluorescence lifetimes at two excitation wavelengths. As suggested by Referee #1, the following sentence was changed from:

(P15, L495-496): "The SIBS was originally designed and marketed to record time-resolved fluorescence lifetime."

To (P15, L495-496): "*The SIBS was originally designed and marketed to record time- and spectrally-resolved fluorescence lifetimes at two excitation wavelengths.*"

[R1.4] Make Fig. 2 higher resolution so it can be seen in detail on a large monitor.

[A1.4] Within the current manuscript version, figures were used in lower resolution to keep file sizes as low as possible. The final version will include high resolution images and figures.

[R1.5] RE: SIBS (Spark Induced Breakdown Spectroscopy) already has a meaning in the measurement of aerosol particles, either single particles or many at a time. It is confusing to see SIBS used for the name of an instrument that has nothing to do with spark induced breakdown. SIBS (original meaning) provides information similar to LIBS, i.e., elemental composition of single-particles or multiple-particles. I imagine SIBS (or LIBS) may eventually be combined with an instrument such as the SIBS described in this paper, to provide both breakdown spectra and fluorescence spectra for each particle. Since the SIBS of Konemann et al., is already marketed, and been around for a while, it is likely too late for this comment to be relevant, but I hope not.

[A1.5] It is indeed unfortunate that two similar acronyms exist for two different instruments. We added the following sentence to hopefully avoid potential misconceptions, including references as suggested by Referee#1:

(P12, L370-373): "*To avoid potential misunderstanding, it is important to note that the SIBS described in this study is not related to spark-induced breakdown spectroscopy instrumentation, which uses the same acronym (e.g., Bauer & Sonnenfroh, 2009; Hunter et al., 2000; Khalaji et al., 2012; Schmidt & Bauer, 2010).*"

It is true that the combination of both breakdown- und fluorescence spectra on single particle scale would provide a completely new level for particle characterization. However, this topic is beyond the scope of this manuscript.

References

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Response to referee comments and suggestions on amt-2018-390 by Könemann et al.

Referee #2 Dr. Ian Crawford

Received: 4 December 2018

General comment:

This paper examines the technical capabilities of the new SIBS UV-LIF bioaerosol spectrometer and describes several technical corrections and calibrations that are necessary to deliver high quality and accurate data products. As a long term WIBS user it is encouraging to see the next generation of high spectral resolution UV-LIF spectrometers that are coming to market being examined in detail early on in their lifecycle; while there is still undoubtedly still utility in broadband spectrally integrated instruments such as the WIBS for broad bioaerosol detection, it has been clear for some time now that deeper specificity/classification requires greater spectral resolution so these technical developments are timely. The authors present a fair assessment of SIBS capability to resolve key biofluorophores and make a number of suggestions and cautions that apply to the SIBS and also UV-LIF spectrometers generally. Overall the paper is well written and the technical validation experiments are well thought out. The results and methodologies reported here will serve as a useful framework for assessing the performance of other multichannel high spectral resolution UV-LIF spectrometers which are entering circulation. I recommend publication after the following comments have been addressed.

Author response: We want to thank Dr. Crawford (Referee #2) for his positive assessment and constructive suggestions.

Specific/technical comment:

[R2.1] L98: Can you please check the size range reported for the WIBS-NEO. It is my understanding that the instrument sizes over the range of 0.5-30 μm.

[A2.1] Thanks a lot for pointing that out. The size range we stated for the WIBS-NEO originated from information we had in the beginning of 2017. Since then, DMT seem to have updated related information. The size range, within the manuscript, is now changed from ~0.3 – 100 μm to ~0.5 – 30 μm for the WIBS-NEO, according to: http://www.dropletmeasurement.com/wideband-integrated-bioaerosol-sensor-wibs-neo

[R2.2] L125: I think that a short sentence summarising some of the validation work would round this out while showing some of the limitations of the instrument/approach. A statement on how the Crawford et al. (2015) method was validated by Gosselin et al. (2016) by showing a good correlation between fungal molecular tracers and assumed fungal clusters but poor agreement between bacterial tracers and assumed bacterial clusters would contextualise this. It may also be worth commenting that the relatively high lower size limit of 0.8 µm used in this study due to instrument limitations may have impacted the latter which may potentially be alleviated by an improved lower detection limit.

[A2.2] As suggested, the following sentences have been added to round out the topic of currently used clustering approaches regarding online LIF:

(P4-5, L130-139): *"For example, it was shown for a rural forest study in Colorado that a cluster derived using WIBS-3 data, assigned to fungal spores (Crawford et al., 2015), correlated well with the mass concentration of molecular fungal tracers (e.g., arabitol and mannitol) measured with offline chemical techniques (Gosselin et al., 2016). In contrast, the clusters in the same study that were assigned to bacteria correlated only poorly with endotoxins, used as bacterial molecular tracers (Gosselin et al., 2016). This provides evidence of a limitation to using LIF instrumentation with low spectral resolution to separate or identify some PBAP types. Additionally, the bacterial cluster allocation might have also been hampered in that case by the minimum detectable particle size of the WIBS (~0.8 µm), resulting in a lower detection efficiency for bacteria."*

[R2.3] L209: Can you comment further on the choice of 1σ thresholding use here. I appreciate that the conventional wisdom used to determine the threshold for WIBS instruments may not carry over here due to the differences in the optical setup but 3σ and 9σ thresholds are used later in the paper when reporting ambient concentrations.

[A2.3] As pointed out by Dr. Crawford, it is currently unknown if thresholding strategies conventionally used for several WIBS models perform similar when applied to the optical setup of the SIBS. For the current manuscript, we decided to use a rather simple 1σ approach, because for the assessment of the spectral accuracy, measuring sets of homogenous particle types (PSLs, biofluorophores), the thresholding plays only a minor role. In contrast, conventional thresholding strategies were applied to a set of ambient data as a first attempt to qualitatively match SIBS results with data derived from established online LIF instruments like the WIBS and UV-APS. In this context, we added the following sentence:

(P7, L223-227): *"Optimization of the thresholding strategy is still an on-going work, for example to investigate whether the often applied 3σ threshold used for the WIBS (e.g., Gabey et al., 2010) also works well with respect to the optical setup of the SIBS. For the assessment of the accuracy of measured fluorescence emissions from reference compounds, a threshold of 1σ was used here."*

[R2.4] L218: This looks like it may be due to coincidence errors arising from multiple particles being present in the sample volume causing odd scattering behaviour. This is a known problem when sampling high concentrations with forward scattering cloud probes, resulting in spectral broadening (e.g., Cooper, 1988).

[A2.4] Thanks a lot for this hint. The stated reference might indeed be an explanation for the effects we have observed for asymmetry factor measurements with the SIBS. The following sentence was added:

(P8, L235-237): "*However, one explanation could be optical coincidences caused by high particle concentrations, resulting in multiple particles being simultaneously present within the scattering volume, as reported by Cooper (1988) using forward-scattering signatures of cloud probes.*"

[R2.5] L435: This is a very interesting point that is raised here about the range irradiance imbalance between xenon lamps. This confirms some of my suspicious about the utility of presenting ABC analysis in general terms without appropriate caveats or a calibration standard and I think this is worth further comment. The Hernandez et al. (2016) work showed some of the results of the issues mentioned here when they compared two WIBS-4As where there were some significantly different classifications between the two units for the same test particle. They speculated that the difference between units was due to detector gain but your results suggest that xenon intensity may significantly contribute towards the observed differences. As a follow on comment this also shows the need for a common calibration reference standard to be adopted by the UV-LIF community (e.g., Robinson et al., 2017). This potentially raises a significant challenge for UV-LIF spectrometers with increased spectral resolution as I don't know if there is likely to be a single fluorophore that will adequately cover the whole spectral range?

[A2.5] This observation is indeed a critical point when it comes to the interpretation of fluorescence data derived from online LIF instruments using similar optical setups. Observed differences, between similar instruments as stated in, e.g., Hernandez et al. (2016), are most likely based on the complex interaction of multiple technical components, batch-to-batch variability etc. However, if prospective experiments verify a general imbalance between xenon sources / optical filtering for the WIBS and SIBS, this issue might turn out to be a major contributor to this topic. We agree with Dr. Crawford that it is absolutely necessary to adopt a calibration standard within the online bioaerosol community. However, to the best of our knowledge, there is currently no compound available that fulfills the requirements (e.g., stability, repeatability, broad spectral range etc.) for being a standard calibrant for multi-channel, multi-excitation LIF-instruments.

Within "5. Summary and conclusions", this existing text passage briefly discuss the data interpretation issue:

(P24, L799-805): "These observations are valid not only for the SIBS, but also for the WIBS-4A and WIBS-NEO and lead to important implications for interpretation of particle data. In particular, a particle that exhibits measurable fluorescence in WIBS channel FL1, but only weak fluorescence in channel FL3 could be assigned as an "A-type" particle in one instrument but an "AC-type" particle in an instrument with slightly stronger xenon 2 irradiance. These differences in classification can be extremely important to interpretation of ambient data (Perring et al., 2015; Savage et al., 2017)."

Additionally, we added the following sentence regarding instrument intercomparisons / calibrant standards:

(P24, L794-799): *"Additionally, alternating irradiance properties might significantly contribute to observed differences in performance of similar instrument types (e.g., Hernandez et al., 2016), expressly underlining the need for a fluorescence calibrant applicable across LIF-instruments (e.g., Robinson et al., 2017). Nevertheless, to the best of our knowledge, there is currently no standard reference available that fulfills the requirements to serve as a calibrant for multi-channel, multi-excitation LIF-instruments."*

[R2.6] L517: In my experience of calibrating forward scattering cloud probes it is often common to find a dip in sizing performance in the lower region of an instruments detection range due to Mie-

Lorenz resonances in the applied Mie curve exceeding the bin thresholds or the bin thresholds being relatively narrow. Mis-sizing can also be further exacerbated by the particles position in the sample area as recently demonstrated by Faber et al. (2018), however this is less likely to be an issue with SIBS/WIBS type instruments as the sample flow jet should be well constrained to the central sampling region. Given that the fit to the calibration has a slope of approximately 1 and a negligible intercept the assumed Mie curve appears to be adequate, however, should there routinely be a dip in the particle size distribution around this size this may explain why.

[A2.6] We considered this possibility, and almost added a comment to the discussion manuscript to this effect. Looking into the Mie curves in more detail, however, we did not find a solid evidence that may serve as an explanation for the effect observed in a size range between 0.6 – 0.8 µm. Because the idea was not strongly supported and to avoid inadvertently leading readers astray, we decided to leave the issue with unknown cause.

[R2.7] Fig. 7: Can you add to the caption what the red line represents. I assume it is the rebinned reference spectra as in Fig. 5.

[A2.7] True, red dashed lines show re-binned reference spectra as stated in Fig. 5 for **c** and **d**. The caption was modified for all corresponding figures (manuscript and supplement) as requested.

[R2.8] Fig. S10: This would be easier to interpret if the two plots were scaled over the same x-axis range.

[A2.8] Within the supplement manuscript, Fig. S10 was modified as requested.

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Spectral Intensity Bioaerosol Sensor (SIBS): An ~~new~~ Instrument for Spectrally Resolved Fluorescence Detection of Single Particles in Real-Time

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Keywords: ~~*SIBS*~~, WIBS, ~~*B*~~bioaerosols, ~~*Single Particle Fluorescence*~~, ~~*F*~~fluorescence ~~*S*~~spectroscopy, ~~*P*~~performance ~~*E*~~evaluation *and characterization*, ~~*P*~~polystyrene ~~*L*~~latex ~~*S*~~spheres, ~~*B*~~biofluorophores, *size and fluorescence calibration*

Abstract

Primary biological aerosol particles (PBAP) in the atmosphere are highly relevant for the Earth system, climate, and public health. The analysis of PBAP, however, remains challenging due to their high diversity and large spatiotemporal variability. For real-time PBAP analysis, light-induced fluorescence (LIF) instruments have been developed and widely used in laboratory and ambient studies. The interpretation of fluorescence data from these instruments, however, is often limited by a lack of spectroscopic information. This study introduces a*n* ~~new~~ instrument – the Spectral Intensity Bioaerosol Sensor (SIBS*, Droplet Measurement Technologies (DMT, Longmont, CO, USA))* – that resolves fluorescence spectra for single particles and, thus, promises to expand the scope of fluorescent PBAP quantification and classification.

The SIBS shares key design components with the latest versions of the Wideband Integrated Bioaerosol Sensor (WIBS) and the findings presented here are also relevant for the widely deployed WIBS-4A and WIBS-NEO as well as other LIF instruments. The key features of the SIBS and findings of this study can be summarized as follows:

- Particle sizing yields reproducible linear responses for particles in the range of 300 nm to 20 µm. The lower sizing limit is significantly smaller than for earlier commercial LIF instruments (e.g., WIBS-4A and the Ultraviolet Aerodynamic Particle Sizer (UV-APS)), expanding the analytical scope into the accumulation mode size range.
- Fluorescence spectra are recorded for two excitation wavelengths (λ_{ex} = 285 and 370 nm) and a wide range of emission wavelengths (λ_{mean} = 302 – 721 nm) with a resolution of 16 detection channels, which is higher than for most other commercially available LIF bioaerosol sensors.
- Fluorescence spectra obtained for 16 reference compounds confirm that the SIBS provides sufficient spectral resolution to distinguish major modes of molecular fluorescence. For example, the SIBS resolves the spectral difference between bacteriochlorophyll and chlorophyll *a*/*b*.
- A spectral correction of the instrument-specific detector response is essential to use the full fluorescence emission range.
- Asymmetry factor (AF) data were assessed and were found to provide only limited analytical information.
- In test measurements with ambient air, the SIBS worked reliably and yielded characteristically different spectra for single particles in the coarse mode with an overall fluorescent particle fraction of ~4 % (3σ threshold), which is consistent with earlier studies in comparable environments.

1. Introduction

Aerosol particles are omnipresent in the atmosphere, where they are involved in many environmental and biogeochemical processes (e.g., Baron & Willeke, 2001; Després et al., 2012; Fuzzi et al., 2006; Hinds, 1999; Pöschl, 2005; Pöschl & Shiraiwa, 2015). Primary biological aerosol particles (PBAP), also termed bioaerosols, represent a diverse group of airborne particles, consisting of whole or fragmented organisms including, e.g., bacteria, viruses, archaea, algae, and reproductive units (pollen and fungal spores), as well as decaying biomass (e.g., Deepak & Vali, 1991; Després et al., 2012; Fröhlich-Nowoisky et al., 2016; Jaenicke, 2005; Madelin, 1994; Pöschl, 2005) and can span sizes from few nanometers up to ~100 µm (Hinds, 1999; Schmauss and Wigand, 1929). The increasing awareness of the importance of PBAP regarding aerosol-cloud interactions, health aspects, and spread of organisms on local, continental or even intercontinental scales has led to a growing interest by scientific researchers and the public (e.g., Després et al., 2012; Fröhlich-Nowoisky et al., 2016; *Yao, 2018*).

Due to inherent limitations (e.g., poor time resolution and costly laboratory analyses) of traditional off-line techniques (e.g., light microscopy and cultivation-based methods) for PBAP quantification, several types of real-time techniques have been developed within the last several decades to provide higher time resolution and lower user costs (e.g., Caruana, 2011; Després et al., 2012; Fennelly et al., 2017; Ho, 2002; *Huffman and Santarpia, 2017*; Jonsson and Tjärnhage, 2014; Sodeau and O'Connor, 2016). One promising category of real-time instruments – meaning that particles are sampled and analyzed both instantly and autonomously – involves application of light- induced fluorescence (LIF). The main principle of this technique is the detection of intrinsic fluorescence from fluorophores ubiquitous in biological cells, such as those airborne within PBAP. These fluorophores include a long list of biological molecules such as aromatic amino acids (e.g., tryptophan and tyrosine), co-enzymes (e.g., reduced pyridine nucleotides (NAD(P)H)), flavin compounds (e.g., riboflavin), as well as biopolymers (e.g., cellulose and chitin) and chlorophyll (e.g., Hill et al., 2009; Li et al., 1991; Pan et al., 2010; Pöhlker et al., 2012, 2013). Detailed information of biological fluorophores can be found elsewhere (Pöhlker et al., 2012 and references therein).

Today, commercial on-line LIF instruments such as the Ultraviolet Aerodynamic Particle Sizer (UV-APS, TSI Inc. Shoreview, MN, USA) and the Wideband Integrated Bioaerosol Sensor (WIBS, developed ~~at~~ *by* the University of Hertfordshire, U.K. and currently licensed and manufactured by Droplet Measurement Technologies (DMT, Longmont, CO, USA)) are commonly applied for research purposes. Detailed descriptions of the UV-APS (e.g., Agranovski et al., 2003; Brosseau et al., 2000; Hairston et al., 1997) and the WIBS series (e.g., Foot et al., 2008; Kaye et al., 2000, 2005; Stanley et al., 2011) are given elsewhere. Concisely, the UV-APS uses an λ_{ex}= 355 nm laser excitation source and spans an emission range between λ_{em}= 420-575 nm. In contrast, the WIBS applies

two pulsed xenon flash lamps emitting at λ_{ex}= 280 and 370 nm, whereas fluorescence emission is detected in three detection channels, λ_{em}= 310 - 400 nm (at λ_{ex}= 280 nm) and λ_{em}= 420 - 650 nm (at λ_{ex}= 280 and 370 nm). Both instruments provide spectrally unresolved fluorescence information. The latest WIBS model is currently the WIBS-NEO, whose design is based on a WIBS-4A but with an extended particle size detection range between ~~~300~~ *~500* nm and ~~100~~ *30* µm (nominal). Both UV-APS and WIBS models have been examined in a variety of laboratory validations (e.g., Agranovski et al., 2003, 2004; Brosseau et al., 2000; Healy et al., 2012; Hernandez et al., 2016; Kanaani et al., 2007; O'Connor et al., 2013; Saari et al., 2013, 2014; Savage et al., 2017; Toprak & Schnaiter, 2013) and have been deployed to investigate both indoor and outdoor atmospheric aerosol via longer-term measurements (e.g., Bhangar et al., 2014; *Calvo et al., 2018*; Crawford et al., 2015b; Fernández-Rodríguez et al., 2018; Foot et al., 2008; Gabey et al., 2010, 2013; Gosselin et al., 2016; Healy et al., 2014; Huffman et al., 2010, 2012, 2013; *Ma et al., 2019*; Perring et al., 2015; Schumacher et al., 2013; Twohy et al., 2016; Ziemba et al., 2016).

Although LIF instruments do not offer the same *qualitative* ability to ~~*qualitatively*~~ identify sampled particles as, e.g., off-line microscopy, mass spectrometry, or culture-based methods, they provide size-resolved information as well as fast sampling and fine-scale temporal information for single particles not accessible with off-line techniques. Nevertheless, these instruments present significant challenges. For example, quantification of PBAP by LIF instruments is hindered by the fact that some biological materials reveal weak fluorescence characteristics that does not rise above detection thresholds (Huffman et al., 2012). In addition to this complication, the detection threshold is not a universally defined parameter and varies for each channel between different units of the same type of instruments (e.g., Hernandez et al., 2016; Savage et al., 2017). Furthermore, unambiguous spectroscopic characterization of bioparticles is fundamentally challenging, because fluorescence spectra of even individual molecules in condensed matter are relatively broad due to radiative decay pathways of excited electrons. Further, bioparticles are chemically complex, each comprised of a mixture of *at least* dozens of types of fluorophores that can each emit a unique emission spectrum that smears together into an even broader fluorescence spectrum from each particle (Hill et al., 2009, 2015; Pan, 2015). Another difficulty is that many non-biological particles, such as certain mineral dusts and polycyclic aromatic hydrocarbons (PAHs), may fluoresce, making it more difficult to distinguish patterns arising from biological particles (e.g., Pöhlker et al., 2012 and references therein; Savage et al., 2017). Lastly, most currently available commercial LIF instrumentation are limited to recording data in 1-3 spectrally integrated emission channels, which limits the interpretation of fluorescence information. Recent efforts to apply more complex clustering algorithms to the spectrally unresolved WIBS-type data are proving helpful at adding additional discrimination *(e.g., Crawford et al., 2015a; Robinson et al., 2013; Ruske et al., 2017; Savage & Huffman, 2018)*~~,~~*. For example, it was shown for*

a rural forest study in Colorado that a cluster derived using WIBS-3 data, assigned to fungal spores (Crawford et al., 2015a), correlated well with the mass concentration of molecular fungal tracers (e.g., arabitol and mannitol) measured with offline chemical techniques (Gosselin et al., 2016). In contrast, the clusters in the same study that were assigned to bacteria correlated only poorly with endotoxins, used as bacterial molecular tracers (Gosselin et al., 2016). This provides evidence of a limitation to using LIF instrumentation with low spectral resolution to separate or identify some PBAP types. Additionally, the bacterial cluster allocation might have also been hampered in that case by the minimum detectable particle size of the WIBS (~0.8 µm), resulting in a lower detection efficiency for bacteria. ~~but aerosol characterization using instrumentation with such low spectra resolution is likely to be fundamentally limited (e.g., Robinson et al., 2013; Ruske et al., 2017; Savage & Huffman, 2018)~~.

The evolution of LIF techniques over the last several decades has significantly expanded our knowledge on spatiotemporal patterns of PBAP abundance in the atmosphere. Nevertheless to further improve the applicability of LIF instrumentation to widespread PBAP detection, it is necessary both to design LIF instrumentation with adequate instrumental properties (e.g., high spectral resolution) and to standardize their operation by characterizing instruments thoroughly with known standards (Robinson et al., 2017). Working toward this goal, a number of LIF instruments *have been developed to analyze* ~~*that offer analysis of*~~ single bioparticles by collecting ~~*providing*~~ resolved fluorescence spectra ~~*have been developed*~~ (e.g., Hill et al., 1999; Pan et al., 2010, 2003; Pinnick et al., 2004; Ruske et al., 2017), however relatively little has been done to offer these commercially. Examples for commercially available instruments providing resolved fluorescence spectra are the PA-300 (λ_{ex}= 337 nm; λ_{em}= 390 – 600 nm, 32 fluorescence detection channels) (Crouzy et al., 2016; Kiselev et al., 2011, 2013) and the follow-up model Rapid-E (λ_{ex}= 337 nm; λ_{em}= 350 – 800 nm, 32 fluorescence detection channels) (http://www.plair.ch/), both manufactured by Plair SA, Geneva, Switzerland. ~~*Beside of resolved fluorescence detection*~~ *In addition to collecting resolved fluorescence spectra*, both instruments also provide measurements of the decay of fluorescence signals, also referred to as fluorescence lifetime.

Introduced here is a*n* ~~new~~ instrument for the detection and characterization of individual particles; the Spectral Intensity Bioaerosol Sensor (SIBS, Droplet Measurement Technologies). Technical properties of the instrument are described in detail and its performance is validated with sizing and fluorescence particle standards, as well as with *particles in* ambient air. Due to the dual excitation and spectrally resolved fluorescence in combination with a broad size detection range, the SIBS has the potential to increase the selectivity of fluorescent biological and non-biological particle detection and discrimination. Because the SIBS uses a comparable optical system as the WIBS-4A and WIBS-NEO, technical details presented here are broadly important to a growing community of scientists

investigating both indoor and outdoor aerosol. Insights and data presented will thus contribute to ongoing discussions within the community of LIF users and will also stimulate discussions about needs for future instrument improvements.

