We greatly value the careful reading and the detailed comments provided by the referees. The responses to the comments of the referees in our direct reply (shown below) and within the revised manuscript (see marked copy) are provided below. The pages and lines indicated below correspond to those in the marked copy.

Response to Referee 1 (Referees' comments are italicized)

1. Referee comment: "*Many experimental details and clarifications are needed to explain and justify this new technique. The discussion of the contribution of these organic acid contributions to WSOC and the many figures that detail organic acid mixing ratios should be pared down, since the species identification here are speculative, and there may be many interferences that have not been adequately considered.*"

Author response: We agree that the assignment of some of the organic acids, with the exception of formic and acetic acids, are speculative. As such, we have revised the manuscript to be more circumspect about the organic acid contribution to $WSOC_g$. Please refer to our response to comment 7.

We have also added more experimental details and discussions on interferences in the revised manuscript. Please refer to our responses to comments on experimental details and interferences below.

2. Referee comment: "*Previous reports have noted that SF6- is a viable reagent ion only for dewpoints below -20 C. The flow tube is at lower pressure here, and that is said to minimize interferences with water vapor (line 167). This needs to be explained much more thoroughly. The very high dilution with dry nitrogen likely does much more to reduce the water vapor interferences, but this is never discussed. This high dilution must reduce sensitivity. Please discuss the trade-off between reduced sensitivity and increased selectivity. Does the high dilution compromise time response, since there is a very small flow of ambient air into the flow tube? Does the large flow of N2 through the ion source increase reagent ion signal?*"

Author response: The referee is correct in stating that there are tradeoffs between reduced sensitivity and increased selectivity when the sampled air is diluted by a large volume of N_2/SF_6 in the CIMS flow tube. The high flow (3.7 slpm) of N_2/SF_6 passed through the ion source increased the SF_6^- reagent ion signal. This does increase sensitivity but the N_2 flow also dilutes the sampled air flow in the flow tube which reduces the sensitivity. We chose these conditions to bring the water interferences to an acceptable level. We don't see significant differences in the time response for sample air flow flows $\geq$ 2 L min^{-1} introduced into the flow tube because a large flow (7 L min^{-1}) is maintained in the sample inlet right up to the CIMS flow tube (Fig. 1). As requested, we have added more discussion on the tradeoffs between reduced sensitivity and increased selectivity when the sampled air is diluted by a large volume of N_2 in the CIMS flow tube and how the high dilution impacts time response in the revised manuscript:

Page 13 line 389: "The 0.3 L min^{-1} sampled air flow is diluted by 3.7 slpm of N_2/SF_6 flow in the flow tube. The ratio of the sampled air flow to the N_2/SF_6 flow introduced into the flow tube is approximately 1:13. While the high N_2/SF_6 flow (3.7 slpm) passed through the radioactive source into the flow tube increased the SF_6^- reagent ion signal, the high dilution

of the sampled air flow in the flow tube reduced the CIMS instrument sensitivity by decreasing the number density of the analytes."

3. Referee comment: "*The flows as shown don't make sense, and volume flow and mass flow don't appear to be correctly distinguished. Accurately describing and discussing the flows is critical, since it is the altered gas flows that have made SF6- viable in a humid environment. Line 154 gives the sampled air flow as a volume flow, but the orifice is said to maintain a constant mass flow. A 220 l/min scroll pump is used to pump a very small flow of 4 L/min through the flow tube, as shown in Figure 1. I suspect that the N2 flow should be written as 3.7 slpm, rather than L/min. But even that doesn't make sense, since it would be ~300 L/min volume flow, and greater than the pump speed. The instrument is said to be similar to Liao et al 2011, but the flows are very different, and these should be discussed completely.*"

Author response: The referee is correct in pointing out that our previously stated flows and pump rates were incorrect. A 367 L min^{-1} scroll pump (Edward nXDS 20i) was used. The N_2/SF_6 flow was 3.7 slpm. These corrections have been made to the revised manuscript:

Page 6 line 166: "The CIMS instrument was comprised of a series of differentially pumped regions: a flow tube, a collisional dissociation chamber, an octopole ion guide, a quadrupole mass filter and an ion detector. These sections were evacuated by a scroll pump (Edward nXDS 20i), a drag pump (Adixen MDP 5011) and two turbo pumps (Varian Turbo-V301), respectively. Ambient air was drawn continuously into the flow tube. A flow of 3.7 slpm of N_2 containing a few ppm of SF_6 (Scott-Marrin Inc.) was passed through a ^{210}Po ion source into the flow tube. SF_6^- anions, which were produced via associative electron attachment in the ^{210}Po ion source, reacted with the sampled ambient air in the flow tube to generate analyte ions."