2. Materials and methods

2.1 Chemicals and materials

Supplemental table S1 summarizes 19 polystyrene latex spheres (PSLs, 5 doped with fluorescent dye) and 6 polystyrene divinylbenzene (PS-DVB) particles, which were purchased from Thermo Fisher (Waltham, MA, USA), Bangs Laboratories Inc. (Fishers, IN, USA), Duke Scientific Corp. (Palo Alto, CA, USA), and Polysciences Inc. (Warrington, PA, USA). A detailed study regarding steady-state fluorescence properties of PSLs and PS-DVB particles used within this study can be found in Könemann et al. (2018). Additionally, we analyzed particles comprised separately of seven pure biofluorophores (tyrosine, tryptophan, NAD, riboflavin, chlorophyll *a* and *b*, and bacteriochlorophyll) (Table S2) as well as one microorganism (*Saccharomyces cerevisiae;* baker's yeast, bought at a local supermarket). Table S2 also includes reference particles used for asymmetry measurements, namely iron oxide (Fe_3O_4), carbon nanotubes, and ammonium sulfate. Ultrapure water (MilliQ, 18 MΩ) and ≥ 99.8 % ethanol (CAS Nr. 64-17-5, Carl Roth GmbH und Co. KG, Karlsruhe, Germany) were used as solvents.

2.2 Aerosolization of reference particles

PSLs were aerosolized from aqueous suspensions with a portable aerosol generator (AG-100; DMT). For both fluorescent and non-fluorescent PSLs, one drop of the suspension (or alternatively three drops for 3 and 4 µm PSLs) was diluted into 10 ml ultrapure water in plastic medical nebulizers (Allied Healthcare, St. Louis, MO, USA). The majority of water vapor from the aerosolization process condenses inside the mixing chamber (~570 cm^3) of the aerosol generator. By using a temperature and relative humidity (RH) sensor (MSR 145 data logger, MSR Electronics GmbH, Seuzach, Switzerland) monitoring the flow directly after the aerosol generator we measured RH values of ~33% (sample flow: 1.4 l/min, dilution: 5 l/min), ~39% (sample flow: 1.4 l/min, dilution: 4 l/min), and ~54% (sample flow: 2.3 l/min, dilution: 2 l/min). Because of the low RH measured, we did not use additional drying (e.g., diffusion dryer) to decrease humidity in the sample flow. Hence, the outlet of the aerosol generator was directly connected to the SIBS inlet with ~30 cm of conductive tubing ($^1/_4$ inch). PSLs were measured for 1 min. Non-fluorescent 4.52 µm PSLs were measured for 2 min, because of the low number concentrations due to poor aerosolization efficiency and gravitational settling of larger particle sizes.

S. cerevisiae was analyzed using a method similar to the one stated above, with the exceptions that the suspension was prepared with a spatula tip of material mixed into ultrapure water and that a diffusion dryer (20 cm, 200 g silica) was added to remove excess water vapor. *S. cerevisiae* was measured for 5 min. Chlorophyll *a*, *b*, and bacteriochlorophyll samples were diluted in 10 ml ethanol. Between each measurement, the setup was cleaned by aerosolizing ultrapure water for 5 min.

PS-DVB particles and biofluorophores (Table S1 and S2) were aerosolized in a dry state. For this purpose, air at a flowrate of ~0.6 l/min was sent through a HEPA filter into a 10 ml glass vial. A small amount of each solid powder sample (~1 g) was placed inside the vial and entrained into the particle-free airstream. Additionally, the sample was physically agitated by tapping the vial. The outlet was connected with ~20 cm conductive tubing into the inlet of the SIBS. The tubing and glass vial were cleaned after each measurement to prevent particle contaminations from previous measurements. Each powder was sampled until cumulative number concentrations > 5000 particles were reached.

In contrast to the monodisperse and spherical PSL standards, the biofluorophore aerosolization process provided a polydisperse and morphologically heterogeneous particle distribution with significant particle fractions at sizes < 1 µm. Therefore, we only used particles in a size range between 1 and 2 µm with sufficient fluorescence intensity values for subsequent data analysis. The only exceptions are the chlorophyll types, where a size range between 0.5 and 2 µm (chlorophyll *a* and *b*) and 0.5 and 1 µm (bacteriochlorophyll) were used due to a less efficient particle aerosolization.

The fluorescent background of the SIBS was measured daily by firing the xenon lamps into the optical chamber in the absence of particles (forced trigger mode). In this case, the diaphragm pump was turned off and the inlet blocked to prevent particles reaching the optical chamber. One forced trigger mode was performed per day with 100 xenon shots per min over a duration of 5 min. The *average* background signal (+ 1σ standard deviation (SD)) was subtracted from derived fluorescence emission of each sample. Additionally, the background signal was reviewed periodically between each biofluorophore measurement to verify that, e.g., optical components ~~*are*~~ *were* not coated with residues from previous measurements. ~~*A*~~ No significant change*s* in background signal *were* ~~*was not*~~ observed between individual measurements. *Optimization of the thresholding strategy is still an ongoing work, for example to investigate whether the often applied 3σ threshold used for the WIBS (e.g., Gabey et al., 2010) also works well with respect to the optical setup of the SIBS. For the assessment of the accuracy of measured fluorescence emissions from reference compounds, a threshold of 1σ was used here.*

For particle asymmetry measurements, iron (II, III) oxide (Fe_3O_4), carbon nanotubes, and ammonium sulfate were aerosolized in dry state, and 2 µm non-fluorescent PSLs and ultrapure water were aerosolized with the aerosol generator method outlined above with SIBS integration times of 3 min

in all cases. Due to the broad distribution of asymmetry factor (AF) values for particles below 1 µm, only the size fraction ≥ 1 µm was used for subsequent analyses. Furthermore, we observed that AF bins between 0 and 1, and AF bin 100 tend to produce increased signal responses, especially for high particle concentrations, for which they were discarded within the analyses. The origin of this effect is unknown~~., but most likely related to detector noise.~~ *However, one explanation could be optical coincidences caused by high particle concentrations, resulting in multiple particles being simultaneously present within the scattering volume, as reported by Cooper (1988) using forward-scattering signatures of cloud probes..*

For collection of particles for microscopy measurements, the sample flow was bypassed and led through a custom-made particle impactor, which was connected to a mass flow controller (D-6321-DR, Bronkhorst High-Tech B.V., Ruurlo, Netherlands) and a membrane pump (N816.1.2KN.18, KNF, Freiburg, Germany). Particles were collected out of the sample flow onto glass cover slips (15 mm diameter) at a flow rate of 2 l/min over a duration of 1 min.

2.3 Reference fluorescence spectra

A Dual-FL fluorescence spectrometer (Horiba Instruments Incorporated, Kyoto, Japan) was used as an offline reference instrument to validate the SIBS spectra. Aqualog V3.6 (Horiba) software was used for data acquisition. The spectrometer was manufacturer-calibrated with NIST Fluorescence Standard Reference Materials (SRMs 2940, 2941, 2942, and 2943). Aforementioned standard fluorophores were analyzed using the SIBS excitation wavelengths at λ_{ex} = 285 and 370 nm. The Dual-FL[1] spectrometer uses a xenon arc lamp as excitation source and a CCD (charge-coupled device) as emission detector, capable of detecting fluorescence emission between 250 and 800 nm. Unless otherwise stated, a low detector gain setting (2.25 e^- per count) and an emission resolution of 0.58 nm was used for all measurements with the Dual-FL. Subsequently, we use the term "reference spectra" for all measurements performed with the Dual-FL. In total, 100 individual spectra were recorded for each sample and averaged spectra were analyzed in Igor Pro (Wavemetrics, Lake Oswego, Oregon USA). Background measurements (solvent[2] in the absence of particles) were taken under the same conditions as for sample measurements and subtracted from the emission signal. For direct comparison to spectra recorded by the SIBS, reference spectra were re-binned by taking the sum of the fluorescence intensity within the spectral bin width of each SIBS detection channel (Table 1).

For PSL measurements, 1.5 µl of each PSL stock solution was diluted in 3.5 ml ultrapure water in a 10 x 10 x 40 mm UV quartz cuvette (Hellma Analytics, Müllheim, Germany) and constantly stirred with a magnetic stirrer to avoid particle sedimentation during measurements. Chlorophyll *a*

[1] Technical information taken from Dual-FL operation manual, rev. A, 30 NOV 2012; Horiba.

[2] ~~*Note that ≥ 99.8 % ethanol was used as solvent for chlorophyll a, b, and bacteriochlorophyll instead of ultrapure water.*~~

and *b* and bacteriochlorophyll were handled equally, however concentrations were individually adjusted to prevent the detector from being saturated and to avoid self-quenching or inner filter effects (Sinski and Exner, 2007). Concentrations were used as follows: chlorophyll *a*: 300 nmol/l, chlorophyll *b*: 1 µmol/l, and bacteriochlorophyll: 3 µmol/l. PSLs, chlorophyll *b*, and bacteriochlorophyll measurements were performed with an integration time of 2 s. For chlorophyll *a* an integration time of 1 s was used.

All other biofluorophores, *S. cerevisiae*, and PS-DVB particles were measured in dry state using a front surface accessory (Horiba). The sample was placed into the surface holder and covered with a synthetic fused silica window. To limit detector saturation from more highly fluorescent particle types, the surface holder was placed at a 70° angle to the fluorescence detector for NAD and riboflavin, 75° for tyrosine, 80° for *S. cerevisiae*, and 85° for tryptophan and PS-DVB particles and subsequently excited at λ_{ex}=285 and 370 nm. Emission resolution and detector gain settings were used as for measurements of samples in solution, except for an integration time of 1 s for all dry samples. Background measurements were performed as described above and subtracted from each sample. Excitation-emission matrices (EEMs) were measured with the same samples as for single wavelength measurements. EEMs were recorded at excitation wavelengths between λ_{ex} = 240 and 800 nm (1 nm increments) and an emission range between λ_{em} = 247 and 829 nm (0.58 nm increments). Exposure times of 1 s were used, except for 2 µm green, 3 µm non-fluorescent PSLs (2 s), and NAD (0.5 s). EEMs were analyzed using Igor Pro.

2.4 Calibration lamps and spectral correction

The relative responsivity of a fluorescence detector can vary substantially across its emission range and, therefore, must be spectrally corrected as a function of emission wavelength (e.g., DeRose, 2007; Lakowicz, 2004). For spectral correction it was important to choose: (i) light sources covering the full spectral emission range of the SIBS, with temporal stability on the timescale of many months and (ii) a calibrated and independent spectrometer to serve as spectral reference.

A deuterium-halogen lamp (DH-Mini; Ocean Optics, Largo, FL, USA) and a halogen projector lamp (EHJ 24 V, 250 W; Ushio Inc., Tokyo, Japan) were used as calibration light sources. Both lamps were connected to a 50 cm optical fiber (FT030, Thorlabs, Newton, NJ, USA) and vertically fixed inside the optical chamber of the Dual-FL spectrometer. An aluminum mirror was attached to the end fitting of the optical fiber, reflecting light in a 90° angle into the detector opening. The projector halogen lamp was allowed to warm up for 30 s before each measurement. For all power levels (100, 150, 200, and 250 W), an integration time of 3 s was used. The DH-Mini was operational for 30 min before each measurement. Settings were used as for the projector halogen lamp, however, due to the low emission a high detector gain setting (9 e^- per count) was used with an integration time

of 25 s. As described in Sect. 2.3, 100 single measurements were taken and averaged (Fig. S1). For the SIBS, both light sources were measured in the same way as for the reference spectra. Measurements were performed with a detector amplification at 610 V (see Sect. 4.2). Background measurements were taken as described in Sect. 2.2. Projector halogen lamp spectra (at all power levels) were recorded for 3 min, the DH-Mini, due to its low emission intensity, for a duration of 5 min.

For the halogen projector lamp, averaged intensity values in each spectral bin were acquired at each power level (150, 200, and 250 W). Spectra measured at 100 W were discarded due to the low and unstable emission at wavelengths shorter than ~500 nm (Fig. S1). Reference spectra and spectra recorded by the SIBS were normalized onto the SIBS detection channel 9 (λ_{mean} = 528.0 nm), which is, theoretically, the detection channel with the highest responsivity (see Sect. 4.3). The individual spectral correction factors were calculated by dividing the reference spectra by the spectra derived from the SIBS. The final correction factors are a combination ~~out~~ of both light sources where the detection channels 1-5 (λ_{mean} = 302.2 – 415.6 nm) include the correction factors for the DH-Mini and the detection channels 6-16 (λ_{mean} = 443.8 – 721.1 nm) the correction factors for the halogen projector lamp. At the intersection between channel 5 and 6, both corrections (DH-Mini, halogen) are in good agreement ($\Delta_{correction}$ = 0.6 in channel 6). For all particle measurements described in the following sections, the background signal and raw sample spectra recorded by the SIBS were multiplied by those correction factors.

2.5 Microscopy of selected reference particles

Bright field microscopy was conducted using an Eclipse Ti2 (Nikon, Tokyo, Japan) with a 60x immersion oil objective lens and an additional optical zoom factor of 1.5, resulting in a 90x magnification. Glass cover slips, used as collection substrates in the particle impactor (Sect. 2.2), were put onto a specimen holder and fixed with tape. Images were recorded using a DS Qi2 monochrome microscope camera with 16.25 megapixels and z-stacks of related images were created using the software NIS-Elements AR (both Nikon).

2.6 Ambient measurement setup and data analysis

The SIBS was operated between the 5th of April to the 7th of May 2018 *from a fourth floor roof laboratory* ~~*on the roof*~~ ~~*(fourth floor inside a roof laboratory) of*~~ at the Max Planck Institute for Chemistry in Mainz, Germany (49°59′28.2”N, 8°13′44.5”E) similar to measurements as described in Huffman et al. (2010) using a UV-APS. The period between the 12th and 18th of April 2018 is described here to highlight the capability of the SIBS to monitor ambient aerosol. Beside of the SIBS,

four additional instruments (data not shown within this study) were connected with ~20 cm conductive tubing ($^1/_4$ inch) to a sample airflow splitter (Grimm Aerosol Technik GmbH & Co. KG, Ainring, Germany). The splitter was connected to 1.5 m conductive tubing ($^5/_8$ inch), bent out of the window, and connected to 2.4 m stainless steel tubing ($^5/_8$ inch, Dockweiler AG, Neustadt-Glewe, Germany) vertically installed. Between a TSP head (total suspended particles, custom-made) and the stainless steel tubing, a diffusion dryer (1 m, 1 kg silica) was installed. Silica was exchanged every third to fourth day and periodic forced trigger measurements were performed *daily*. The total flow was ~8.4 l/min.

For measurements presented here, ~~only~~ particles were *only* included if they showed fluorescence emission in at least two consecutive spectral channels. This filter was applied to limit noise introduced from measurement artifacts from a variety of sources and will need to be investigated in more detail. The conservative analysis approach here suggests that the values reported are likely to be a lower limit for fluorescent particle number and fraction. The observations are in line with previous measurements, however, giving general support that the SIBS measurements are reasonable. Note that the maximum repetition rate of the xenon lamps is 125 Hz, corresponding to maximum concentrations of 20 particles per cm^{-3} (see Sect. 3.3). Because ~50% of the total coarse particle number were excited by xenon 1 and xenon 2, the fluorescent particle concentrations and fluorescent fractions are corrected accordingly.

3. Design and components of the SIBS

The SIBS is based on the general optical design of the WIBS-4A (e.g., Foot et al., 2008; Healy et al., 2012; Hernandez et al., 2016; Kaye et al., 2005; Perring et al., 2015; Robinson et al., 2017; Savage et al., 2017; Stanley et al., 2011) with improvements based on a lower particle sizing limit, resolved fluorescence detection, and a broader emission range. The instrument provides information about size, particle asymmetry, and fluorescence properties for individual particles in real-time. The excitation wavelengths are optimized for the detection of the biological fluorophores tryptophan, NAD(P)H, and riboflavin. However, other fluorophores in PBAP will certainly fluoresce at these excitation wavelengths as many of them cluster in two spectral fluorescence "hotspots" as summarized in Pöhlker et al. (2012 and references therein) and as shown for WIBS-4A measurements by Savage et al. (2017). Figure 1 shows an overview of excitation wavelengths and emission ranges of the UV-APS, WIBS-4A, WIBS-NEO, and SIBS for bioaerosol detection in relation to the spectral location of selected biofluorophores, such as tyrosine, tryptophan, NAD(P)H, riboflavin, and chlorophyll *b*. At $\lambda_{ex} = 285$ nm, the SIBS excites fluorophores in the "protein hotspot", at $\lambda_{ex} = 370$ nm fluorophores in the "flavin/coenzyme hotspot" (Pöhlker et al., 2012). In contrast to the UV-APS, the SIBS is able to detect fluorescence signals from chlorophyll due to the extended upper spectral range

of detection (up to λ_{em} = 721 nm). Both the WIBS-4A and WIBS-NEO cover the spectral emission range for chlorophyll *b*, however, cannot provide resolved spectral information to separate it from other fluorophores. Table 2 summarizes and compares parameters and technical components of the SIBS, WIBS-4A, and WIBS-NEO. The individual components are described in detail in the subsequent sections.

To avoid potential misunderstanding, it is important to note that the SIBS described in this study is not related to spark-induced breakdown spectroscopy instrumentation, which uses the same acronym (e.g., Bauer & Sonnenfroh, 2009; Hunter et al., 2000; Khalaji et al., 2012; Schmidt & Bauer, 2010). The DMT SIBS discussed here was recently used as part of a study investigating aerosols in several ambient outdoor environments (Nasir et al., 2018), but the study here is the first to discuss important technical details of the instrument design and operation.

3.1 Aerosol inlet and flow diagram

The design for the aerosol inlet of the SIBS is identical to the inlet of the WIBS-4A and WIBS-NEO. A detailed flow diagram is shown in Figure S2. Aerosol is drawn in via an internal pump as laminar air flow through a tapered delivery nozzle (Fig. S2a) where sheath (~2.2 l/min) and sample flow (~0.3 l/min) are separated.

3.2 Size and shape analysis

After passing the delivery nozzle, entrained particles traverses a 55 mW continuous-wave diode laser at λ = 785 nm (#2 in Fig. 2 and position #1 in Fig. S3). Unlike in the WIBS-4A and WIBS-NEO (635 nm diode laser), the triggering laser in the SIBS is in the near-infrared (IR) region (> 700 nm) and, therefore, outside the detectable emission range of the 16-channel photomultiplier tube (PMT) to avoid scattered light from the particle trigger laser being detected (see Fig. 1). The side and forward scattered light is collected and used for subsequent measurements. Side scattered light is collected by two concave mirrors, which are directed at 90° from the laser beam axis, and reflect the collected light onto a dichroic beam splitter (#7 in Fig. 2). A PMT (H10720-20, Hamamatsu Photonics K.K., Japan) converts incoming light signals into electrical pulses, which are used for particle triggering and sizing (#6 in Fig. 2). For the determination of the optical particle size, the SIBS uses a calculated calibration curve according to the Lorenz-Mie Theory, assuming spherical and monodisperse PSLs with a refractive index of 1.59 (Brandrup et al., 1989; Lorenz, 1890; Mie, 1908). Compared to aerodynamic sizing, which depends on particle morphology and density (e.g., Reid et al., 2003; Reponen et al., 2001), the calculated optical diameter can vary significantly if the assumption of sphericity is

not fulfilled. In contrast, optical sizing is not as affected by differences in material density. The instrument operator must thus be aware of uncertainties in measured particle size due to, e.g., particle morphology, spatial orientation of a particle when traversing the trigger laser or changing refractive indices. In contrast to the WIBS-4A, the SIBS and WIBS-NEO detect ~~a~~ *the full* range of particle sizes ~~between~~ *(SIBS:* ~0.3 and 100 µm (nominal)*; WIBS-NEO: ~0.5 and 30 µm (nominal))*~~,~~ ~~achievable~~ by using one PMT gain setting instead of switching between a "Low Gain" and "High Gain" setting. Physical and technical details of this Gain-switching method are patent pending and are not publicly available.

The forward-scattered light is measured by a quadrant PMT (#5 in Fig. 2) to detect the scatter asymmetry for each particle (Kaye et al., 1991, 1996). A OG-515 long pass filter (Schott AG, Mainz, Germany) prevents incoming light from the xenon flash lamps in a spectral range below 515 ± 6 nm from reaching the Quadrant PMT. To calculate the AF, the root-mean-square variations for each quadrant of the PMT of the forward-scattered light intensities are used (Gabey et al., 2010). The AF broadly relates whether a particle is more spherical or fibril. Theoretically, for a perfectly spherical particle, the AF would be 0, whereas an elongated particle would correspond to an AF of 100 (Kaye et al., 1991). However, due to electrical and optical noise of the Quadrant PMT, the AF value of a sphere is usually between ca. 2 and 6 (according to WIBS-4A service manual (DOC-0345 Rev A)). Because the AF value depends on physical properties of optical components, the baseline for spherical particles may shift even within identical instruments (Savage et al., 2017). For example, the study by Toprak & Schnaiter (2013) reported an average AF value for spherical particles of 8 using a WIBS-4A. In contrast, AF values shown by Foot et al. (2008) were, on average, below ~5 for spherical particles measured with a WIBS-2s prototype.

3.3 Fluorescence excitation

Two xenon flash lamps (L9455-41, Hamamatsu) (#3 and #4 in Fig. 2) are used to induce fluorescence. They emit light pulses, which exhibit a broad excitation wavelength range of 185 to 2000 nm. The light is optically filtered to obtain a *relatively monochromatic* ~~*defined*~~ excitation wavelength. Further information about spectral properties of the xenon flash lamps can be found elsewhere (Specification sheet TLSZ1006E04, Hamamatsu, May 2015). Figure 3 displays relevant optical properties of the lamps and filters used within the SIBS, WIBS-4A, and WIBS-NEO. For the SIBS, a BrightLine® FF01-285/14-25 (Semrock Inc., Rochester, NY, USA) single-band bandpass filter is used with λ_{mean} = 285 nm and an effective excitation band[3] of 14 nm width is used for xenon 1. For xenon 2,

[3] The effective excitation band is defined as "guaranteed minimum bandwidth" (GMBW), describing the spectral region a bandpass filter transmits light relative from the mean wavelength. For example, a GMBW of 14 nm means that light is transmitted in a 7 nm spectral range above and below the mean wavelength.

the single-band bandpass filter BrightLine® FF01-370/36-25 (Semrock) is used with λ_{mean} = 370 nm and with an effective excitation band of 36 nm width. The only difference between all three instruments is that the WIBS-4A and WIBS-NEO use a different single-band bandpass filter for xenon 1 (Semrock, BrightLine® FF01-280/20-25; λ_{mean} = 280 nm; effective excitation band of 20 nm). The excitation light beam *for all three instruments* is focused on the sample flow within the optical cavity, resulting in a rectangular beam shape of ~5 mm by 2 mm. Xenon 1 is triggered when particles pass position 2 in Figure S3 and approximately 10 µs later xenon 2 is triggered as the particles move further to position 3 in Figure S3. After firing, the flash lamps need ~5 ms to recharge. During the recharge period, particles are counted and sized but no fluorescence information is recorded. The maximum repetition rate of the xenon lamps yields a measurable particle number concentration of ~2 x 10^4 l^{-1} (corresponding to 20 cm^{-3}).

Irradiance values from light sources becomes a crucial factor when interpreting derived fluorescence data of LIF instruments because the fluorescence intensity is directly proportional to the intensity of incident radiant power, described by the relationship:

$$F = \phi I_0(1-e^{-\varepsilon bc}) \quad (1)$$

ϕ: quantum efficiency, I_0: intensity of incident light, ε: molar absorptivity, *b*: path length (cell), *c*: molar concentration (Guilbault, 1990).

To measure the irradiance of each xenon lamp after optical filtering, we used a thermal power head (S425C, Thorlabs), which was placed at a distance of 11.3 cm (focus length from xenon arc bow to sample flow intersection) from the xenon lamp measuring over a duration of 1 min at 10 xenon shots per s. By measuring new xenon lamps, we observed an average irradiance of 14.8 mW/cm^2 for xenon 1 and 9.6 mW/cm^2 for xenon 2, corresponding to ~154 % higher irradiance (spectrally integrated) from xenon 1. A second set of lamps, used intermittently for three years including several months of continuous ambient measurements and a lab study with high particle concentrations, exhibited average irradiance values of 10.8 mW/cm^2 (1σ SD 1.8 mW/cm^2) for xenon 1 and 4.9 mW/cm^2 (1σ SD 1.9 mW/cm^2) for xenon 2, corresponding to ~220 % higher irradiance from xenon 1. Comparing the nominal, transmission-corrected irradiance data from the two xenon lamps provided by the lamp supplier (Fig. 3a and 3b, red dashed lines), an irradiance imbalance between xenon 1 and xenon 2 can be assumed for all three LIF instruments discussed here (SIBS, WIBS-4A, and WIBS-N~~eo~~*EO*).

Results shown here are comparable to multiple WIBS studies (e.g., Hernandez et al., 2016; Perring et al., 2015; Savage et al., 2017), where fluorescence emission intensities at λ_{ex} = 280 nm (xenon 1) also show a tendency to be higher than those at λ_{ex} = 370 nm (xenon 2).

3.4 Spectrally resolved fluorescence detection

Fluorescence emission from excited particles is collected by two parabolic mirrors in the optical cavity and delivered onto a custom-made dichroic beam splitter (Semrock, #7 in Fig. 2). The beam splitter allows transmission of incoming light between ~300 and 710 nm, with an average transmission efficiency of 96%. At wavelengths shorter than 300 nm, the transmission decreases rapidly to < 20% at 275 nm. At the upper detection end of the SIBS (λ_{mean} = 721 nm), the transmission efficiency decreases to ~89%. The scattering light from the diode laser is reflected at a 90° angle onto the PMT used for particle detection and sizing. At the excitation wavelength of 785 nm, the reflection efficiency is stated at ~95% (Fig. S4).

After passing the dichroic beamsplitter, the photons are led into a grating polychromator (A 10766, Hamamatsu) (#8 in Fig. 2). A custom-made transmission grating (Hamamatsu) is used to diffract incoming light within a nominal spectral range between 290.8 – 732.0 nm. In case of the SIBS, a grating with 300 g/mm groove density and 400 nm blaze wavelength is used, resulting in a nominal spectral width of 441.2 nm and a resolution of 28.03 nm/mm. After passing the transmission grating, the diffracted light hits a 16-channel linear array multi-anode PMT (H12310-40, Hamamatsu) (#9 in Fig. 2) with defined mean wavelengths for each channel as shown in Table 1.

For each single particle detected, two spectra are recorded, at λ_{ex} = 285 and 370 nm. The detectable band range of the PMT overlaps the excitation wavelength of xenon 2. Therefore, a notch optical filter (Semrock) is placed between the optical chamber and the grating polychromator to prevent the detector from being saturated. Incoming light at wavelengths shorter than 300 nm and from 362 to 377 nm is blocked from reaching the PMT resulting in a reduced spectral bin width for detection channels 1, 3 and 4. The first three detection channels are omitted because their mean wavelengths are below λ_{ex} = 370 nm (see also Fig. 1). Accordingly, the emission spectra for xenon 2 excitation begin at channel 4 (λ_{mean} = 387.3 nm).

Technical data (xenon flash lamps, filters, dichroic beam splitter, PMT responsivity, and transmission grating) described in the previous sections (3.3 and 3.4) were provided by Hamamatsu and Semrock. Note that transmission/reflection efficiencies of the dichroic beam splitter, cathode radiant sensitivity of the PMT, and diffraction efficiency data are modeled. Thus, individual components may differ slightly from modeled values, even within the same production batch. Neither company assumes data accuracy or provides warranty, either expressed or implied.

The SIBS was originally designed and marketed to record time- and spectrally-resolved fluorescence lifetimes at two excitation wavelengths. The fluorescence lifetime of most biofluorophores, serving as targets for bioaerosol detection, are usually below 10 ns (e.g., Chorvat & Chorvatova, 2009; Herbrich, et al., 2012; O'Connor et al., 2014; Richards-Kortum & Sevick-Muraca, 1996). However, by choosing xenon lamps as excitation source, recording relevant fluorescence lifetimes

in this ns range is hampered by the relatively long decay time of the xenon lamp excitation pulse (~1.5 µs). In principle, fluorescence lifetime measurements would be possible if the xenon lamps were replaced by appropriate laser excitation sources in the SIBS optical design.

3.5 Software components and data output

The SIBS uses an internal computer (#10 in Fig. 2) with embedded LabView-based data acquisition software allowing the user to control functions in real time and change multiple measurement parameters. As an example, the "Single Particle" tab out of the SIBS interface is shown in Figure S5. Here, the user can define, e.g., the sizing limits of the SIBS (upper and lower threshold) and the minimum size of a particle being excited by the xenon flash lamps. Furthermore, forced trigger measurements can be performed while on this particular tab. Subsequently, the term "forced trigger measurement" will be replaced by "background signal measurement". A local Wi-Fi network is installed so that the SIBS can be monitored and controlled remotely. A removable hard drive is used for data storage. Data is stored in a HDF5 format to minimize storage space and optimize data write speed. Resulting raw data are processed in Igor Pro. As an example: by using a minimum sizing threshold of 500 nm, the SIBS data output per day, operating in a relatively clean environment (~40 particles per cm^{-3}), can span several hundreds of MB. In contrast, the data output can increase up to ~3 GB daily in polluted areas (~680 particles per cm^{-3}). By lowering the minimum sizing threshold to 300 nm, the data volume can exceed 10 GB per day when sampling in a moderately polluted environment (~180 particles per cm^{-3}).

4. Results and data validation

4.1 Validation of SIBS sizing

To validate the optical sizing of the SIBS, twenty particle size standards were analyzed, covering a broad size range from 0.3 to 20 µm in particle diameter. Overall, the particle size measurements from the SIBS (optical diameter) show good agreement with the corresponding measurements of physical diameter reported by PSL and PS-DVB manufacturers (Fig. 4). For the SIBS ~~and WIBS-NEO~~, the manufacturer states a nominal minimum size detection threshold of 300 nm. Figure 4 shows that a linear response between optical particle size and physical particle size extends *down* to at least 300 nm. Smaller particles were not investigated. The upper size detection threshold is reported by the manufacturer to be nominally 100 µm. However, the upper limit was not investigated here due to the difficulty in aerosolizing particles larger than this. In most field applications, the upper particle size cut is often far below this value due to unavoidable sedimentation losses of large particles in the inlet

system (e.g., Moran-Zuloaga et al., 2018.; Von der Weiden et al., 2009). Note that the size distributions of physical diameter for PS-DVB standards are broader compared to the PSL standards, as reported by the manufacturer (Table S1). This also translates to broader distributions of optical diameter measured by the SIBS *for PS-DVB* than for ~~*the*~~ PSL particles. The 0.356 µm PSL sample was an outlier with respect to the overall trend, showing an optical diameter of 0.54 µm. We suspect that this deviation between physical and optical size can be explained by a poor quality of this particular PSL sample lot rather than an instrumental issue, and so it was not included in the calculation of the trend line (Fig. 4). Furthermore, the SIBS was shown to *slightly* undersize the PSLs between 0.6 and 0.8 µm, however, the overall trend exhibits a coefficient of determination of $r^2 > 0.998$.

As mentioned in Sect. 3.2, an important point regarding the SIBS and WIBS-NEO is that the size calibration within the unit cannot be changed by the user, meaning that the PMT output voltages are transformed directly to outputted physical diameter within the internal computer using a proprietary calculation. It is still important, however, for the user to perform sizing calibration checks frequently to verify and potentially post-correct particle sizing of all particle sizing instruments, including the SIBS and WIBS-NEO.

4.2 Amplification of fluorescence detector

As with all optical detection techniques, adequate understanding of detection thresholds is an essential aspect of instrument characterization and use (e.g., Jeys et al., 2007; Savage et al., 2017). Application of appropriate voltage gain settings must be applied to the physical detection process so as not to lose information about particles that cannot be recovered by post-processing of data. Yet particles in the natural atmosphere exhibit an extremely broad range of fluorescence intensities (many orders of magnitude), arising from the breadth of quantum yields for fluorophores occurring in aerosols and from the steep increase of fluorescence emission intensity with particle size (2^{nd} to 3^{rd} power) (e.g., Hill et al., 2015; Könemann et al., 2018; Sivaprakasam et al., 2011; Swanson & Huffman, 2018). This range of fluorescence properties is generally broader than the dynamic range of any single instrument, and so a UV-LIF instrument can be operated, e.g., to either: (i) apply a higher detector gain to allow high sensitivity toward detecting weakly fluorescing particles, often from rather small particles (< 1 µm), at the risk of losing fluorescence information for large or strongly fluorescent particles due to detector saturation or (ii) apply a lower detector gain to preferentially detect a wide range of more highly fluorescent particles, but at the risk of not detecting weakly fluorescent or small particles.

Amplification voltage of the 16-channel PMT used in the SIBS can be adjusted between 500 and 1200 V. Each of the 16 detection channels can also be individually adjusted using digital gain settings within the SIBS acquisition software. This channel-specific gain does not affect the amplification

process (e.g., the dynode cascade), but rather modifies the output signal of single detection channels digitally. The digital gain is applied only after the signal collection process, and so cannot compensate for a signal that is below the noise threshold or that saturates the detector. The digital gain was thus left at the maximum gain level (255 arbitrary units (a.u.)) for all channels during particle measurements discussed here.

To explore the influence of amplification voltage on particle detectability, 0.53 µm purple PSLs were chosen to arbitrarily represent the lower limit of detectable fluorescence intensity. Using larger (0.96 µm) particles comprised of the same purple fluorophore, Könemann et al. (2018) showed that the particles were only narrowly detectable above the fluorescence threshold in each of the three channels of a WIBS-4A (same unit as used in Savage et al., 2017) and so the smaller, 0.53 µm PSLs were chosen here as a first proxy for the most weakly fluorescing particles we would expect to detect. To improve the signal to noise ratio (SNR) for the lower fluorescence detection limit, the PMT amplification voltage was varied in seven steps between 500 and 1000 V (corresponding to a gain from 10^3 to 10^6, specification sheet TPMO1060E02, Hamamatsu, June 2016) for purple PSLs and background signals (Fig. 5a). Whereas PSL spectra at a PMT amplification of 500 V were indistinguishable from the background signal (+ 1σ SD), spectra show a discernable peak at λ_{mean} = 415.6 nm above 600 V. Subsequently, the SIBS was operated with a PMT amplification voltage of 610 V corresponding to the lowest SNR threshold accepted (Fig. 5a, b). The detection of small biological particles was tested by measuring the emission spectrum of *S. cerevisiae* as an example of a PBAP (see also Pöhlker et al., 2012). On average, the size of intact *S. cerevisiae* particles range between ~2 – 10 µm (e.g., Pelling et al., 2004; Shaw et al., 1997). To test the ability of the SIBS to detect low intensity emissions, we separately analyzed *S. cerevisiae* particles between 0.5 and 1 µm, which most likely includes cell fragments caused by the aerosolization process (Fig. 5c). The tryptophan-like emission, peaking in detection channel 2 (λ_{mean} = 330.6 nm) for λ_{ex} = 285 nm, reveals intensity values below 100 a.u., which are comparable to fluorescence intensity values derived from 0.53 µm purple PSLs (detection channel 5, λ_{mean} = 415.6 nm, Fig. 5d). These two tests for *S. cerevisiae* and 0.53 µm purple PSLs confirmed the instruments ability to detect emission spectra from particles at least as strongly fluorescent as these two test cases, leaving a wide range to detect larger and more intensely fluorescing particles. By using a 3σ SD threshold, the fluorescence peak at λ_{mean} = 415.6 nm of 0.53 µm purple PSLs is still detectable but cannot be distinguished from the background signal at a 6σ SD threshold anymore. Therefore, fluorescence intensity values at the lower detection limit should be treated with care. Corrected spectra of both *S. cerevisiae* and 0.53 µm purple PSLs can be found in the supplement (Fig. S6). By operating the SIBS at relatively low detector amplification, very weak fluorescence, especially from small particles (< 1 µm) might not exceed the detection threshold dur-

ing field applications and would be missed. Further investigation will be necessary to choose amplification voltages appropriate for individual applications where smaller or otherwise weakly fluorescent particles might be particularly important. For all subsequent measurements discussed here, a PMT amplification voltage of 610 V was used.

Saturation only occurred for 15 and 20 µm non-fluorescent PS-DVB particles. As highlighted in Figure S7, the polystyrene/detergent signal (Könemann et al., 2018) at λ_{ex} = 285 nm for 10 µm PS-DVB particles can be spectrally resolved (Fig. S7b), whereas the spectrum for 15 µm PS-DVB particles (Fig. S7e) is altered due to single particles (~10 % out of 400 particles) saturating the detector (at 62383 a.u.). By comparing the defined lower detection end (Fig. 5) to the upper end (Fig. S7), a quantitative difference of approximately three orders of magnitude can be estimated, indicating a wide detectable range at the chosen amplification voltage setting.

4.3 Wavelength-dependent spectral correction of detector

The 16 cathodes of the PMT should be considered as independent detectors with wavelength dependent, individual responsivity and amplification characteristics. In combination with physical properties of technical components (e.g., excitation sources, optical filters, gratings), an instrumental-specific spectral bias might result in incorrect or misleading spectral patterns if not corrected (e.g., DeRose, 2007; DeRose et al., 2007; Holbrook et al., 2006). To compensate for such potential instrumental biases, we used a spectral correction approach as described in Sect. 2.4. The spectral correction factors are comparable to the theoretical responsivity of the PMT with the highest correction for channels 1-4 (λ_{mean} = 302.2 – 387.3 nm) and 14-16 (λ_{mean} = 666.5 – 721.1 nm) (Fig. 6). Channel 8 (λ_{mean} = 500.0 nm) shows the highest responsivity and channels 6 and 7 (λ_{mean} = 443.8 and 471.9 nm) exhibit a noticeable lower responsivity than their adjacent channels (see also Sect. 4.4.1). The spectral correction shows several peaks (e.g., detector channels 3, 5, and 8) and dips (e.g., detector channels 4, 6, and 7) (Fig. 6), however, this pattern is due to gain variations for different channels and is not noise.

It is important to note that the detector settings and spectral correction uniquely refer to the SIBS unit as it was used for the current study. Due to technical and physical variability as stated above, it is likely that the spectral correction required for other SIBS units would be somewhat different. Furthermore, the wavelength-dependent detector correction may change over time due to material fatigue or contaminations in the optical chamber affecting background signal measurements. Periodic surveillance and adjustments are therefore required, especially after measurements where the instrument was exposed to high particle concentrations or was operated during extreme weather or environmental conditions (e.g., temperature, humidity, vibration). For particle sizing verification, we recommend the use of 0.5, 1, and 3 µm non-fluorescent PSLs. Regarding a fluorescence response check,

we recommend 2 µm green and 2 µm red PSLs for the validation of the spectral responsivity maximum and the upper (near-IR) detection range. To our knowledge, no fluorescent dyed PSLs are available to verify the response within the lower spectral detection range (UV) of the SIBS. However, the polystyrene signal of 3 µm non-fluorescent PSLs (Fig S7g, h, i, see also Könemann et al., 2018) represents a compromise between signal strength at λ_{ex} = 285 nm and aerosolization efficiency (compared to PSLs with larger sizes) for a spectral responsivity validation.

4.4 Fluorescence spectra of standards

4.4.1 PSL standards

The SIBS spectra for the four different PSL standards, covering an emission range from UV to near-IR, generally agree well with the corresponding reference spectra (Fig. 7). Each of the two excitation wavelengths probe separate fluorescent modes, which appear at approximately the same emission wavelength for a given PSL type (e.g., λ_{em} = ~580 nm for red PSLs, Fig. 7j), as discussed by Könemann et al. (2018). Moreover, even the rather weak polystyrene and detergent fluorescence, systematically associated with PSL suspensions (Könemann et al., 2018), is resolved by the SIBS at λ_{ex} = 285 nm and λ_{em} = ~300 nm (Fig. 7b, e, h, k). It is further noteworthy that emission~~s~~ *intensity* at λ_{ex} = 285 nm ~~are~~ *is* generally higher than derived emission~~s~~ *intensity* at λ_{ex} = 370 nm (Fig. 7c, f, i, l), supporting the finding that a particle receives higher irradiance values from xenon 1 than from xenon 2 (see also Sect. 3.3).

As mentioned in Sect. 4.3, detection channels 6 and 7 require relatively large correction factors. For 2.07 µm purple PSLs (Fig. 7b, c), the SIBS spectra closely match the references spectra after correction. For the 2.1 µm blue PSLs (Fig. 7e, f), however, the corrected spectrum matches the reference spectrum well, except at detection channel 6 (λ_{mean} = 443.8 nm), where the SIBS spectrum is lower than the reference spectrum by approximately 50%. This effect was also observed for 1 µm blue PSLs (Thermo Fisher, B0100), doped with the same fluorophore (data not *shown* ~~*included in this study*~~). The reason for this *discrepancy* ~~*malfunction*~~ is unknown. Nevertheless, because this effect only occurs noticeably for highly fluorescent blue PSLs and NAD (see also Sect. 4.4.2), one explanation could be that the instrument-dependent dynode cascade (the electronic amplification stages) for this particular detection channel is suppressed, resulting in a lower amplification efficiency. In this case, relatively low signals could be amplified correctly, whereas medium or high intensity emission could only be amplified up to a certain level. The amplification threshold for detection channel 6 is, however, unknown and needs further verification.

4.4.2 Biofluorophore standards

Figure 8 and 9 highlight fluorescence spectra of different biofluorophores measured by the SIBS, which correspond to related reference spectra (compare also Pöhlker et al., 2012), showing that amino acids (fluorescence emission only at λ_{ex} = 285 nm), co-enzymes and flavin compounds (fluorescence emission at λ_{ex} = 285 and 370 nm), and chlorophylls (fluorescence emission only at λ_{ex} = 370 nm) can be spectrally distinguished.

The uncorrected spectrum of tryptophan (Fig. S9) highlights the necessity of a spectral correction to compensate for the low detector responsivity within the UV and near-IR bins. If the fluorescence signal of tryptophan remains uncorrected, the spectra is shifted slightly to longer wavelengths (red-shifted) due to the low responsivity of channel 2 in comparison to channel 3, resulting in misleading spectral information. For NAD (Fig. 8h, i), fluorescence intensity values of channel 6 are lowered due the suppressed amplification efficiency in this particular channel as described for blue PSLs (Sect. 4.4.1).

All biofluorophores (except chlorophyll types) were aerosolized as dry powders (see Sect. 2.2) to avoid fluorescence solvatochromism effects~~, means solvent-dependent spectral shifts relative to the dry fluorophore state, which serves as a reference case here~~ (e.g., Johnson et al., 1985). Solvatochromism of fluorophores in aqueous solution – the only atmospherically relevant case – typically shifts fluorescence emissions to longer wavelengths due to the stabilized excited state caused by polar solvents (Lakowicz, 2004). This spectral red-shift can be seen in Figure S10, where the peak maximum for NAD shows a difference of ~15 nm between a dry and water-solv*ateded* state, whereas riboflavin reveals an even higher shift of ~37 nm. Here, solvatochromism serves as an example for fluorescence spectra that vary substantially as a function of the fluorophore's microenvironments (e.g., solvent polarity, pH, temperature).

Each of the three types of chlorophyll exhibit the weakest emission of all biofluorophores measured within this study, however the SIBS was able to detect the fluorescence signal at λ_{ex} = 370 nm for all three (Fig. 9). The spectral difference between chlorophyll *a* and *b* is only minor at λ_{ex} = 370 nm ($\Delta\lambda$ = 8.3 nm) for which the spectral resolution of the SIBS is not capable of distinguishing between ~~*both*~~ types (Fig. 9a, b, c, d and Fig. S11) (e.g., French et al., 1956; Welschmeyer, 1994). Nevertheless, the SIBS shows the ability to distinguish between chlorophyll *a* and *b*, and bacteriochlorophyll due to the red-shift in the bacteriochlorophyll spectrum ($\Delta\lambda$ = 28.5 nm at λ_{ex} = 370 nm, between chlorophyll *a* and bacteriochlorophyll). This may provide a further discrimination level regarding algae, plant residues, and cyanobacteria. Bacteriochlorophyll also shows a second and even stronger emission peak at λ_{ex} = 370 nm (λ_{em} = ~800 nm) that could help further distinguish it from chlorophyll *a* and *b*, but the SIBS spectrometer cannot currently detect this far into the IR (e.g., Rijgersberg et al., 1980; Van Grondelle et al., 1983).

Overall, fluorescence emissions recorded by the SIBS are in good agreement with measured reference spectra. However, care must be taken as to the interpretation of fluorescence emissions covering broad spectral ranges, which span regimes with large differences between individual correction factors (e.g., channel 15 (λ_{mean} = 693.9 nm, Fig.7l) and channel 2 (λ_{mean} = 330.6 nm, Fig.8k). For the SIBS, namely the first two UV detection channels and the last two near-IR channels *should* ~~*have to*~~ be treated with care. Further investigation~~*s*~~ ~~*are*~~ is required for a careful assessment of how the spectral correction can be applied properly *with respect to* ~~*onto*~~ fluorescent and non-fluorescent atmospheric particles.