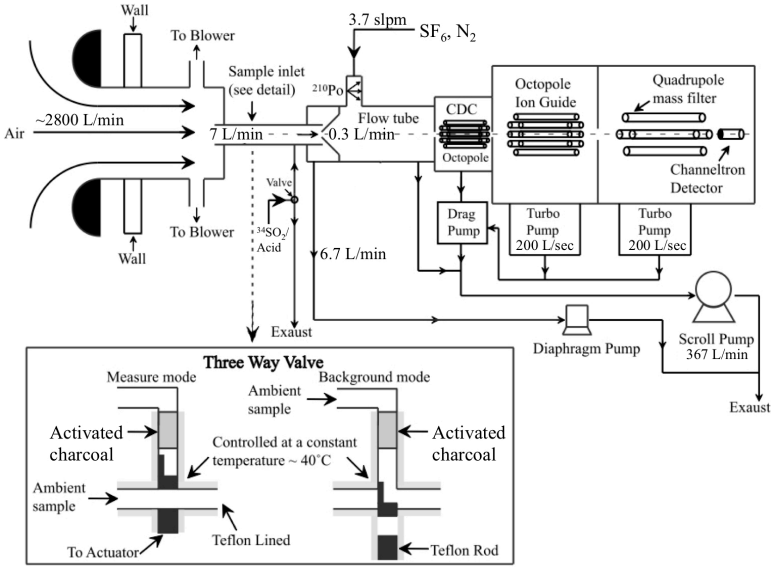

Figure 1: The CIMS instrument and inlet configuration used in the field study. The automated three-way sampling valve is shown in the inset. The figure was adapted from Liao et al. (2011).

4. Referee comment: "*Please state and demonstrate time response, and preferably show a*

calibration and a zero. Detection limits are given for 2.5 min. I don't understand the relation between the 2.5 min back- ground and the 0.04 s duty cycle for each mass (line 946). Please describe the measurement frequency - how often is a measurement made? I'm confused as to whether these are 0.04 s measurements, 2.5 min measurements, or 1 hr measurements (as used in the figures). And please be specific about the integration time for the mixing ratios. For example, are the range of mixing ratios in the abstract (line 22) for 1 hr averages?"

Author response: The dwell time for each m/z ion was set to 0.5 s and measurements of these ions were obtained every ~13 s. These measurements were averaged to generate 1-hour averaged mixing ratios of the various gases. Mixing ratios presented in this paper are the 1-hour averaged mixing ratios. We have added details on the frequency of measurements and stated explicitly that the data reported are 1-hour averages in the revised manuscript:

Page 7 line 192: "Ions monitored during the field study included m/z 45, 59, 65, 73, 75, 79, 82, 87, 89, 101, 102, 103, 108, 117, 131 and 148. The assignment of these ions will be discussed in section 3. The dwell time for each m/z ion was set to 0.5 s and measurements of these ions were obtained every ~13 s, which resulted in a ~4 % (= 0.5/13 x 100 %) duty cycle for each ion monitored. The data presented in this paper was averaged to 1-hour intervals unless stated otherwise."

The author is correct in stating that the range of mixing ratios reported in the abstract are the 1-hour averaged values. This is clarified in the revised manuscript:

Page 1 line 21: "1-hour averaged ambient concentrations of organic acids ranged from a few parts per trillion by volume (ppt) to several parts per billion by volume (ppb)."

Each background measurement period lasted ~4 min. However, ion signals for the different organic acids took up to 1.5 min to stabilize during the switch between ambient, calibration and background measurements during the field study. Hence, ion signals obtained during the first 1.5 min were not included in the calculation of the average and standard deviation of ion signals measured during background mode. This resulted in a 2.5 min (= 4 min – 1.5 min) integration time period for background measurements. The duty cycle (0.04 or 4 %) was calculated from the dwell time of each m/z ion monitored (0.5 s) and the measurement frequency (13 s). We clarified how detection limits were calculated from the background measurements in the revised manuscript:

Page 9 line 257: "The detection limits of the organic acids were estimated as 3 times the standard deviation values (3σ) of the ion signals measured during background mode. Although each background measurement period lasted ~4 min, ion signals of the different organic acids took up to 1.5 min to stabilize during the switch between ambient, calibration and background measurements during the field study. Thus, ion signals measured during the first 1.5 min were not included in the calculation of the average and standard deviation of ion signals measured during background mode. Table 1 summarizes the average detection limits of the organic acids for 2.5 min averaging periods which corresponds to the length of a background measurement with a 4 % duty cycle for each m/z."