4.5 Particle asymmetry measurements

The AF of spherical particles such as PSLs (Fig. 10a, b) and ultrapure water droplets is approximately 10 (Table 3), which is slightly higher than reported values for spherical particles by, e.g., Savage et al. (2017) (AF= ~5) or Toprak & Schnaiter (2013) (AF= ~8) using a WIBS. It is noteworthy that the AF of water droplets increases slightly with increasing droplet size and, therefore, contributes to the mean value (Fig. S13). This effect is most likely based on a decreasing surface tension with increasing droplet size for which the droplet morphology is changed to a more oval shape within the sample flow. A similar effect regarding a potential droplet deformation using an Airborne Particle Classifier (APC) was observed by Kaye et al., (1991). Even if the morphology of ammonium sulfate (crystalline, Fig. 10d) and Fe_3O_4 (irregular clusters, Fig. 10f) is diverse, the~~*ir*~~ difference~~*s*~~ in AF~~*s*~~ is only minor (~13 and 14, Table 3), indicating that most naturally occurring aerosols (e.g., sea salt, soot, various bacterial and fungal clusters) will occur in a AF regime between ~10 and 20. Only rod-shaped carbon nanotubes (110-170 nm diameter, 5-9 µm length) show increased AFs with a mean value at ~22 (Table 3) at which also, e.g., bacteria would occur (Fig. 10h). No particles observed exhibited average AF values >25, as would have been expected for, e.g., carbon nanotubes. Because the range of AF values for homogenous particles is relatively broad and the differences between morphologically diverse particle types is only minor (Table 3), the question can be raised to what extent particles could be distinguished based on the AF under ambient conditions. *Similar broad AF ranges were found in Healy et al., (2012), measuring sodium chloride, chalk, and several pollen and fungal spores types.* As also discussed by Savage et al. (2017), the AF values reported by SIBS and WIBS units should be treated with extreme care.

The validation of asymmetry measurements is challenging due to unavoidable particle and aerosolization effects (e.g., particle agglomeration and spatial orientation within the sample flow) and the lack of standardized procedures for AF calibrations. Measurements performed in this study do, therefore, only serve as a rough AF assignment. Moreover, even if both the SIBS and WIBS use the same technical components for defining AFs, a direct intercomparison cannot be applied due to technical

variability (e.g., PMT related signal-to-noise ratio or the alignment of optical components). Additionally, it is currently unknown in how far the 785 nm diode laser of the SIBS affect asymmetry measurements compared to the WIBS using a 635 nm diode laser.

4.6 Initial ambient measurements

Several weeks of initial ambient SIBS measurements ~~have been~~ *were* conducted on the roof of the Max Planck Institute for Chemistry in Mainz, Germany. At *a nearby building site* ~~the same location~~, Huffman et al. (2010) conducted one of the first ambient UV-APS studies *in the year 2006*. Moreover, Toprak & Schnaiter (2013) conducted a WIBS-4A study at a comparable site in central Germany *from 2010 to 2011*. The aim of this brief ambient section is to validate that the SIBS-derived key aerosol and fluorescence data are *reasonable and relatively* consistent with the aforementioned studies. We found a good agreement between the coarse mode (≥1 µm) number concentrations ($N_{T,c}$) of the SIBS ($N_{T,c}$ ranging from *0.25* ~~0.19~~ to *1.59* ~~1.24~~ cm^{-3}, with a mean of *0.76* ~~0.59~~ cm^{-3})~~,~~ *and previous data from* the UV-APS (mean $N_{T,c}$: 1.05 cm^{-3} (Huffman et al., 2010))~~,~~ and the WIBS-4A (mean $N_{T,c}$: 0.58 cm^{-3} (Toprak and Schnaiter, 2013)) (Fig.11a). Furthermore, good agreement was found between coarse mode fluorescent number concentrations ($N_{F,c}$) of the SIBS ~~with a 3σ SD threshold~~ (mean $N_{F,c\ (3\sigma)}$: *0.025* ~~0.019~~ cm^{-3}), the UV-APS (mean $N_{F,c}$: 0.027 cm^{-3} (Huffman et al., 2010)), and the WIBS-4A ~~with a 3σ SD threshold~~ (mean $N_{F,c\ (3\sigma)}$: 0.031 cm^{-3} (Toprak and Schnaiter, 2013)) (Fig.11a). Similarly, the fraction of fluorescent particles in the coarse mode ($N_{F,c}/N_{T,c}$) compares well between SIBS ~~with a 3σ SD threshold~~ (mean $N_{F,c(3\sigma)}/N_{T,c}$: 4.2 %), the UV-APS (mean $N_{F,c}/N_{T,c}$: 3.9 % (Huffman et al., 2010)), and the WIBS-4A ~~with a 3σ SD threshold~~ (mean $N_{F,c(3\sigma)}/N_{T,c}$: 7.3 % (Toprak and Schnaiter, 2013)) (Fig.11b). Expectedly, a 1σ SD threshold gives much higher *SIBS* fluorescent fractions of 39.2 %, whereas a 6σ SD threshold corresponds with much lower fluorescent fractions of 1% (Fig.11b). Note that no prefect match between our results and the studies by Huffman et al. (2010), and Toprak & Schnaiter (2013) can be expected, since the measurements took place with different sampling setups and during different seasons. Furthermore, the spectrally resolved SIBS data makes the definition of fluorescent fraction more complex than for UV-APS and WIBS data (see Sect. 2.6). However, the overall good agreement confirms that the SIBS produces reasonable results in an ambient setting. Further, the single particle fluorescence spectra are reasonable with respect to typical biofluorophore emissions (Pöhlker et al., 2012). Exemplary spectra (λ_{ex} = 285 and 370 nm) of ambient single particles can be found in the supplement (Fig.S14). An in-depth analysis of extended SIBS ambient datasets is subject of ongoing work.

5. Summary and conclusions

Real-time analysis of atmospheric bioaerosols using commercial LIF instruments has largely been restricted to data recorded in only 1-3 spectrally integrated emission channels, limiting the interpretation of fluorescence information. Instruments that can record resolved fluorescence spectra over a broad range of emission wavelengths may thus be required to further improve the applicability of LIF instrumentation to ambient PBAP detection. Introduced here is the SIBS *(DMT, Longmont, CO, USA),* ~~as a new aerosol fluorescence detector,~~ which *is an instrument that* provides resolved fluorescence spectra (λ_{mean} = 302 – 721 nm) from each of two excitation wavelengths (λ_{ex} = 285 and 370 nm) for single particles. The current study introduces the SIBS by presenting and experimentally validating its key functionalities. This work critically assesses the strengths and limitations of the SIBS with respect to the growing interest in real-time bioaerosol quantification and classification. It should be noted that the study is an independent evaluation that was not conducted*,* ~~or~~ endorsed*, or co-authored* by the manufacturer *or representatives*. Overall, this work confirms a precise particle sizing between 300 nm and 20 µm and the particle discrimination ability based on spectrally resolved fluorescence information of several standard compounds.

The SIBS was operated at a low PMT detector amplification setting (610 V) to retain capacity to detect large or brightly fluorescent particles. It was confirmed, however, that even weak fluorescence signals from 0.53 µm purple PSLs and from small *S. cerevisiae* fragments (0.5 - 1 µm) can be clearly distinguished from the background signal. Saturation events were only observed for the polystyrene/detergent signal from relatively large 15 and 20 µm PS-DVB particles. Nevertheless, the fluorescence intensity detection threshold is highly instrument-dependent due to the complex interaction of single technical components across individual instruments. For example, xenon 1 exhibited ~154 % higher irradiance than xenon 2 (both new lamps) due to differences in the properties of xenon emission and the optical filters used. For used xenon lamps (> 4000 hours of use), an even higher difference of ~220 % was observed. Thus, a defined fluorescence detection threshold will most likely change over time due to, e.g., material fatigue. *Additionally, alternating irradiance properties might significantly contribute to observed differences in performance of similar instrument types (e.g., Hernandez et al., 2016), expressly underlining the need for a fluorescence calibrant applicable across LIF-instruments (e.g., Robinson et al., 2017). Nevertheless, to the best of our knowledge, there is currently no standard reference available that fulfills the requirements to serve as a calibrant for multi-channel, multi-excitation LIF-instruments.* ~~These o~~*O*bservations *in this study* are valid not only for the SIBS, but also for the WIBS-4A and WIBS-NEO and lead to important implications for interpretation of particle data. In particular, a particle that exhibits measurable fluorescence in WIBS channel FL1, but only weak fluorescence in channel FL3 could be assigned as an “A-type” particle

in one instrument but an "AC-type" particle in an instrument with slightly stronger xenon 2 irradiance. These differences in classification can be extremely important to interpretation of ambient data (e.g., Perring et al., 2015; Savage et al., 2017).

The PMT used in the SIBS shows a wavelength-dependent sensitivity distribution along all 16 detection channels. To compensate for this characteristic and to be able to use the broadest possible fluorescence emission range, the measured emission spectra were corrected with respect to reference spectra acquired from deuterium and halogen lamps. A spectral correction over a broad emission range also introduces drawbacks, however, that LIF-instrument users should keep in mind while interpreting derived fluorescence information. In particular, the first two (UV) and the last two (near-IR) detection channels should be treated with care, because they ~~*have*~~ *require* larger correction factors compared to adjacent channels. Ultimately, the correction factor and amplification voltages applied to the detector will be experiment-specific and will need to be investigated with respect to individual experimental aims. *To this extent, possible differences between instruments and important calibrations complicate the concept of the instrument being commercially available. Individual users may desire to be able to purchase the SIBS as a "plug-and-play" detector, but using without a critical understanding of these complexities would not be appropriate at this time and could lead to inadvertent misinterpretation of the data.*

Fluorescence spectra of fluorescent PSLs, amino acids, co-enzymes, and flavins measured by the SIBS agree well with corresponding spectra recorded with an offline reference spectrometer. Thus, the SIBS was shown to be capable of clearly distinguishing between different particle types based on resolved fluorescence information. Furthermore, the extended fluorescence emission range (λ_{em} = > 700 nm) enables the SIBS also to distinguish chlorophyll *a* and *b* from bacteriochlorophyll, potentially opening new possibilities for the detection of, e.g., algae, plant residues, and cyanobacteria.

Particle asymmetry measurements revealed that spherical PSLs have an AF of 9.9 (± 3.6), whereas other materials (ammonium sulfate, Fe_3O_4, and carbon nanotubes) show AF values of 13.1 (± 8.1), 14.4 (± 7.4), and 21.6 (± 12.7), respectively. Because differences of measured AF value between morphologically diverse particle types are small and within the ranges of uncertainty for the measurement of a given set of particles, it is questionable how well particles can be distinguished based on the AF *as presently measured* by the quadrant PMT ~~*as presently measured*~~. Users of SIBS and WIBS instruments should apply extreme care if using AF data. It is also likely that different instrument units may have very different AF responses with respect to this measurement. At a minimum, each individual unit needs to be rigorously calibrated to known particle types to determine if AF values are sufficiently different (e.g., separated by several standard deviations) to justify scientific conclusions based on the metric.

Exemplary ambient data, measured between the 12th and 18th of April 2018 on the roof of the Max Planck Institute for Chemistry in Mainz (Germany), are consistent with LIF measurement data using a UV-APS (Huffman et al., 2010) and a WIBS-4A (Toprak and Schnaiter, 2013). Total coarse particle number concentrations revealed a mean value of *0.76* ~~0.59~~ cm^{-3} (1.05 cm^{-3} (Huffman et al., 2010); 0.58 cm^{-3} (Toprak and Schnaiter, 2013)) of which ~4.2% are considered to be fluorescent using a 3σ SD threshold (3.9% (Huffman et al., 2010); 7.3% (Toprak and Schnaiter, 2013)), including only particles that show fluorescence emission in~~,~~ at least~~,~~ two adjoining detection channels. Using a 1σ and 6σ SD threshold results in fluorescent fractions of 39.2% and 1% respectively. However, the applicability of different threshold strategies for the SIBS is currently under investigation and needs further verifications.

The results suggest that the SIBS has the ability to increase the selectivity of detection of fluorescent biological and non-biological particles by use of two excitation wavelengths and 16-channel resolved fluorescence information in combination with a broad detectable emission range. The applicability of described methods onto ambient datasets is currently under investigation. Data shown here and the detailed insights of technical components used in the SIBS will be broadly beneficial for users of LIF instruments providing resolved fluorescence information, but also for users of various generations of WIBS and other LIF instruments widely applied within the bioaerosol community.

6. Data availability

The data of the key results presented here can be provided upon request. For specific data requests, please refer to the corresponding authors.

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Appendix A: List of acronyms and symbols.

Acronym/Symbol	Description
AF	Asymmetry factor
APC	Airborne Particle Classifier
CCD	Charge-coupled device
DMT	Droplet Measurement Technologies
EEM	Excitation-emission matrix
EM	Emission
EX	Excitation
IR	Infrared
LIF	Light-induced fluorescence
N	Particle number concentration (cm^{-3})
$N_{T,c}$	*N* of total coarse particles (1-20 µm)
$N_{F,c(n\sigma)}$	*N* of fluorescent coarse particles (1-20 µm) at 1, 3, or 6σ
NAD	Nicotinamide adenine dinucleotide
NAD(P)H	Nicotinamide adenine dinucleotide and nicotinamide adenine di-nucleotide phosphate
NIST	National institute of standards and technology
PBAP	Primary biological aerosol particles
PMT	Photomultiplier tube
PAH	Polycyclic aromatic hydrocarbons
PSL	Polystyrene latex sphere
PS-DVB	Polystyrene-divinylbenzene
SD	Standard deviation
SIBS	Spectral intensity bioaerosol sensor
SNR	Signal to noise ratio
TSP	Total suspended particles
UV	Ultraviolet
UV-APS	Ultraviolet aerodynamic particle sizer
Vis	Visible light
WIBS	Wideband integrated bioaerosol sensor

Table 1. Lower, mean, and upper wavelength at each PMT detection channel. Nominal data according to manufacturer Hamamatsu.

Channel	λ_{lower} (nm)	λ_{mean} (nm)	λ_{upper} (nm)
1	298.2	302.2	316.2
2	316.6	330.6	344.6
3	345.0	359.0	362.5
4	377.5	387.3	401.3
5	401.5	415.6	429.7
6	429.8	443.8	457.8
7	457.9	471.9	485.9
8	486.0	500.0	514.0
9	514.0	528.0	542.0
10	541.9	555.9	569.9
11	569.7	583.7	597.7
12	597.4	611.4	625.4
13	625.0	639.0	653.0
14	652.8	666.5	680.2
15	679.9	693.9	707.9
16	707.1	721.1	735.1

1147 **Table 2.** Parameters and technical components of the SIBS in comparison to the WIBS-NEO and
1148 WIBS-4A. Data are taken from manufacturer information.

	SIBS	**WIBS-NEO**	**WIBS-4A**
First production	*2015*	*2016*	*2009*
Measured parameters	Particle size Asymmetry Factor Fluorescence spectra	Particle size Asymmetry Factor Integrated fluorescence in 3 channels	Particle size Asymmetry Factor Integrated fluorescence in 3 channels
Particle size range	~0.3 – 100 µm	~~0.3~~ *0.5* – ~~100~~ *30* µm	~0.5 – 20 µm
Maximum concentration	~2 x 10^4 particles/L	~2 x 10^4 particles/L	~2 x 10^4 particles/L
Fluorescence excitation	λ_{ex}= 285 and λ_{ex}= 370 nm	λ_{ex}= 280 and λ_{ex}= 370 nm	λ_{ex}= 280 and λ_{ex}= 370 nm
Fluorescence emission	λ_{mean}= 302 – 721 nm (16-channel PMT)	λ_{em}= 310-400 nm and λ_{em}= 420-650 nm	λ_{em}= 310-400 nm and λ_{em}= 420-650 nm
Flow rate	Sample flow:~0.3 l/min Sheath flow:~2.2 l/min (re-circulating)	Sample flow:~0.3 l/min Sheath flow:~2.2 l/min (re-circulating)	Sample flow:~0.3 l/min Sheath flow:~2.2 l/min (re-circulating)
Laser	785 nm diode laser, 55 mW	635 nm diode laser, 15 mW	635 nm diode laser, 12 mW
Pump	Diaphragm pump	Diaphragm pump	Diaphragm pump
Power requirements	200 W, 90 - 230 VAC	150 W, 90 - 230 VAC	150 W, 90 - 230 VAC
Weight (kg)	20.1	12.5	13.6
Dimension W x L x H (cm)	42.5 x 61.5 x 23.5	45.1 x 36.2 x 24.1	30.4 x 38.2 x 17.1

Table 3. Asymmetry factor (AF) values for reference particles. Values are based on the mean of a Gaussian fit applied onto each particle histogram (see also Fig. 10), including 1σ SD.

	AF
2 µm non-fluorescent PSLs	9.9 ± 3.6
Ultrapure water	11.9 ± 2.9
Ammonium sulfate	13.1 ± 8.1
Fe_3O_4	14.4 ± 7.4
Carbon nanotubes	21.6 ± 12.7

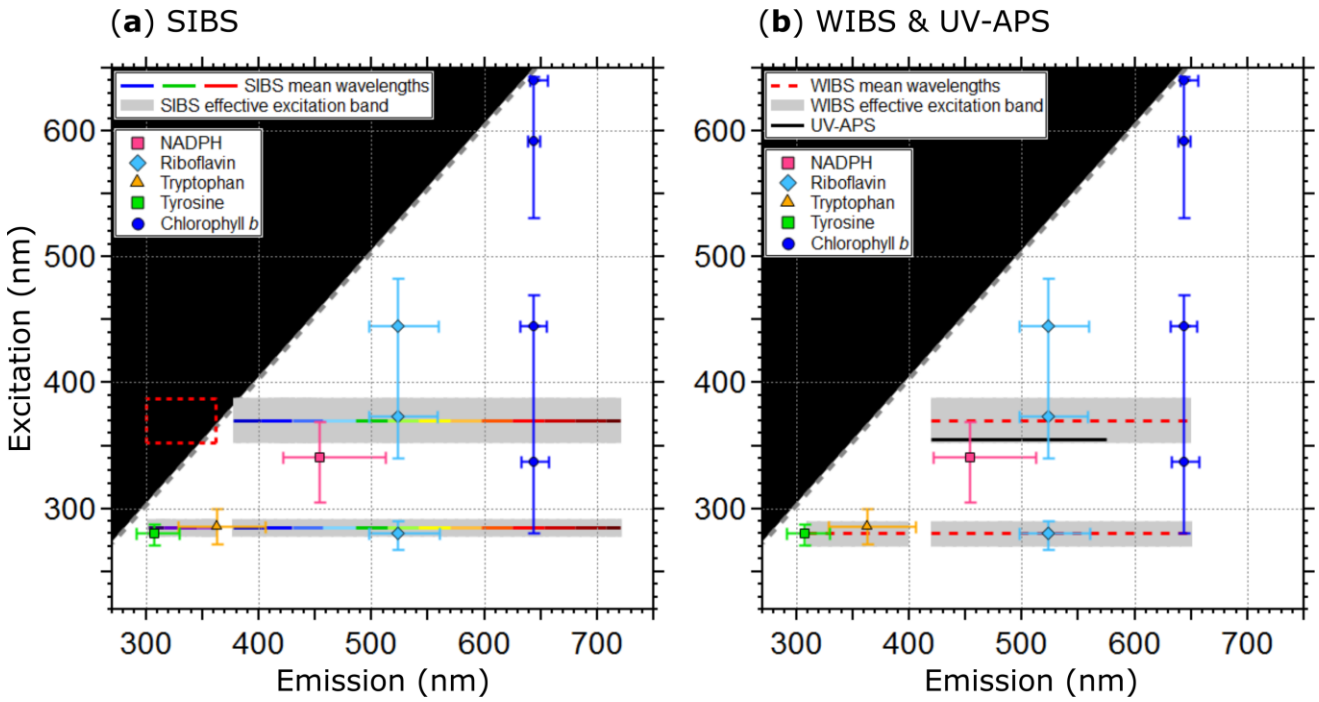

Figure 1. Optical design and overview of excitation and emission specifications of the LIF instruments UV-APS, WIBS, and the SIBS with spectral locations of the autofluorescence modes of the biofluorophores tyrosine, tryptophan, NAD(P)H, riboflavin, and chlorophyll *b* (as examples). Here the term WIBS includes the WIBS-4A and WIBS-NEO, because both instruments use the same optical components. Spectral properties of the emission bands of LIF instruments are illustrated as horizontal lines. The color-coded bars in (**a**) illustrate the spectrally resolved fluorescence detection of the two excitation wavelengths (λ_{ex} = 285 and 370 nm) by the SIBS. The "blind spot" (white notch) at λ_{ex} = 285 nm between λ_{em} = 362 - 377 nm (**a**) originates from a notch optical filter, used to block incident light from the excitation sources. Grey dashed lines show the 1st order elastic scattering. At λ_{ex} = 370 nm, the detection range of the SIBS includes the spectral range where $\lambda_{em} < \lambda_{ex}$, for which fluorescence is not defined and so data within the red dashed rectangle is omitted (**a**). Grey bars indicate the effective excitation bands of optical filters used for the WIBS and SIBS (see also Sect. 3.3 and Fig. 3). The effective excitation bands in the WIBS and SIBS occur in a spectral range spanning several nanometers (up to 36 nm), in contrast to the UV-APS (black line, **b**), which uses a laser source with a defined excitation (Figure adapted from Pöhlker et al., 2012).

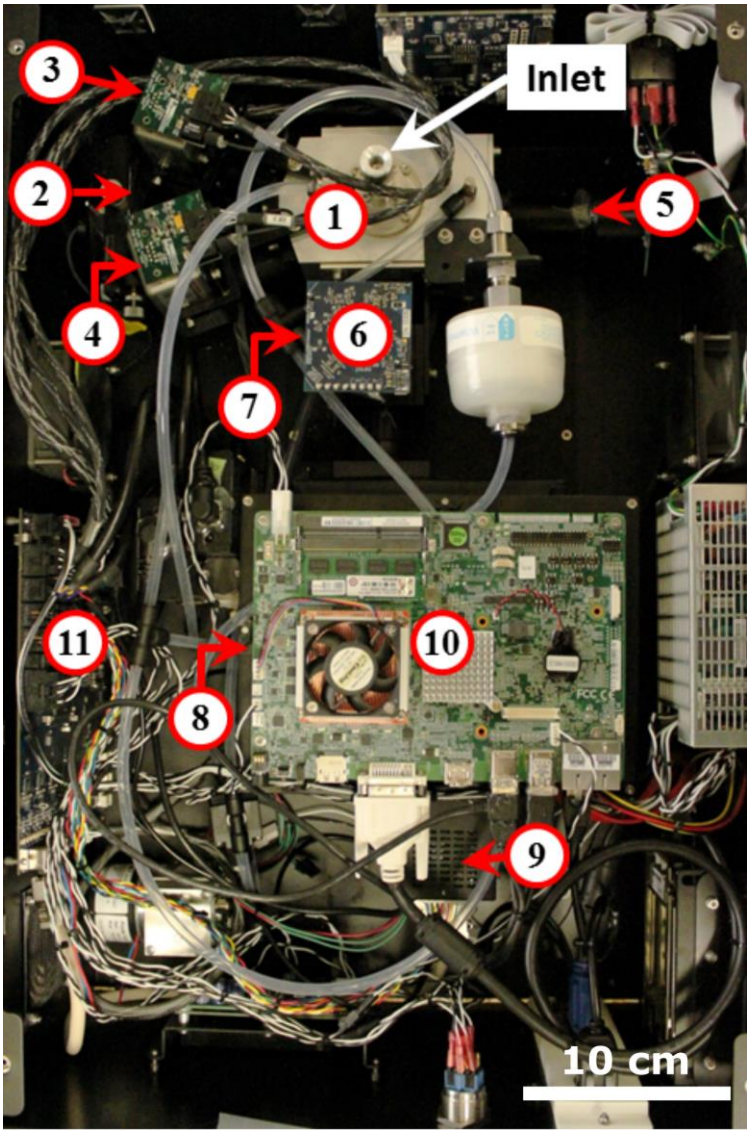

Figure 2. Technical components within SIBS body. (1) Optical cavity. (2) Continuous wave diode laser used for particle detection and sizing. (3) and (4) Xenon light sources. (5) Quadrant PMT used for the determination of particle asymmetry. (6) PMT used for particle detection and sizing. (7) Dichroic beamsplitter separates side-scattered light (particle sizing) and fluorescence emission (not visible; below component (6)). (8) Grating polychromator (below component (10)). (9) 16-channel PMT used for detection of fluorescence. (10) Embedded computer unit. (11) Control-board.

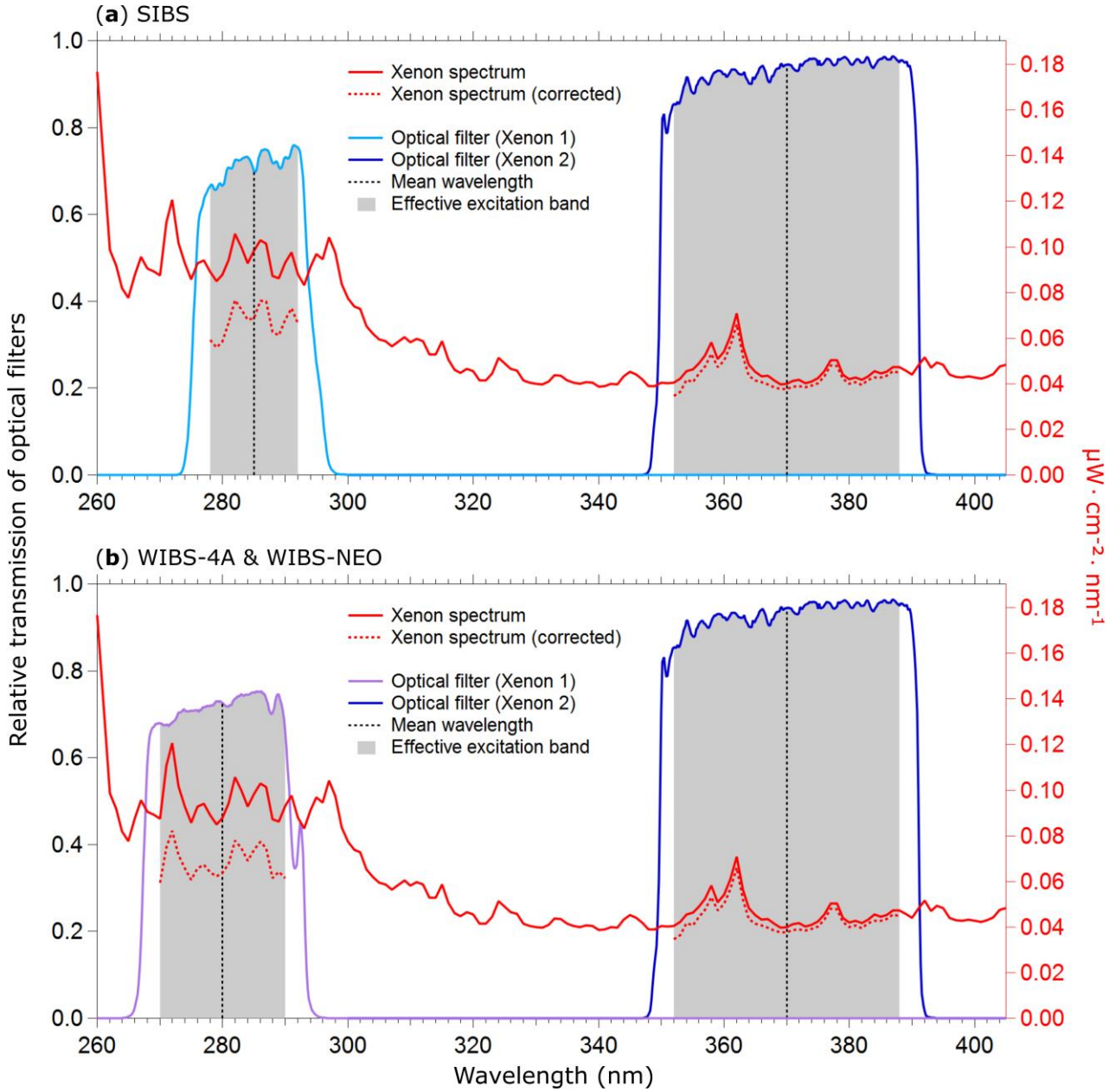

Figure 3. Irradiance from xenon flash lamps based on specifications of lamps and optical filters. Purple and blue lines show optical transmission of filters (left axes) applied to select excitation wavelength. Gray bands indicate where filter transmit light relative from the mean wavelength. Red lines show theoretical irradiance values of the xenon flash lamp (right axes): solid line (raw output), dashed line (relative output after filtering). Relative output shown as raw output multiplied by effective excitation band of the bandpass filters used in the: (**a**) SIBS ($\Delta\lambda_{ex\ (Xenon1)}$ = ~14 nm; $\Delta\lambda_{ex\ (Xenon2)}$ = ~36 nm), and (**b**) WIBS-4A and WIBS-NEO ($\Delta\lambda_{ex\ (Xenon1)}$ = ~20 nm; $\Delta\lambda_{ex\ (Xenon2)}$ = ~36 nm). Xenon lamp operating conditions: 600 V main voltage, 0.22 µF main capacitance, 126 Hz repetition rate, 500 mm distance. (Data courtesy: Xenon flash lamps / Hamamatsu; Single-band bandpass filters / Semrock).

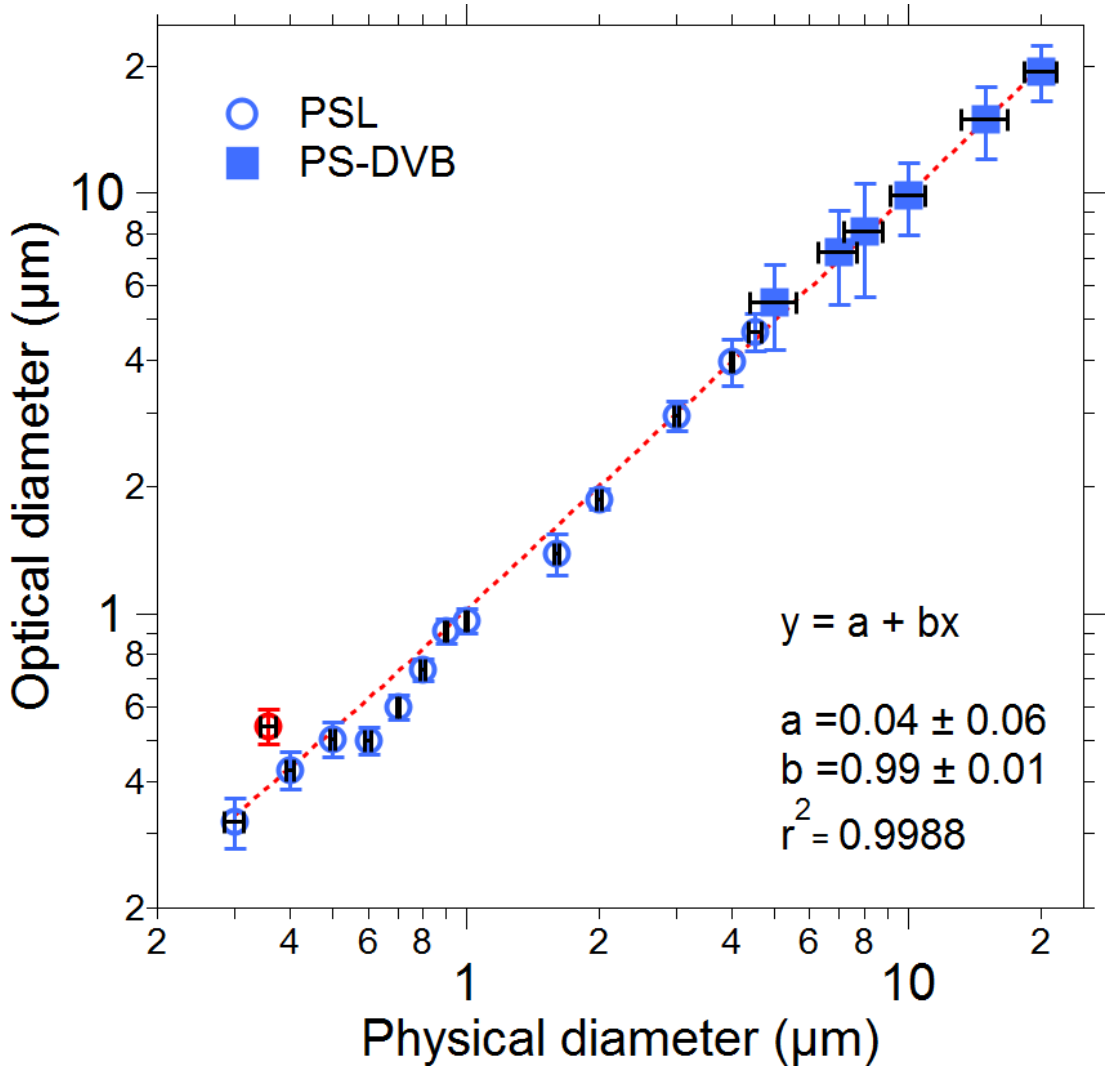

Figure 4. Size calibration of SIBS. Black horizontal bars indicate 1σ SD as stated by each manufacturer (Table S1). Optical diameter values and related 1σ SD are based on a Gaussian fit, which was used to average size distributions of several thousand homogeneous particles for each measurement. The linear fit (red dashed line) excludes the 0.356 µm PSL sample (red marker), an outlier potentially caused by a poor quality PSL batch. Only non-fluorescent particle standards were used for determining the sizing accuracy.

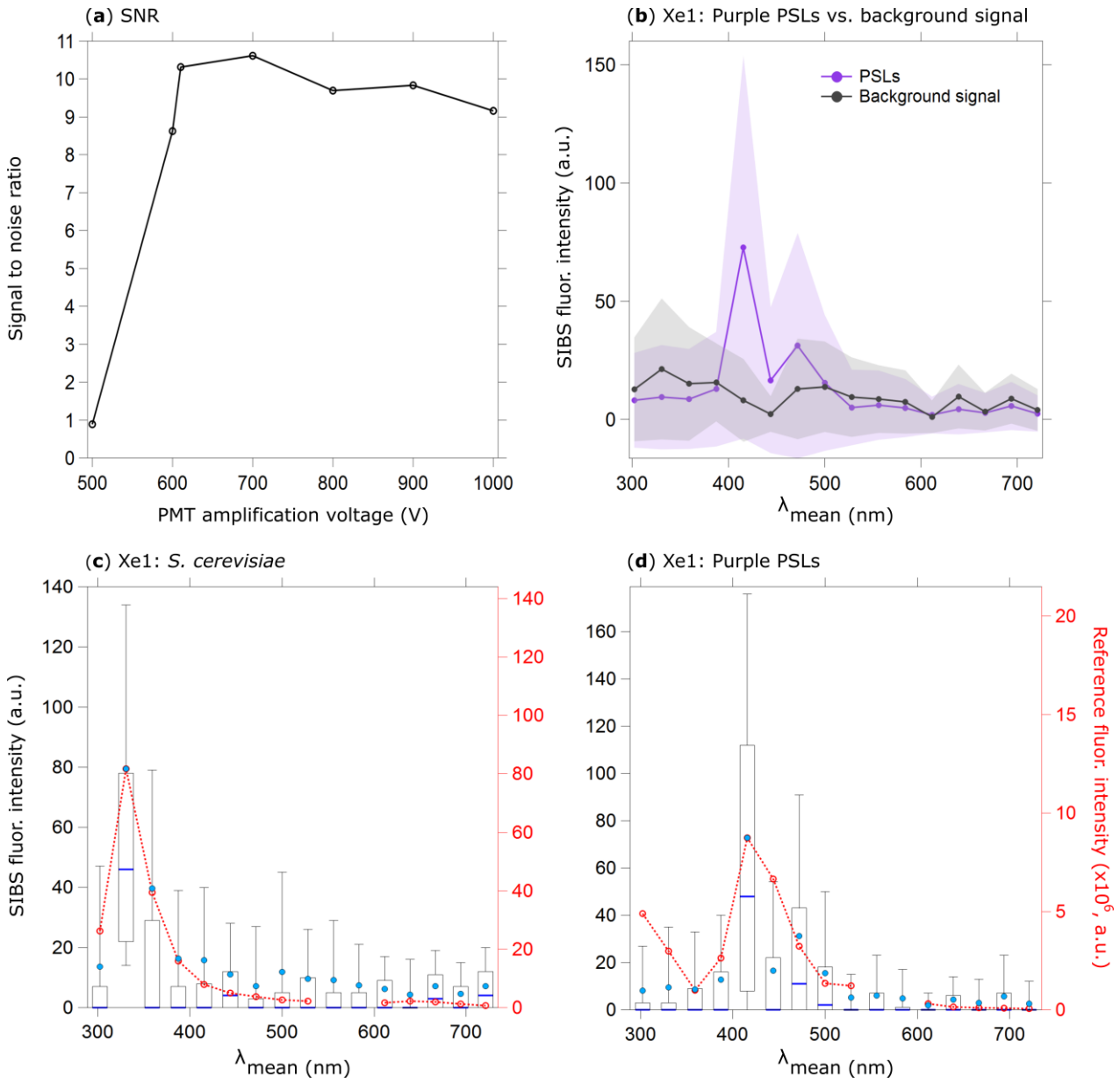

Figure 5. SIBS signal to noise ratio (SNR) in (**a**): emission of 0.53 µm purple PSLs (5260 particles, background signal + 1σ SD subtraction) divided by background signal at different PMT amplification voltages (both at Xe1, channel 5, averaged, and uncorrected). Background signal measured over 5 min. In (**b**), fluorescence emission in contrast to background signal at a PMT amplification voltage of 610 V are shown (same parameters as in (**a**)). Shaded area: 1σ SD. Fluorescence intensity values are shown in arbitrary units. Fluorescence emission spectra of (**c**) *S. cerevisiae* (yeast; 2048 particles, 0.5 – 1 µm) and (**d**) PSLs (as in (**b**)). Red dashed lines and markers (right axes) show averaged and re-binned reference spectra. Box and whisker plots (left axes) show SIBS spectra: median (blue line), mean (circle), boxes 75 and 25 percentile, whiskers 90 and 10 percentile. Data coinciding with 1st or 2nd order elastic scattering were removed from reference spectra.

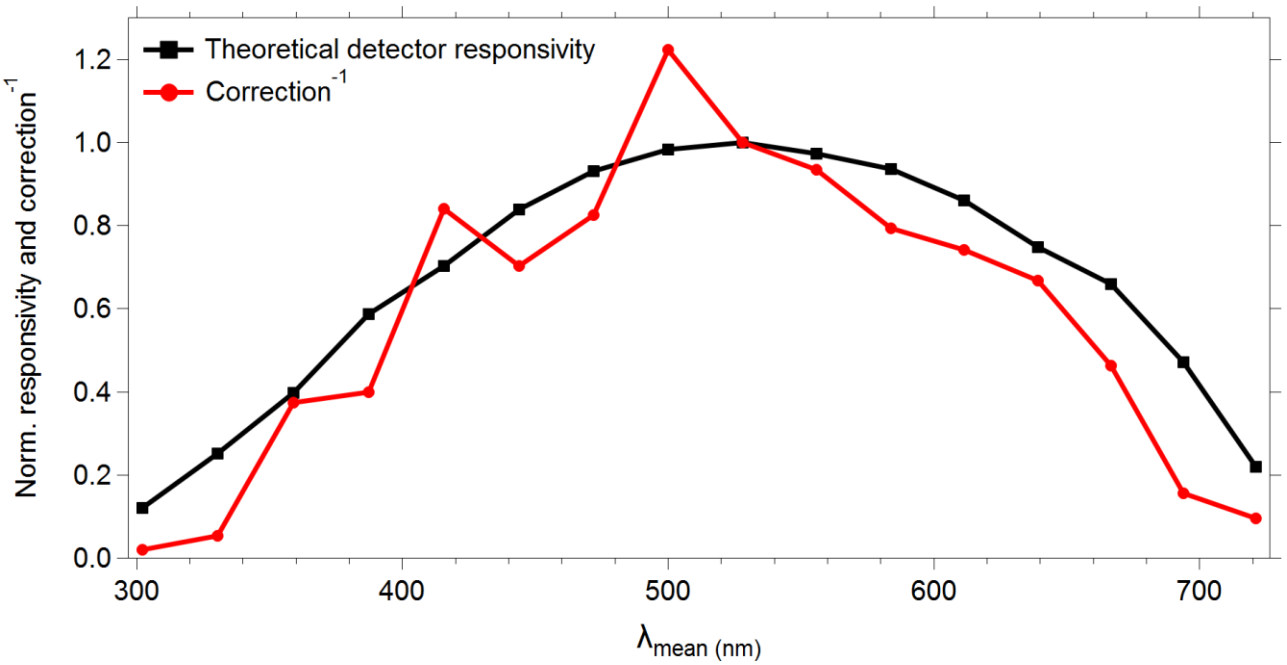

Figure 6. Normalized theoretical detector responsivity and spectral correction. Theoretical detector responsivity derived from measured cathode radiant sensitivity multiplied by the diffraction efficiency (as shown in Figure S8). Note that red line shows inverse of spectral correction to match detector response.

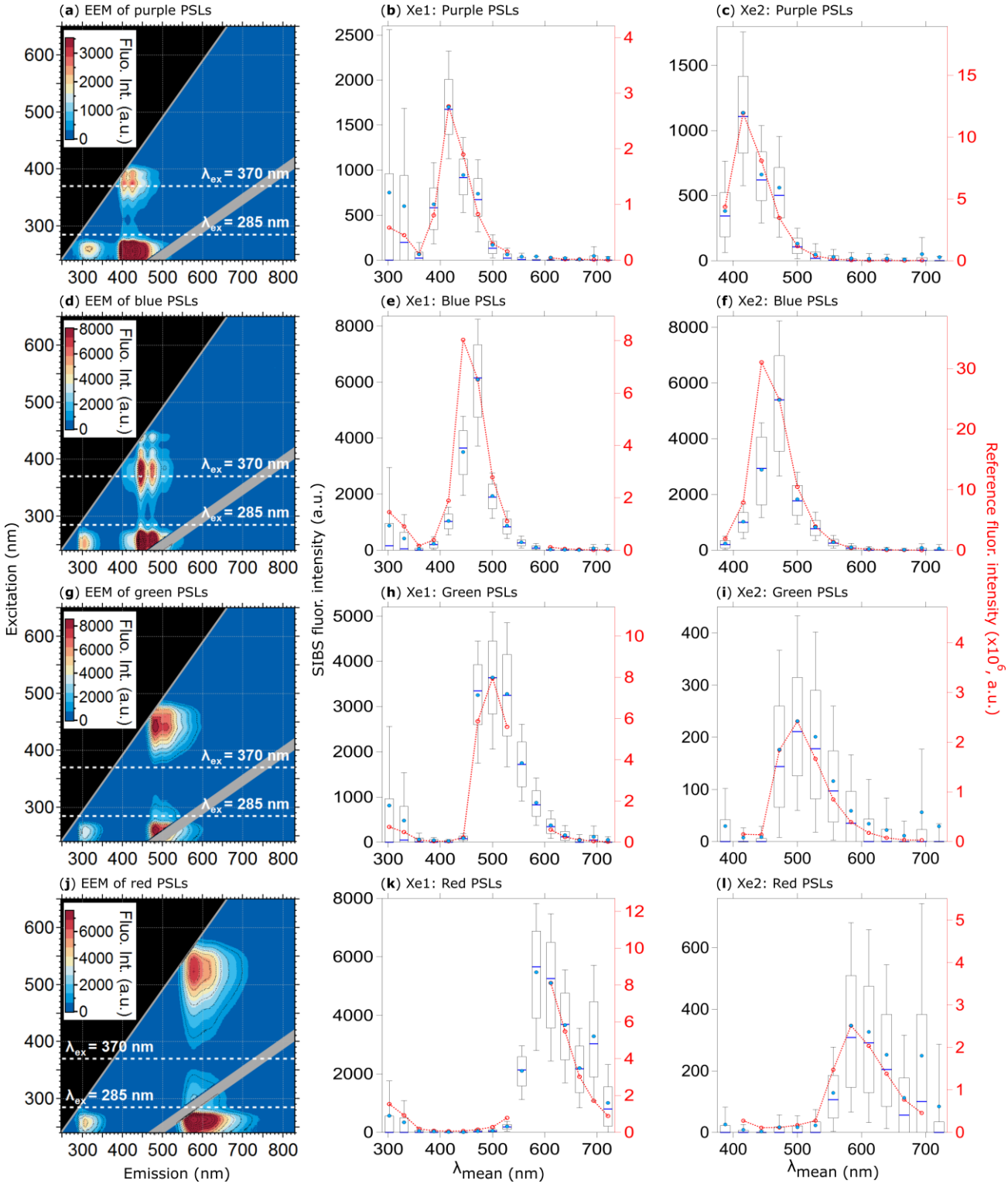

Figure 7. Fluorescence emission spectra of PSLs. Steady-state fluorescence signatures displayed as EEMs (left column) and spectra at Xe1 and Xe2 (middle, right columns) for: 2.07 µm purple (**a, b** and **c**, 1082 particles), 2.1 µm blue (**d**, **e** and **f**, 1557 particles), 2 µm green (**g**, **h**, and **i**, 1174 particles), and 2 µm red PSLs (**j**, **k,** and **l**, 1474 particles). Within EEMs: white dashed lines show SIBS excitation wavelengths (λ_{ex} = 285 and 370 nm), grey diagonal lines indicate 1st and 2nd order elastic scattering bands (both bands were subtracted automatically by the Aqualog V3.6 software). *Red dashed lines and markers (right axes; middle, right columns): averaged and re-binned reference spectra.*