As requested, we have added a section discussing the CIMS instrument time response and a figure showing a calibration and background measurement in the revised manuscript:

Page 14 line 426: "3.1.3. Background and calibration measurements

Figure S4 shows an example of the CIMS instrument response during the switch between background, calibration and ambient measurements of formic and acetic acids during the field study. The 13 s time resolution data was used to determine the CIMS instrument time response. Formic (m/z 45, 65 and 108) and acetic (m/z 79) acid ion signals took ~1.5 min to reach a steady state after switches between ambient, calibration and background measurements (Figs. S4a and S4c).

The decays in the formic and acetic acid ion signals and times required for them to reach steady state after the removal of calibration gases during the switch from standard addition calibration to ambient sampling were used to determine the CIMS response time. The signal decays were fitted using double exponential functions. For formic acid, the m/z 45, 65 and 108 ion signals decayed to $1/e^2$ in 37 ± 2, 33 ± 2 and 32 ± 2 s, respectively (Fig. S4b). For acetic acid, the m/z 79 ion signal decayed to $1/e^2$ in 42 ± 2 s (Fig. S4d).”

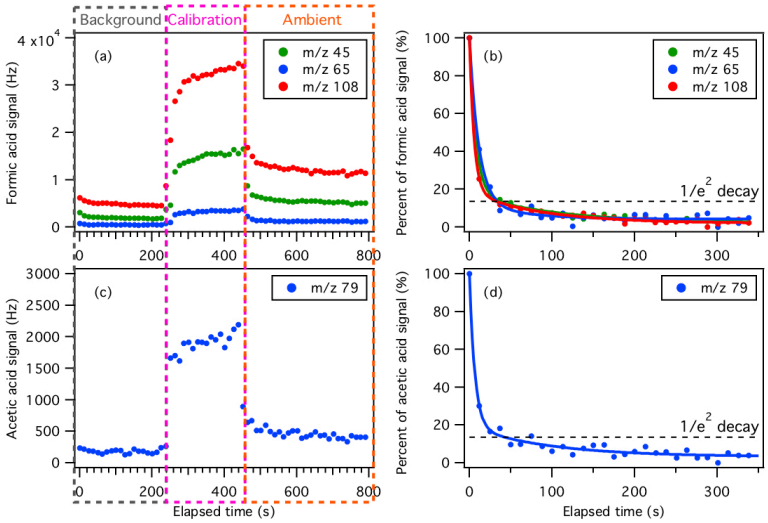

Figure S4: Example of the CIMS instrument response during switches between background, calibration and ambient measurements of (a) formic, and (c) acetic acids. Panels (b) and (d) show the percent of formic and acetic acid ion signals after the removal of a 6.75 ppb of formic acid and 5.87 ppb of acetic acid standard addition calibration as a function of time. All the data shown here are 13 s time resolution data. Double exponential fits to each m/z ion are shown as colored solid lines. Black dashed lines show the times for the ions to decay to $1/e^2$.

5. Referee comment: “*Detection limits and abundances in the figures are written in ppb, but for most cases, ppt would be a more appropriate unit and much easier to read.*”

Author response: We have made the requested changes in the revised manuscript.

6. Referee comment: “*Giving sensitivities relative to 34SO2 is confusing and seems unnecessary, especially since that is a different ion-molecule reaction than all the organic acid reactions studied here and it has a water dependence. Providing absolute sensitivities will be much more useful for anyone who wants to compare this ion chemistry with others. And please discuss how these sensitivities compare to other techniques.*”

Author response: As requested, we have provided the absolute sensitivities of organic acids and discussed how these sensitivities compare with other CIMS techniques in the revised manuscript:

Page 12 line 363: "Table 1 shows a summary of the sensitivities of X^- and X^-•HF ions of organic acids. The average sensitivities of the $HCOO^-$ (m/z 45) and $HCOO^-$•HF (m/z 65) ions of formic acid were 1.29 ± 0.22 and 0.29 ± 0.05 Hz ppt^{-1}, respectively, while the average sensitivities of the CH_3COO^- (m/z 59) and CH_3COO^-•HF (m/z 79) ions of acetic acid were 1.46 ± 0.29 and 0.30 ± 0.06 Hz ppt^{-1}, respectively. A weak ^{210}Po ion source (< 1 mCi) was used by SF_6^--CIMS instrument during the field study, hence these sensitivities will be substantially higher if a stronger radioactive source is used. Nevertheless, these sensitivities are compared to formic and acetic acid sensitivities measured by a high-resolution time-of-flight chemical ionization mass spectrometer (Aerodyne Research Inc.) that utilized I^- reagent ions during the field study. Although the formic acid sensitivity measured by I^--CIMS (1.33 ± 0.28 Hz ppt^{-1}) was comparable to that measured by SF_6^--CIMS, the acetic acid sensitivity measured by I^--CIMS (< 0.1 Hz ppt^{-1}) was substantially lower than that measured by SF_6^--CIMS. Previous studies have similarly reported low acetic acid sensitivity measured by I^--CIMS (Aljawhary et al., 2013; Lee et al., 2014)."