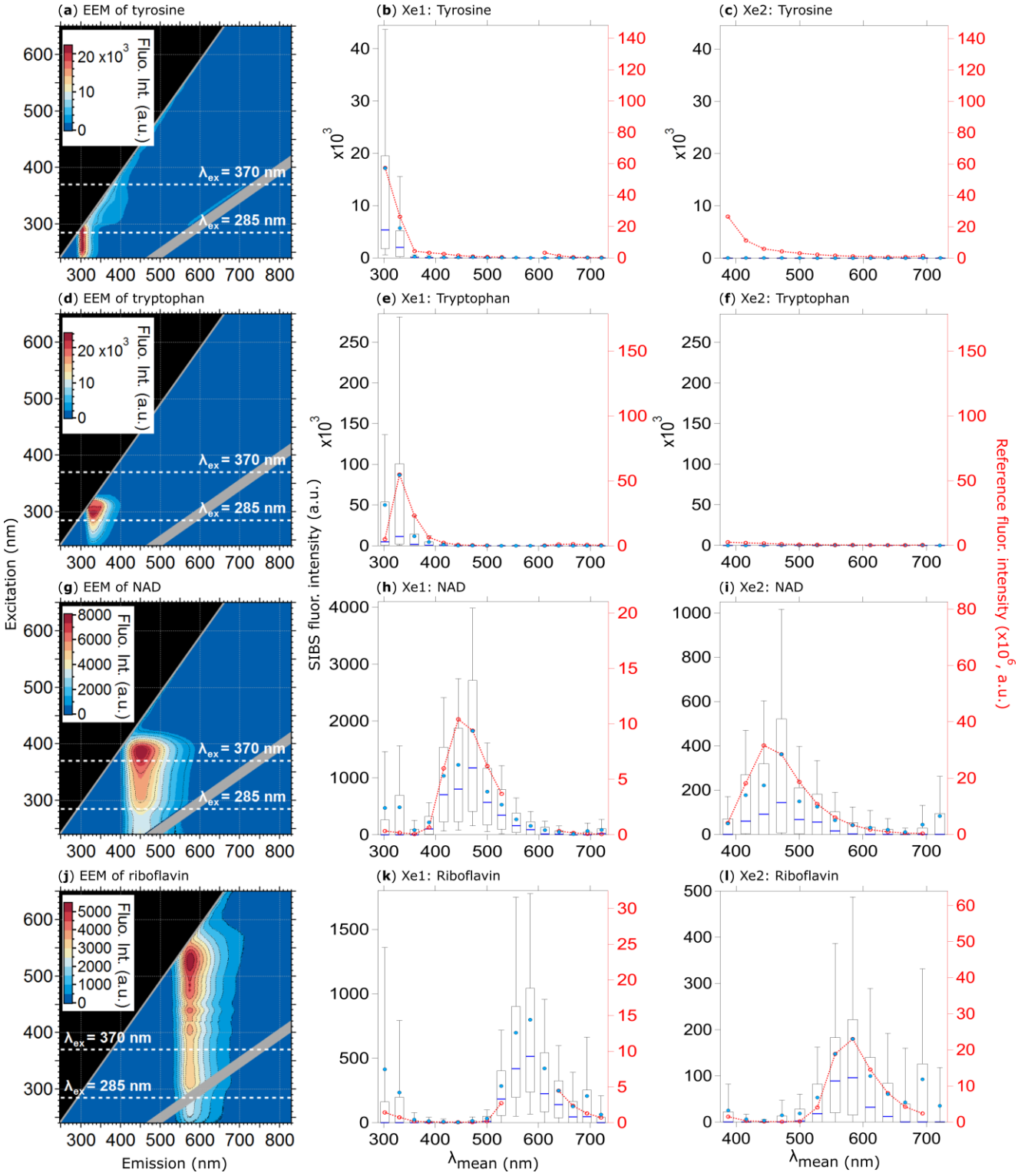

Figure 8. Fluorescence emission spectra of biofluorophores. EEMs (left column) and spectra at Xe1 and Xe2 wavelengths (middle and right columns) shown for: tyrosine (**a**, **b**, and **c**, 209 particles), tryptophan (**d**, **e**, and **f**, 193 particles), NAD (**g**, **h**, and **i**, 376 particles), and riboflavin (**j**, **k**, and **l**, 205 particles). *Red dashed lines and markers (right axes; middle, right columns): averaged and re-binned reference spectra.* All biofluorophores were size-selected between 1 and 2 µm.

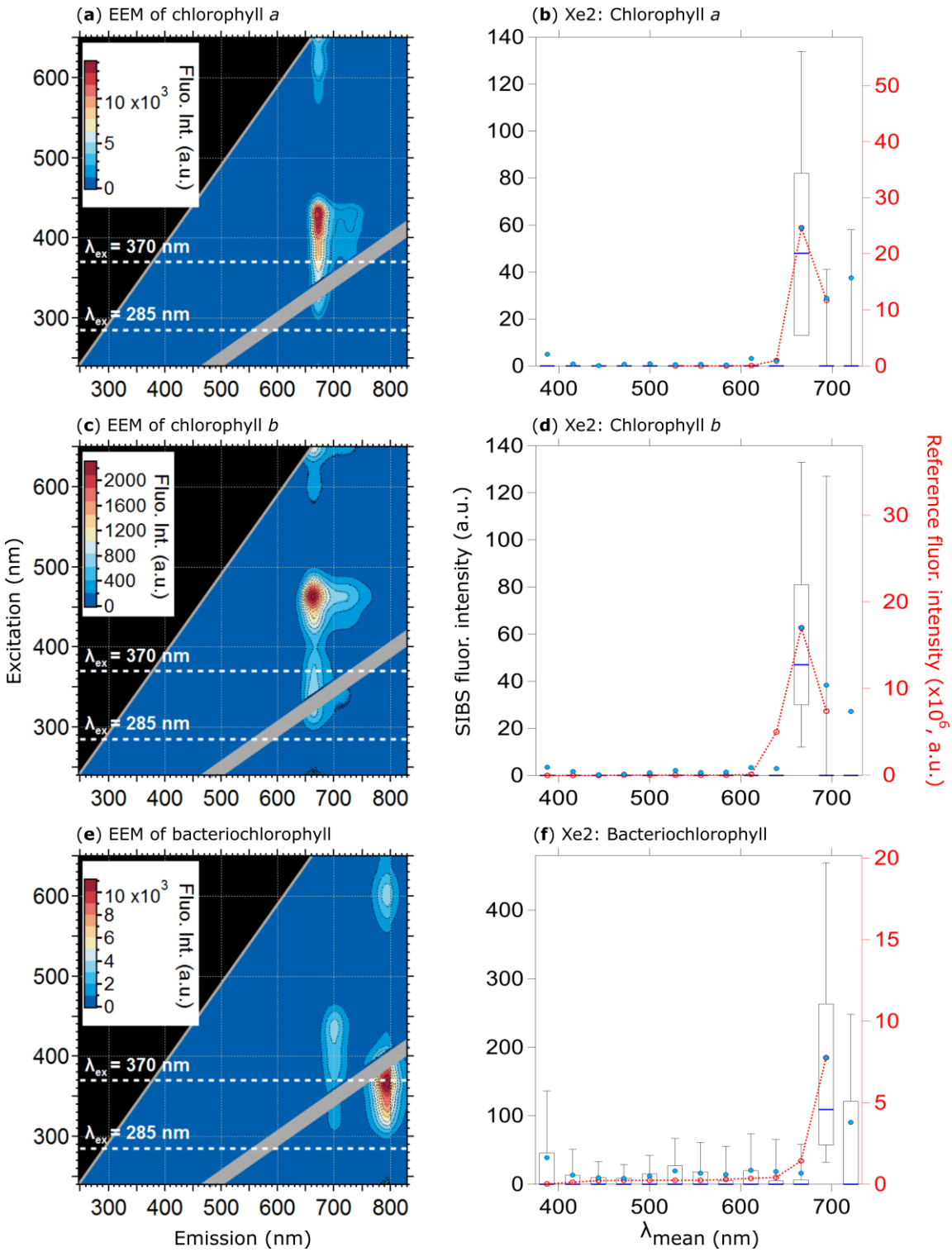

Figure 9. Fluorescence emission spectra of three chlorophyll types. Highlighted are EEMs (left column) and spectra at Xe2 (right columns) for: chlorophyll *a* (**a** and **b**, 370 particles), chlorophyll *b* (**c** and **d**, 585 particles), and bacteriochlorophyll (**e** and **f**, 633 particles). *Red dashed lines and markers (right axes; right column): averaged and re-binned reference spectra.* Size range chlorophyll *a* and *b*: 0.5 - 2 µm, size range bacteriochlorophyll: 0.5 - 1 µm. Emission spectra at Xe1 are excluded due to a fluorescence artifact caused by solved components from the polymer of the aerosolization bottles (Fig. S12).

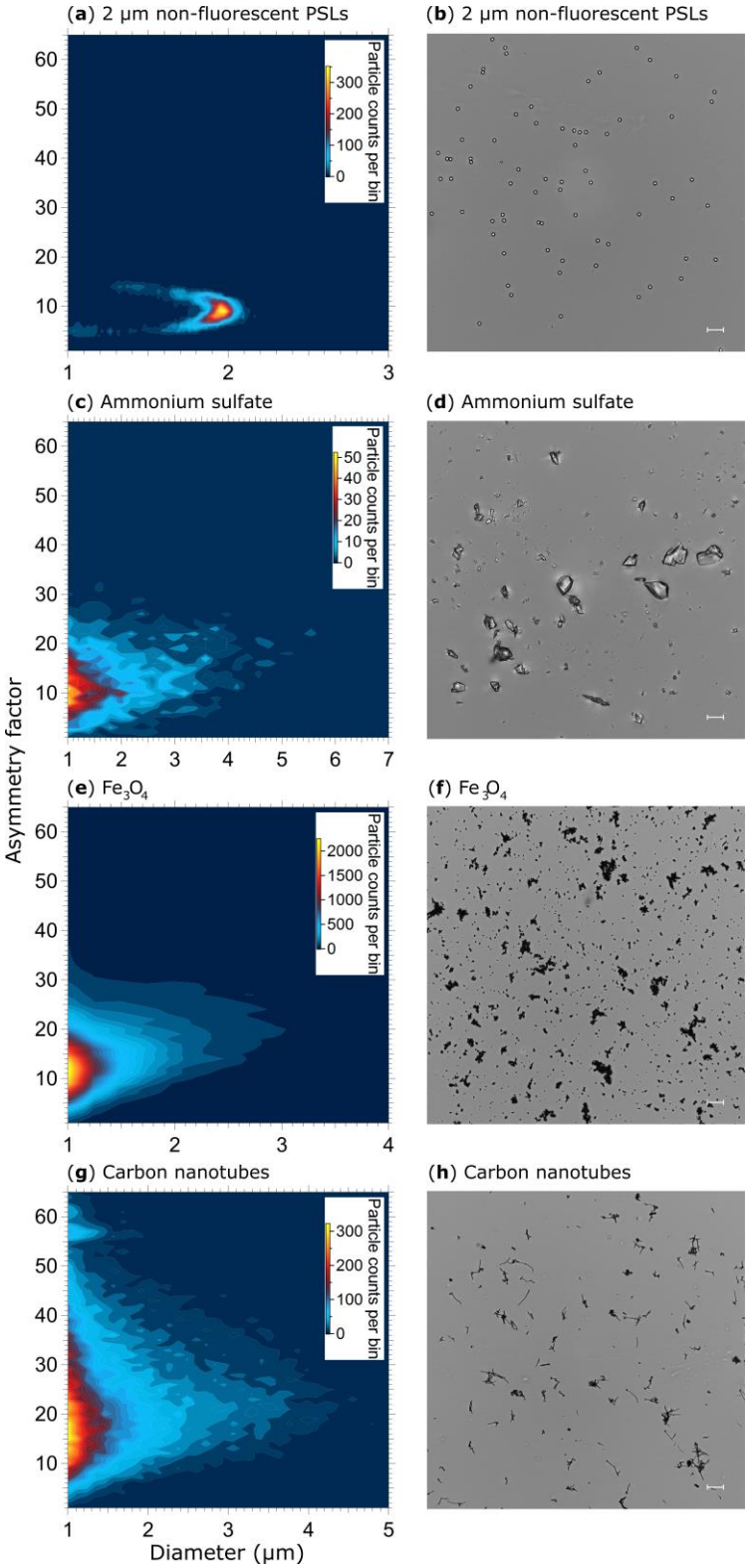

Figure 10. Particle asymmetry. Shown are particle density histograms (left column) and microscopy images (right column) for: 2 µm non-fluorescent PSLs (**a** and **b**, 17836 particles), ammonium sulfate (**c** and **d**, 3496 particles), Fe_3O_4 (**e** and **f**, 65097 particles), and carbon nanotubes (56949 particles, **g**). Scale bar (right column) indicates a length of 10 µm.

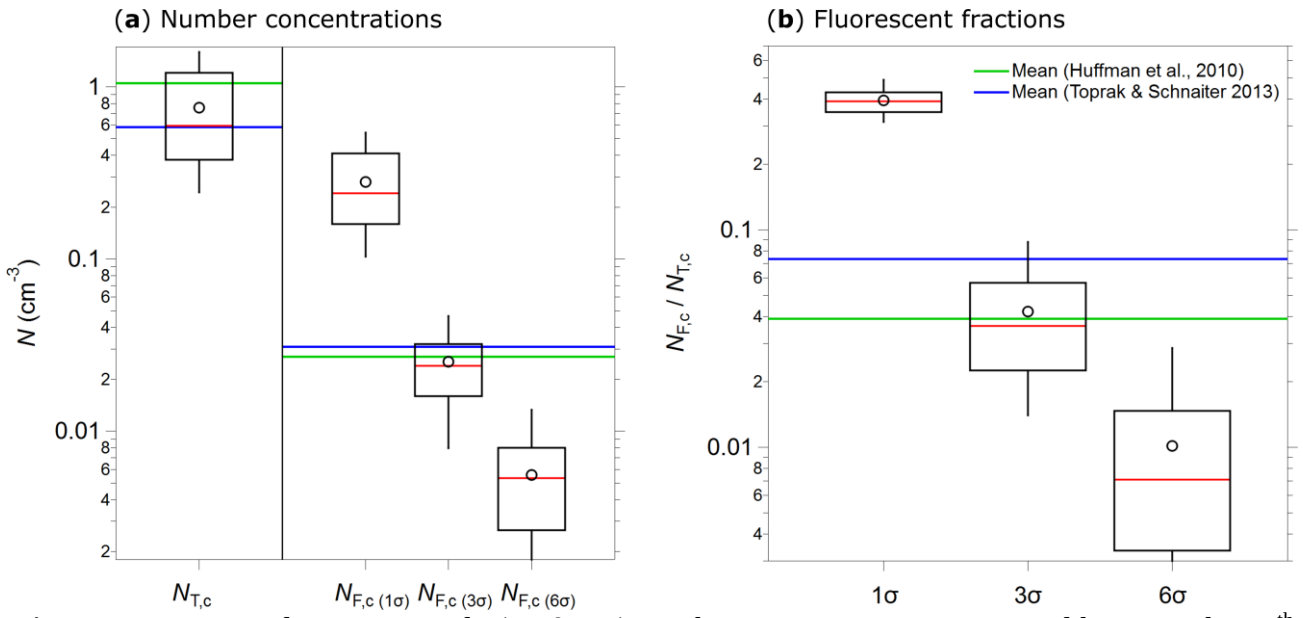

Figure 11. Integrated coarse particle (1-20 µm) number concentrations, measured between the 12th and 18th of April 2018 (5 min average), for total particles ($N_{T,c}$, fluorescent and non-fluorescent) and coarse fluorescent particles ($N_{F,c}$) after 1, 3, and 6σ SD background signal subtraction (**a**). The fluorescent fractions of integrated coarse particle number concentrations ($N_{F,c}$ / $N_{T,c}$) at 1, 3, and 6σ SD are shown in (**b**). Median (red line), mean (black circles), boxes 75 and 25 percentile, whiskers 95 and 5 percentile (**a** and **b**). Data from Huffman et al. (2010) (green lines) and Toprak & Schnaiter, (2013) (blue lines) were taken for comparison (**a** and **b**).

Supplement to

Spectral Intensity Bioaerosol Sensor (SIBS): An ~~new~~ Instrument for Spectrally Resolved Fluorescence Detection of Single Particles in Real-Time

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This file includes:

Supplementary Tables S1 to S2

Supplementary Figures S1 to S14

Table S1. Summary of physical properties of Polystyrene latex spheres (PSLs) and polystyrene-divenylbenzene particles (PS-DVB) used in this study. Stated properties are taken from manufacturer information. SD: Standard deviation, RI: Refraction index at 589 nm and 25˚C.

Diameter (µm)	SD (µm)	Confidence	RI	Material	Color / Dye	$\lambda_{ex}/\lambda_{em}$ (nm)	Provider	Catalog code
0.3	0.0148	CV= 5.1%	1.59	PSL	Non-fluorescent	Non-fluorescent	Polysciences Inc.	64015
0.356	0.014	CV= 3.9%	1.59	PSL	Non-fluorescent	Non-fluorescent	Polysciences Inc.	64016
0.4	0.0073	CV= 1.8%	1.59	PSL	Non-fluorescent	Non-fluorescent	Thermo-Fisher	3400A
0.5	0.0079	CV= 1.6%	1.59	PSL	Non-fluorescent	Non-fluorescent	Duke Scientific Corporation	3500A
0.53	N/A	N/A	1.59	PSL	Plum Purple / Proprietary	360 / 420	Bangs Laboratories Inc.	FS03F
0.6	0.010	CV= 1.7%	1.59	PSL	Non-fluorescent	Non-fluorescent	Thermo-Fisher	3600A
0.7	0.0083	CV= 1.2%	1.59	PSL	Non-fluorescent	Non-fluorescent	Thermo-Fisher	3700A
0.8	0.0083	CV= 1%	1.59	PSL	Non-fluorescent	Non-fluorescent	Thermo-Fisher	3800A
0.9	0.0041	CV= 0.5%	1.59	PSL	Non-fluorescent	Non-fluorescent	Thermo-Fisher	3900A
1	0.010	CV = 1.0%	1.59	PSL	Non-fluorescent	Non-fluorescent	Duke Scientific Corporation	4009A
1.6	0.020	CV= 1.3	1.59	PSL	Non-fluorescent	Non-fluorescent	Thermo-Fisher	4016A
2	0.021	CV= 1.0%	1.59	PSL	Non-fluorescent	Non-fluorescent	Thermo-Fisher	4202A
2	N/A	CV= < 5%	1.59	PSL	Red / Firefli™ Fluorescent Red	542 / 612	Thermo-Fisher	R0200
2	N/A	CV= < 5%	1.59	PSL	Green / Firefli™ Fluorescent Green	468 / 508	Thermo-Fisher	G0200
2	N/A	CV= < 5%	1.59	PSL	Blue / Firefli™ Fluorescent Blue	368,388,412 / 445, 445, 473	Thermo-Fisher	B0200B
2.07	0.15	N/A	1.59	PSL	Plum Purple / Proprietary	360 / 420	Bangs Laboratories Inc.	FS05F
3	0.032	CV=1.1%	1.59	PSL	Non-fluorescent	Non-fluorescent	Thermo-Fisher	4203A
4	0.04	CV = 1.0%	1.59	PSL	Non-fluorescent	Non-fluorescent	Thermo-Fisher	4204A
4.52	0.15	CV = 3.0%	1.59	PSL	Non-fluorescent	Non-fluorescent	Polysciences Inc.	17135
5	0.6	CV=11%	1.59	PS-DVB	Non-fluorescent	Non-fluorescent	Thermo-Fisher	DC-05
7	0.7	CV= 10%	1.59	PS-DVB	Non-fluorescent	Non-fluorescent	Thermo-Fisher	DC-07
8	0.8	CV= 10%	1.59	PS-DVB	Non-fluorescent	Non-fluorescent	Thermo-Fisher	DC-08
10	0.9	CV=9.2%	1.59	PS-DVB	Non-fluorescent	Non-fluorescent	Thermo-Fisher	DC-10
15	1.8	CV=11%	1.59	PS-DVB	Non-fluorescent	Non-fluorescent	Thermo-Fisher	DC-15
20	1.7	CV=8.9%	1.59	PS-DVB	Non-fluorescent	Non-fluorescent	Thermo-Fisher	DC-20

Table S2. Summary of reference particles used within this study. All biofluorophores, iron oxide (Fe_3O_4), and carbon nanotubes were purchased from Sigma-Aldrich, St. Louis, MO, USA. Ammonium sulfate was purchased from Fisher Scientific, Hampton, NH, USA.

Reference particles	**CAS Nr.**
Bacteriochlorophyll	17499-98-8
Chlorophyll *a*	479-61-8
Chlorophyll *b*	519-62-0
NAD	606-68-8
Riboflavin	83-88-5
Tryptophan	73-22-3
Tyrosine	556-02-5
Fe_3O_4	1317-61-9
Carbon nanotubes	308068-56-6
Ammonium sulfate	7783-20-2

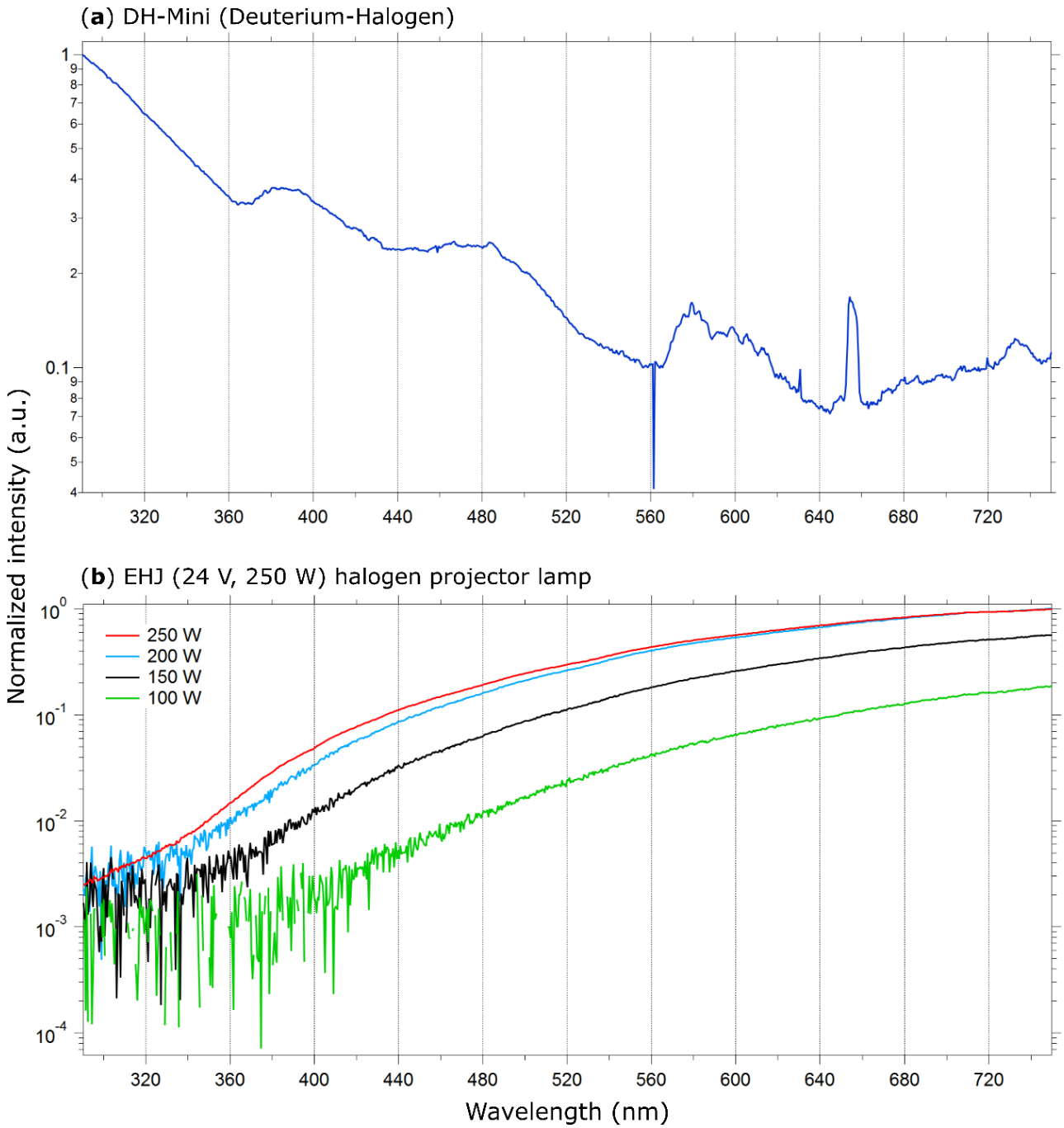

Figure S1. Normalized and averaged calibration lamp spectra. In (**a**), the spectrum of a deuterium-halogen lamp (DH-Mini, Ocean Optics) is shown, in (**b**) the spectra of a halogen projector lamp (EHJ 24V250W, Ushio), both measured with the Dual-FL spectrometer (Horiba).

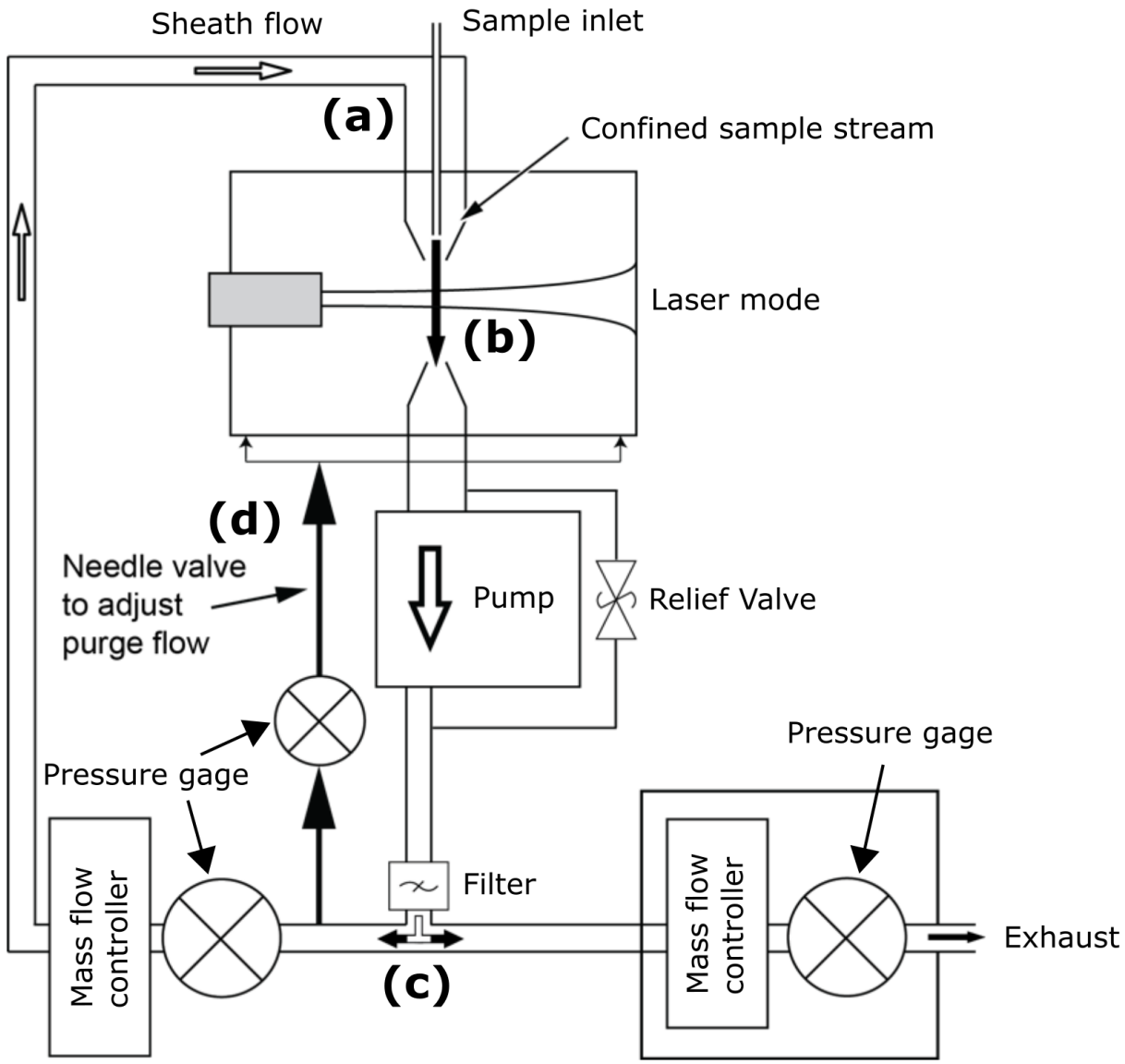

Figure S2. Flow diagram for the SIBS. Aerosol is drawn via a tapered delivery nozzle (**a**) into the optical cavity. The intersection of sample flow and laser beam defines a sampling volume with approximately 0.7 mm in diameter and 130 µm depth (**b**). The sheath flow is filtered through a HEPA filter and recirculates in the system (**c**). A small purge flow, which is adjusted by a needle valve (**d**), constantly purges the optical cavity (Modified, image courtesy: DMT).

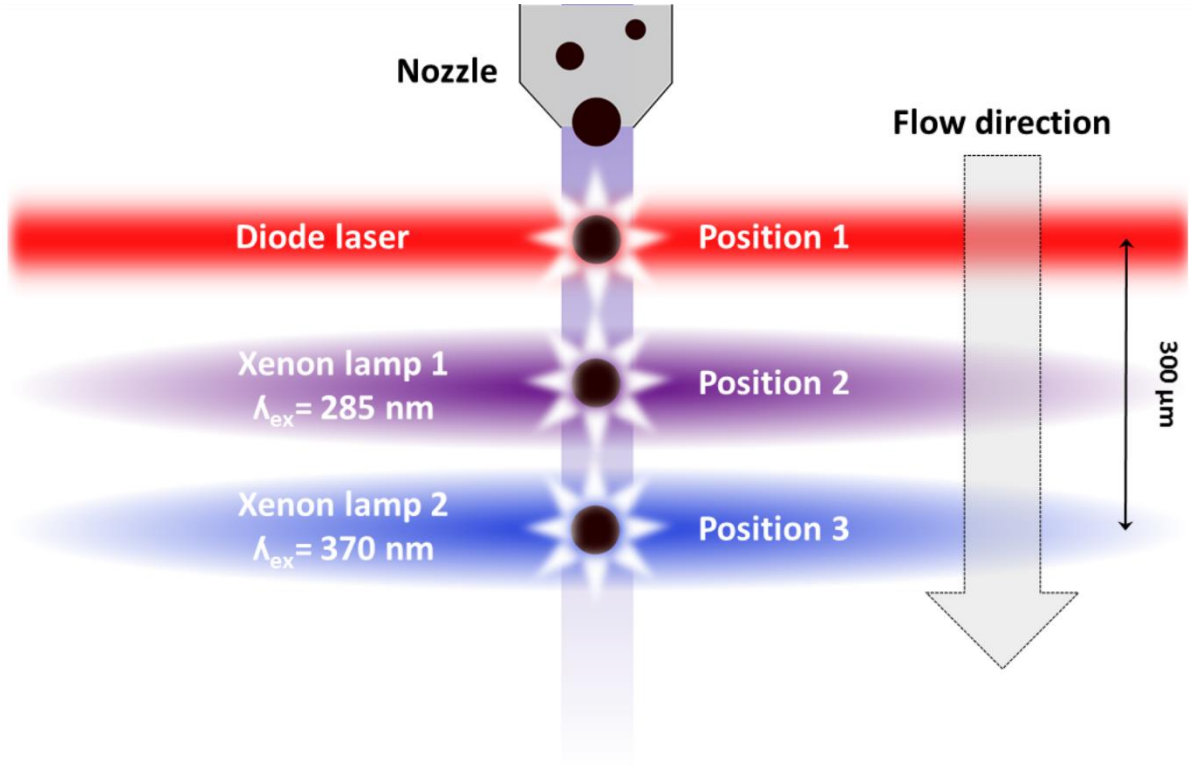

Figure S3. Schematic diagram of particle size and fluorescence detection. Position 1: Particles scatter light in all directions after being illuminated by a diode laser (λ = 785 nm). Position 2: Xenon lamp 1 is firing at λ_{ex} = 285 nm. Position 3: Xenon lamp 2 is firing at λ_{ex} = 370 nm. The measurement cycle from position 1 to position 3 takes ~25 µs over a distance of ~300 µm. (Modified, adapted from WIBS-4A service manual (DOC-0345 Rev A), DMT; 2012).

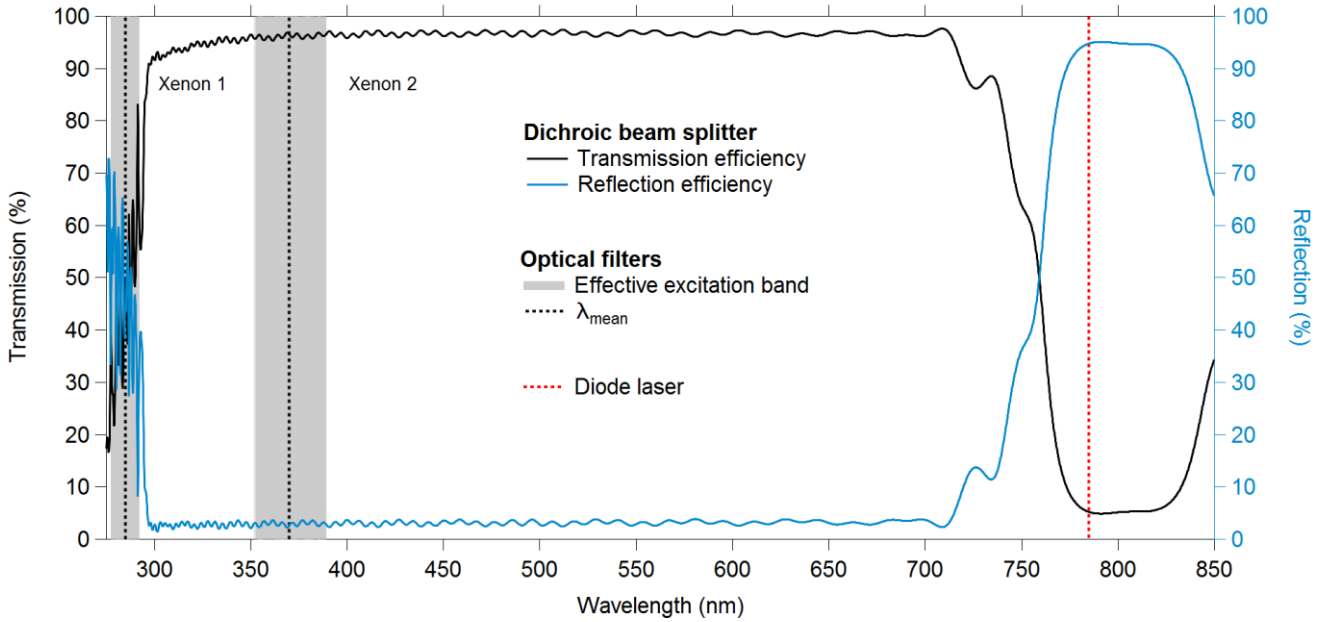

Figure S4. Transmission and reflection efficiency of the dichroic beam splitter. The beam splitter transmits fluorescence emission (black line) to the grating polychromator and reflects scattering light (blue line) to the particle sizing- and detection PMT. (Data courtesy: Semrock).

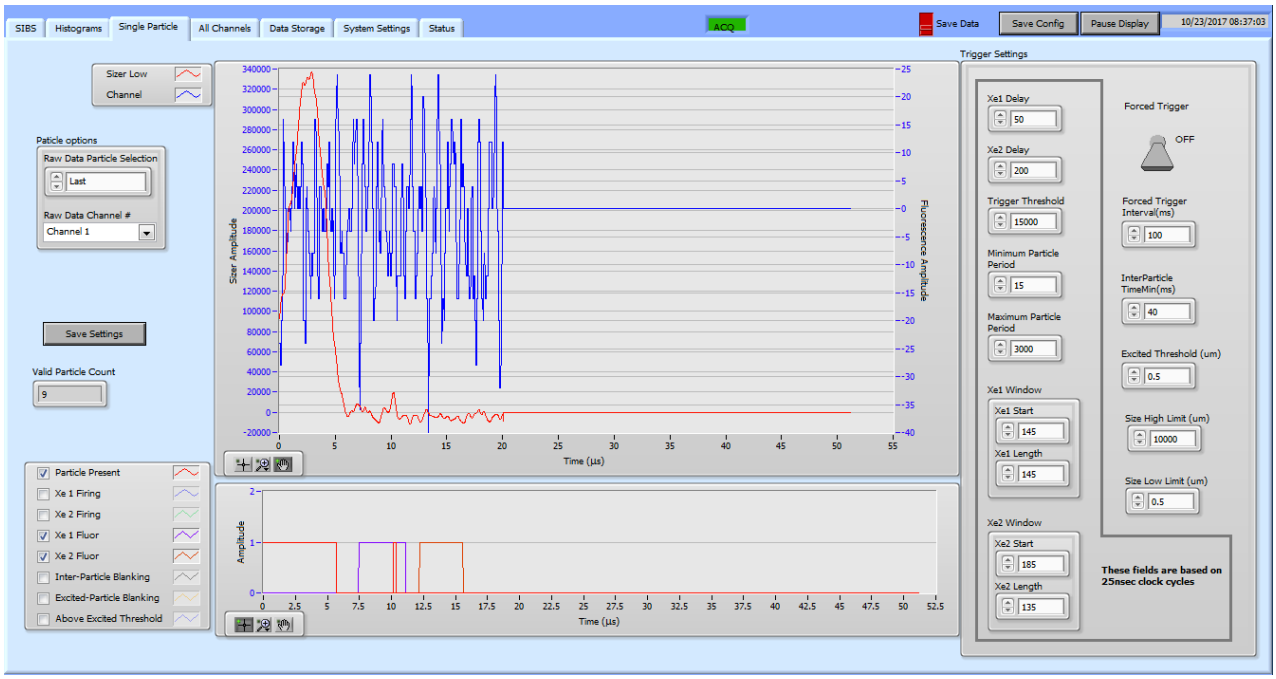

Figure S5. "Single Particle" tab of the SIBS user interface.

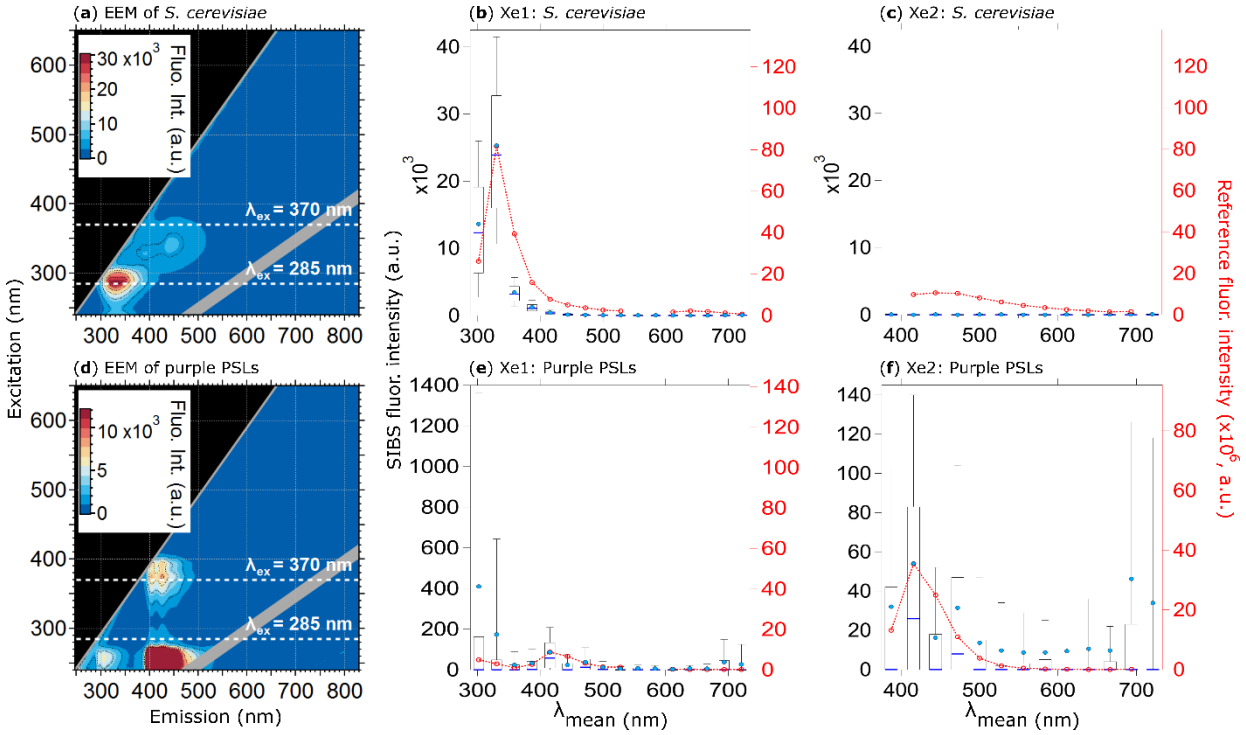

Figure S6. Corrected fluorescence emission of *S. cerevisiae* and 0.53 µm purple PSLs. Steady-state fluorescence signatures displayed as EEMs (left column) and spectra at Xe1 and Xe2 (middle, right columns) for: *S. cerevisiae* (**a, b** and **c**, size range between 4 - 10 µm, 1057 particles), and 0.53 µm purple PSLs (**d**, **e,** and **f**, 5260 particles). Within EEMs: white dashed lines show SIBS excitation wavelengths (λ_{ex} = 285 and 370 nm), grey diagonal lines indicate 1st and 2nd order elastic scattering bands (both bands were subtracted automatically by the Aqualog V3.6 software). *Red dashed lines and markers (right axes; middle, right columns): averaged and re-binned reference spectra.*

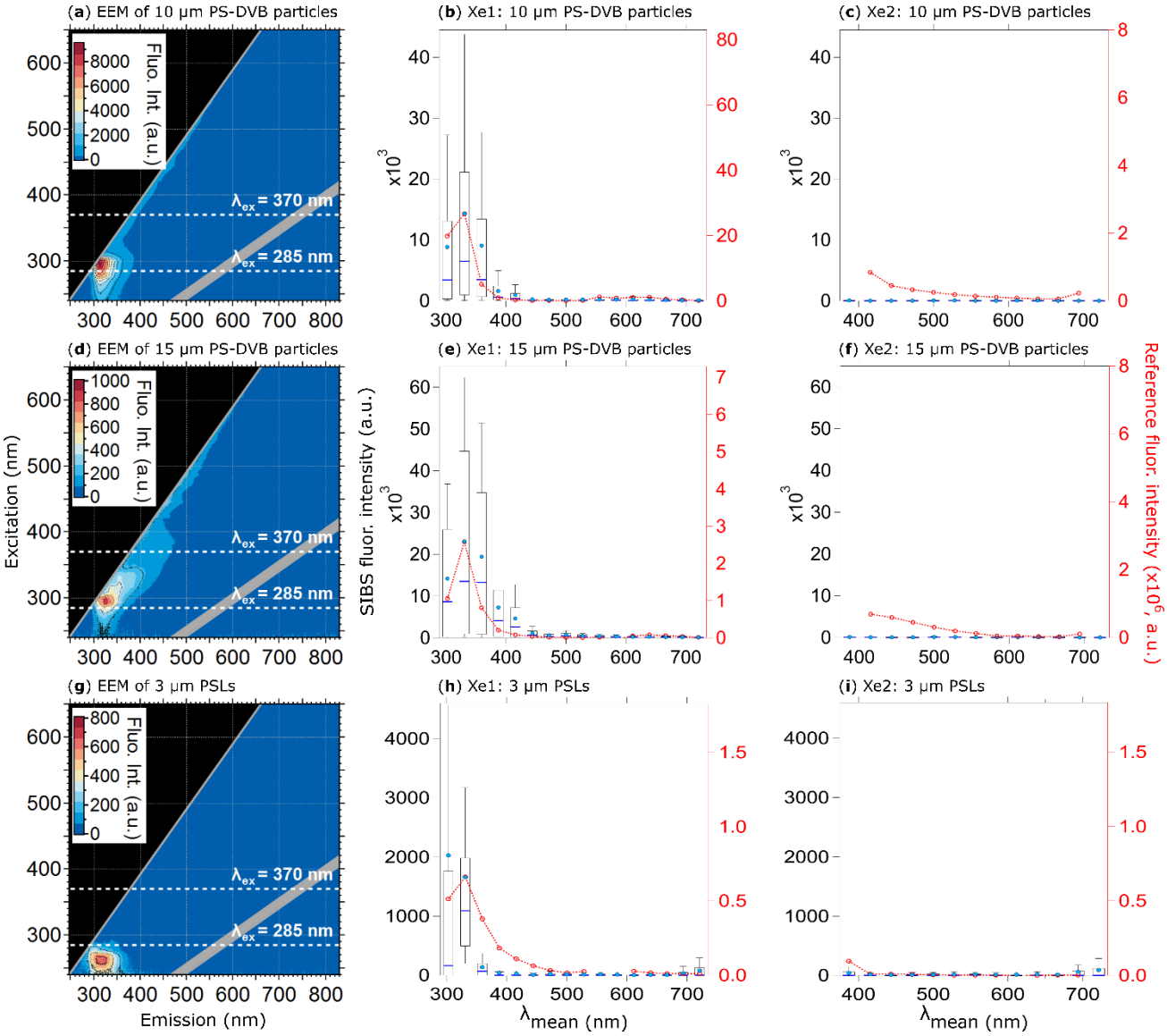

Figure S7. Fluorescence signatures of non-fluorescent particles. Highlighted are EEMs (left column) and spectra at Xe1 and Xe2 (middle, right columns) for: 10 µm (**a**, **b,** and **c**, uncorrected, 367 particles) and 15 µm (**d**, **e,** and **f**, uncorrected, 400 particles) PS-DVB particles, and 3 µm PSLs (**g**, **h,** and **i**, corrected, 2396 particles). *Red dashed lines and markers (right axes; middle, right columns): averaged and re-binned reference spectra.*

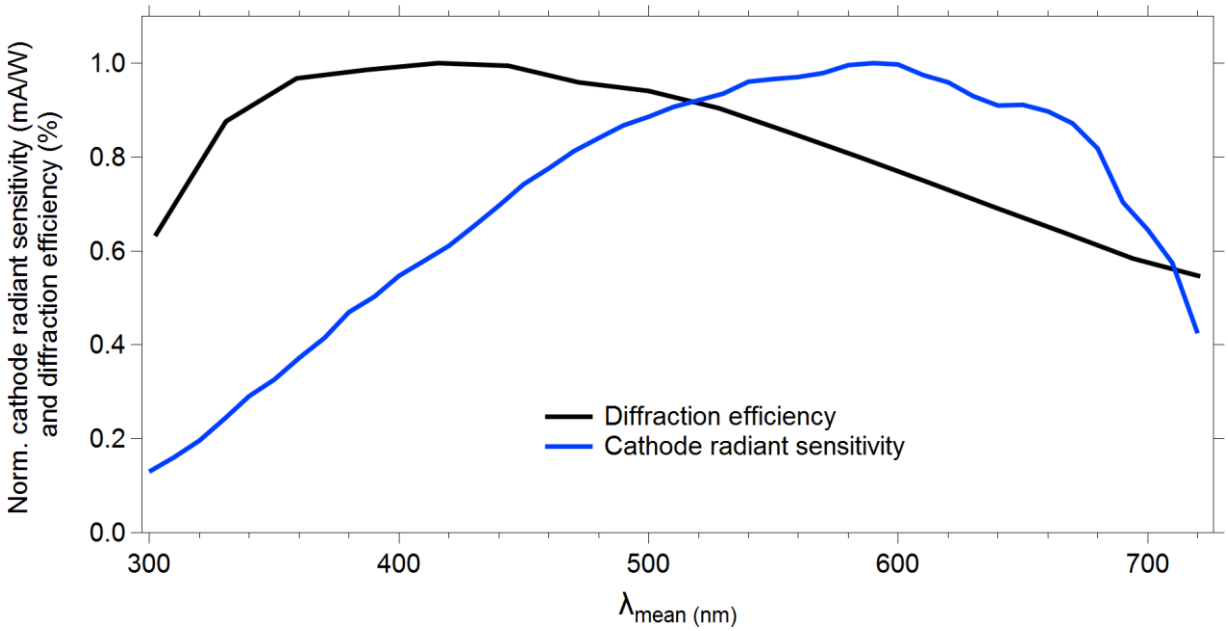

Figure S8. Normalized cathode radiant sensitivity of the PMT and diffraction efficiency of the grating. The cathode radiant sensitivity multiplied by the diffraction efficiency results in the theoretical detector responsivity shown in Figure 6. (Data courtesy: Hamamatsu).

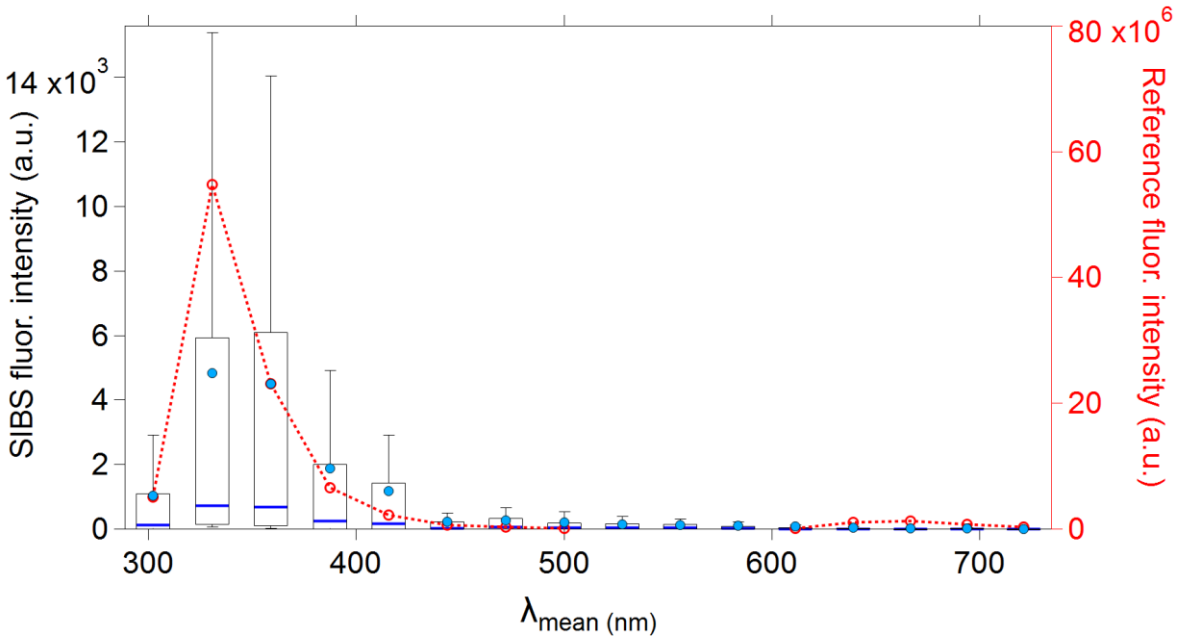

Figure S9. Uncorrected fluorescence spectra of tryptophan at Xe1 in a size range between 1 – 2 µm. *Red dashed line and markers (right axes): averaged and re-binned reference spectra.*

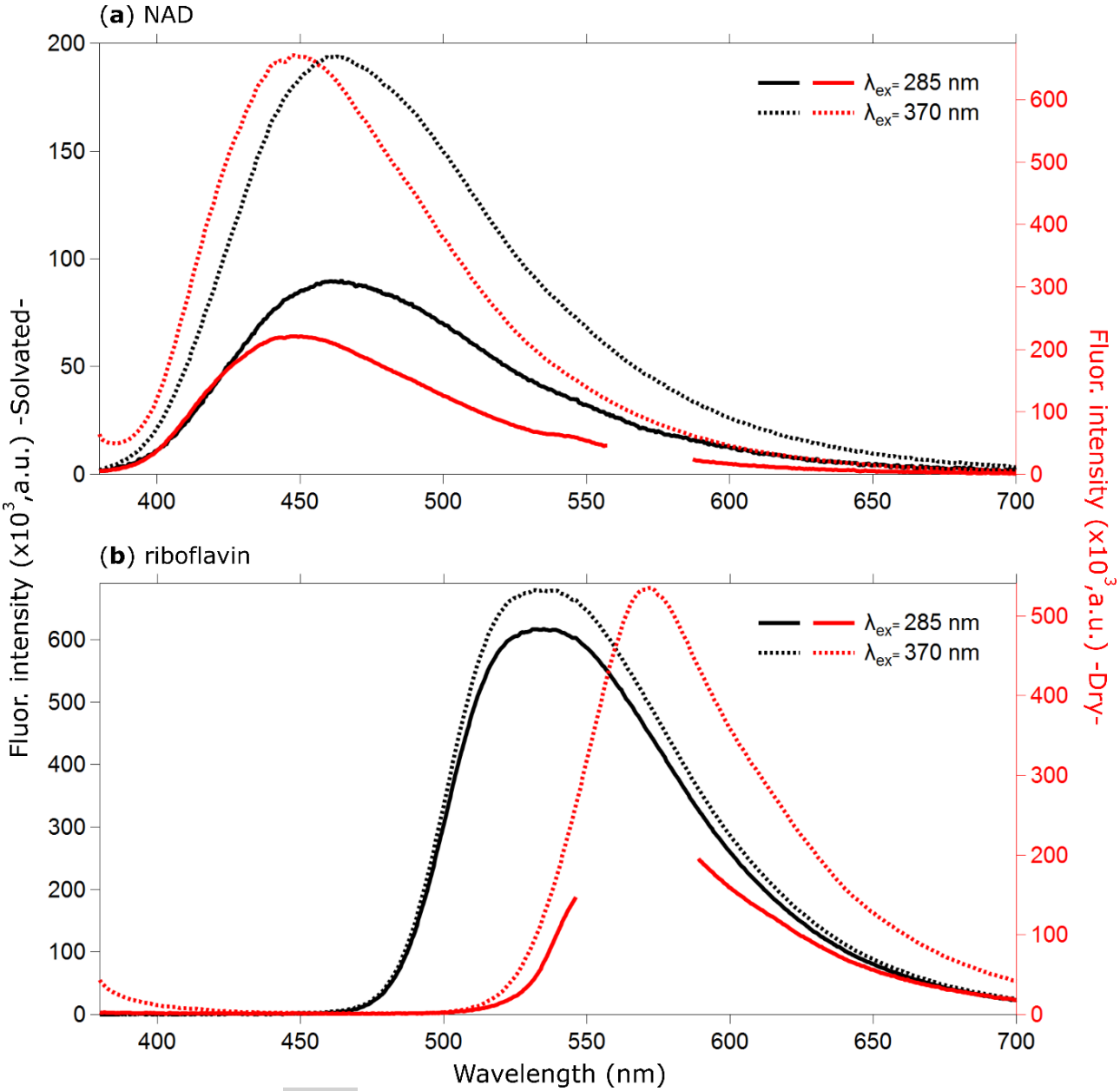

Figure S10. Dry vs. solv*ateded*. Shown are reference spectra for NAD (**a**) and riboflavin (**b**) in dry and solv*ateded* state. Data coinciding with 2nd order elastic scattering were removed (***a and*** **b**, red solid line). Peak maxima: NAD (dry): ~448 nm, NAD (solv*ateded*): ~463 nm, riboflavin (dry): ~572 nm, riboflavin (solv*ateded*): ~535 nm.

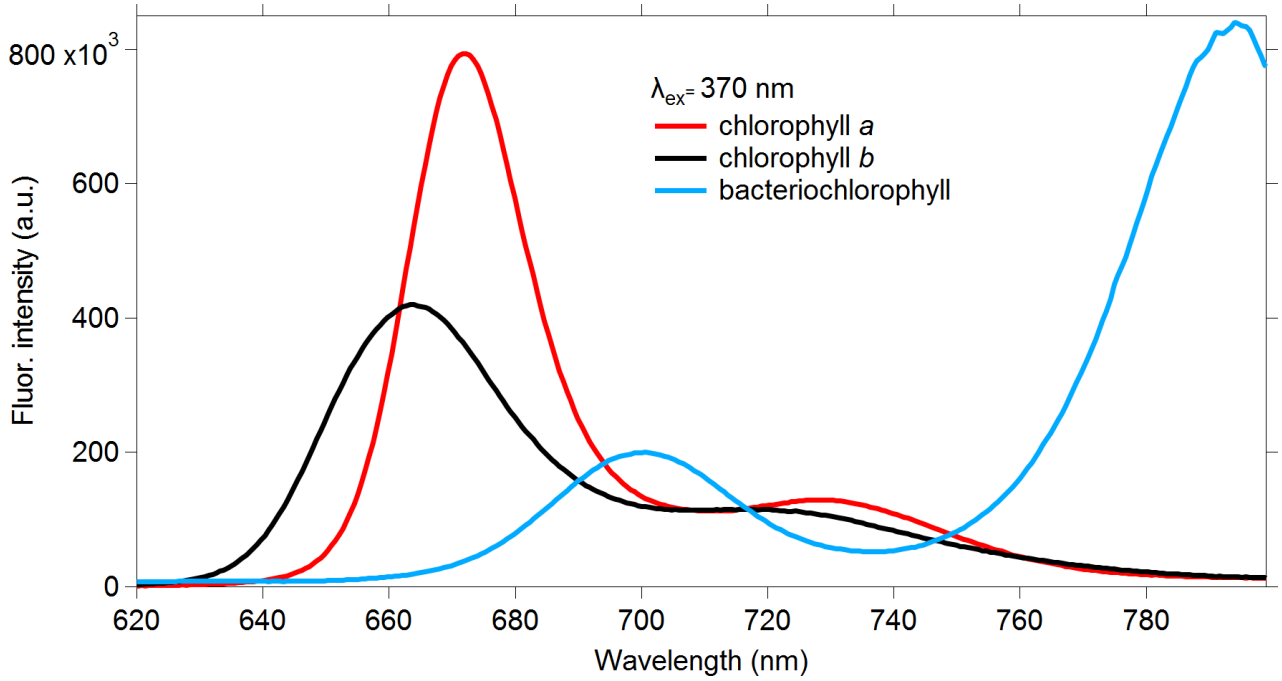

Figure S11. Fluorescence spectra of different chlorophyll types. Shown are reference spectra for chlorophyll *a*, *b*, and bacteriochlorophyll at λ_{ex} = 370 nm.

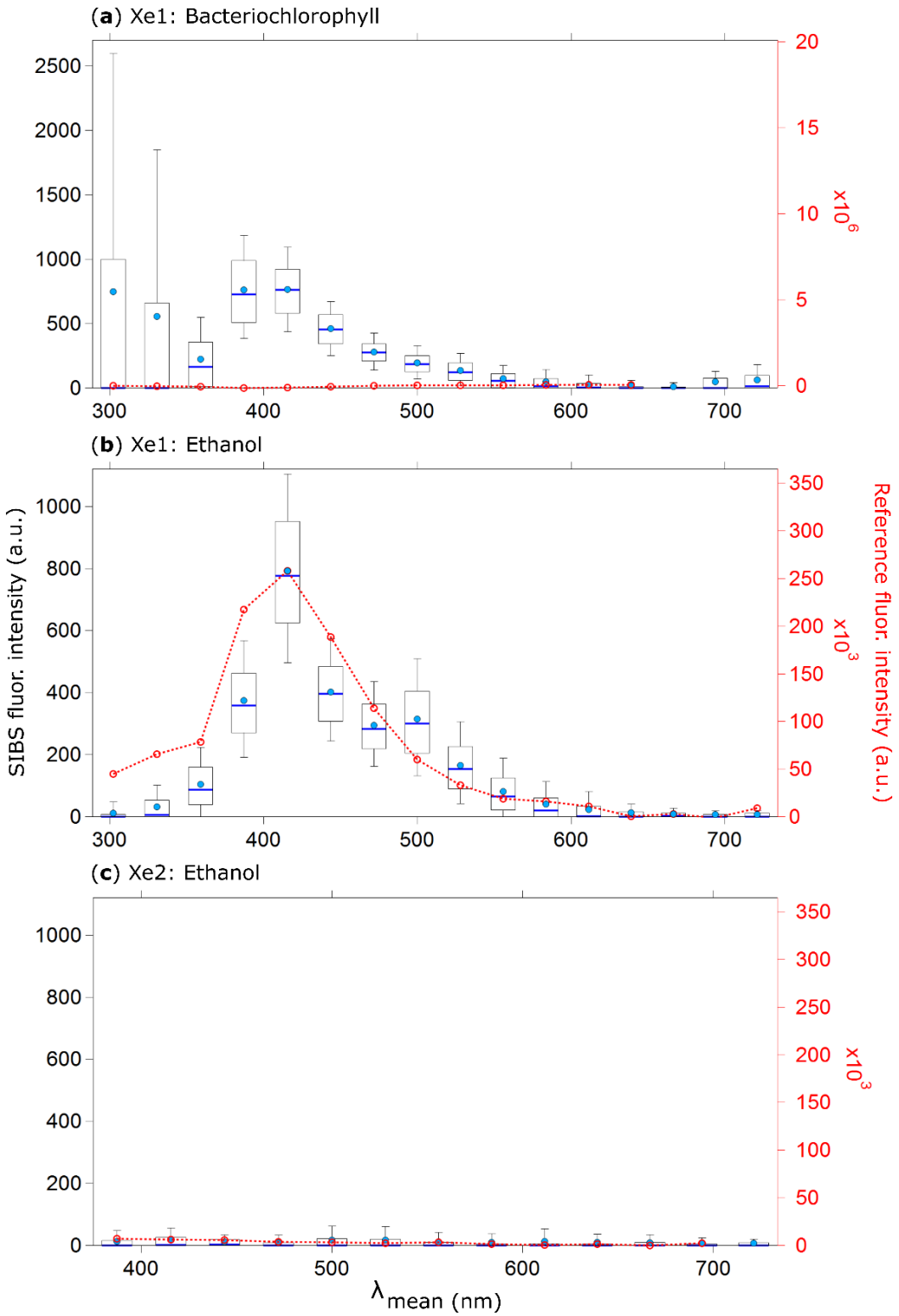

Figure S12. Fluorescence spectra of ethanol artefact. Highlighted are fluorescence spectra of bacteriochlorophyll at Xe1 (**a**) and uncorrected spectra of ethanol, after being vortexed for 15 min in nebulizer plastic bottles, at Xe1 (**b**) and Xe2 (**c**). *Red dashed lines and markers (right axes;): averaged and re-binned reference spectra.* Since no distinct fluorescence signal is detectable at Xe2 (**c**), the fluorescence emission of chlorophyll *a*, *b* and bacteriochlorophyll is considered to be unaffected.

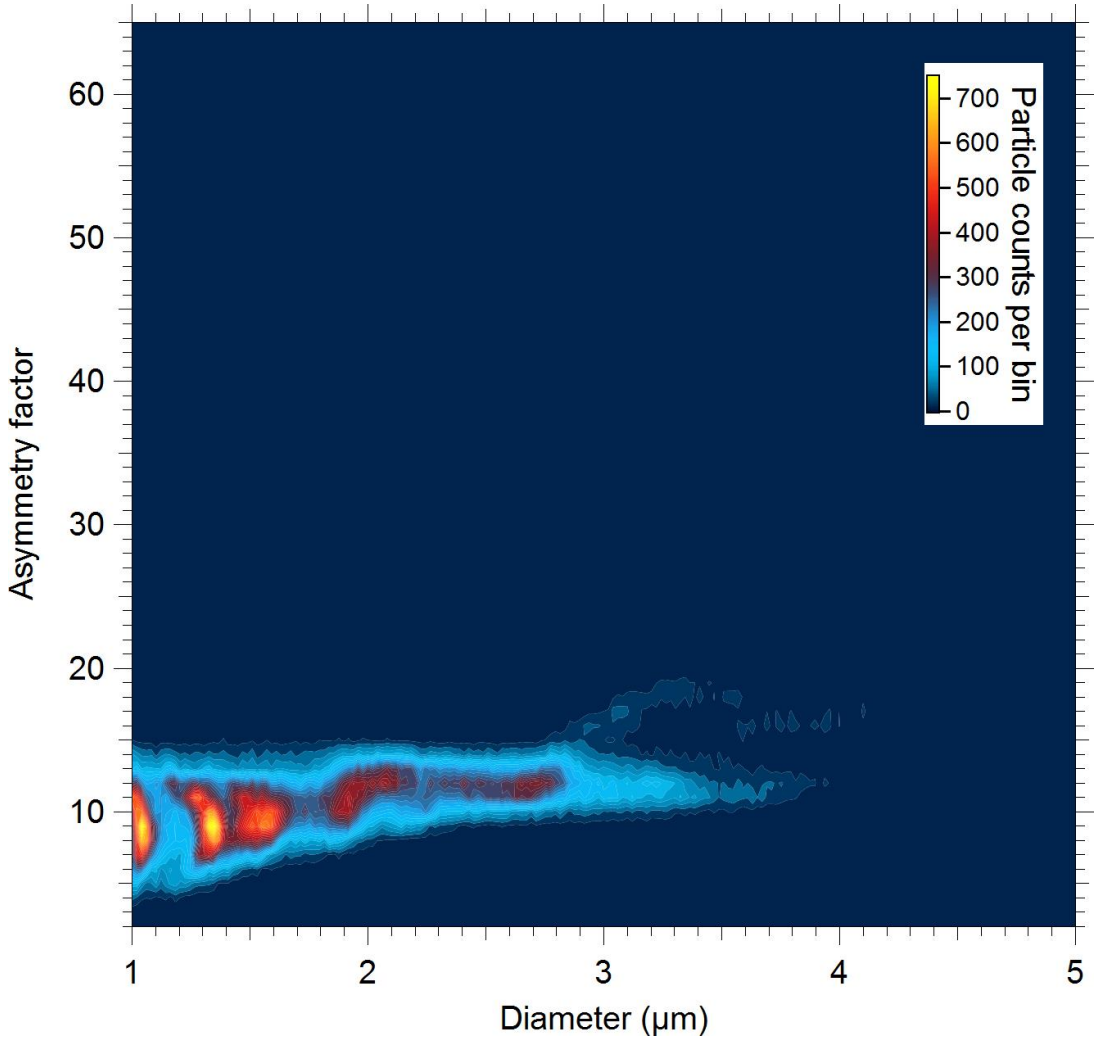

Figure S13. Particle asymmetry of ultrapure water droplets (163178 particles) displayed as particle density histogram.

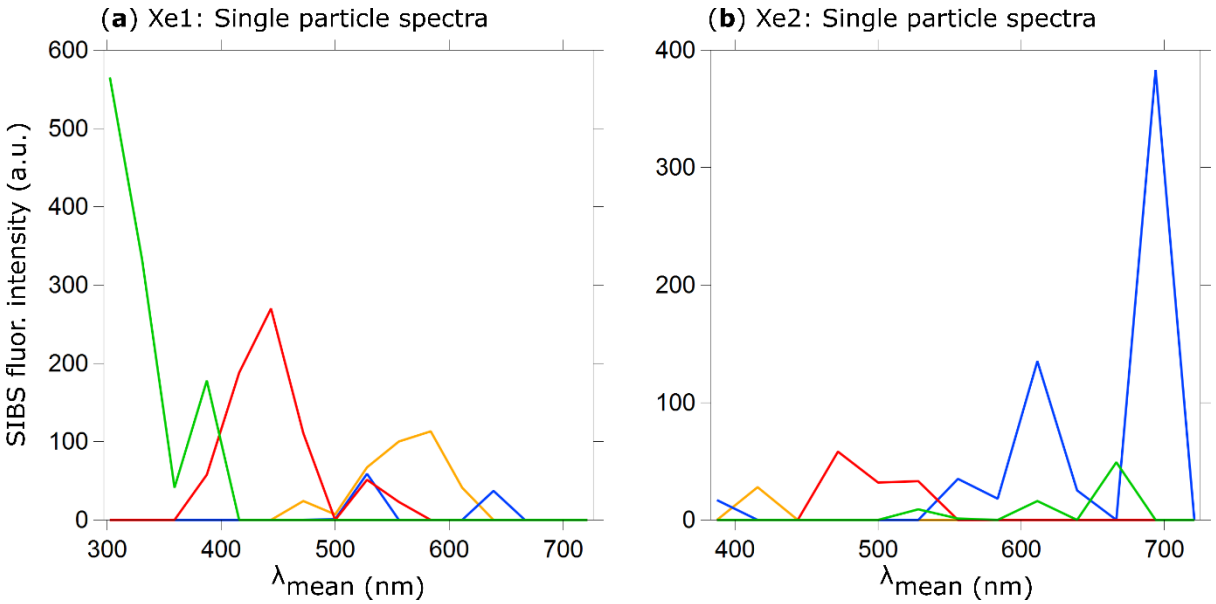

Figure S14. Exemplary fluorescence spectra of single ambient particles at Xe1 (**a**) and Xe2 (**b**).