References:

Aljawhary, D., Lee, A. K. Y., and Abbatt, J. P. D.: High-resolution chemical ionization mass spectrometry (ToF-CIMS): application to study SOA composition and processing, Atmospheric Measurement Techniques, 6, 3211-3224, 10.5194/amt-6-3211-2013, 2013.

Lee, B. H., Lopez-Hilfiker, F. D., Mohr, C., Kurten, T., Worsnop, D. R., and Thornton, J. A.: An Iodide-Adduct High-Resolution Time-of-Flight Chemical-Ionization Mass Spectrometer: Application to Atmospheric Inorganic and Organic Compounds, Environmental Science & Technology, 48, 6309-6317, 10.1021/es500362a, 2014.

Table 1: Summary of organic acids of interest, their detection limits and sensitivities of their X^- and X^-•HF ions[a]

Organic Acid	Detection limit (ppt)[b]	Sensitivity (Hz ppt^{-1})	
		X^-	X^-•HF
Formic acid	30	1.29 ± 0.22	0.29 ± 0.05
Acetic acid	60	1.46 ± 0.29	0.30 ± 0.06
Oxalic acid	1	6.38 ± 0.32	0.97 ± 0.05
Butyric acid	30	0.41 ± 0.01	0.12 ± 0.004
Glycolic acid	2	5.53 ± 0.11	1.64 ± 0.03
Propionic acid	6	2.05 ± 0.02	1.26 ± 0.01
Valeric acid	10	0.76 ± 0.008	0.35 ± 0.004

[a]Only organic acids with calibration measurements are shown.
[b]Detection limits are approximated from 3 times the standard deviation values (3σ) of the ion signals measured during background mode. Shown here are the average detection limits of the organic acids for 2.5 min averaging periods which corresponds to the length of a background measurement at a 4 % duty cycle for each mass.

7. Referee comment: "*Please discuss the many possible interferences to the organic acids listed.*

The discussion of calibrations for many compounds was valuable, but then compounds for which calibrations weren't successful (pyruvic, for example) were dismissed. It appears arbitrary to ignore compounds if the calibration didn't work. Those other compounds almost certainly contribute to the signals and make the calculations of WSOC fraction extremely unreliable. For example, the signals at oxalic likely are dominated by or include contributions from lactic acid (both mass 90), butyric includes contributions from pyruvic (both mass 88), propionic includes contributions from glyoxylic acid (both mass 74). The authors correctly note that the mass assignments are speculative. It is OK to speculate, but from then on it would be appropriate to report mixing ratios only as upper limits for that compound and then forego calculations that rely on quantifying abundance. The sensitivities of the reported compounds span a wide range (over an order of magnitude). Thus, if the signal at one of the reported masses comes instead from an isomer with a much higher or lower sensitivity, the organic acid abundance would be drastically over or under estimated. There needs to be considerably more work to justify the WSOC contributions."

Author response: The comparison of the total carbon contributed by the organic acids to the $WSOC_g$ serves as a zeroth order check that the assignment of ion peaks to the different organic acids are plausible. We agree that the assignment of some ion peaks are speculative, hence we are more circumspect in our discussion of the carbon mass fraction of $WSOC_g$ comprised of organic acids in the revised manuscript:

Page 19 line 573: "To estimate the fraction of $WSOC_g$ that is comprised of organic acids, the total organic carbon contributed by formic, acetic, oxalic, butyric, glycolic, propionic and valeric acids is compared to the $WSOC_g$ measurements. We emphasize that the ion peak assignment of some of these organic acids are speculative. Hence, this comparison primarily serves as a zeroth order check to determine if the peak assignments are plausible. Figures 6a and 6b show the time series and diurnal profiles of $WSOC_g$ and the organic carbon contributed by the measured organic acids. Formic and acetic acids comprised majority of the total organic carbon contributed by the measured organic acids (study averages of 31 and 38 %, respectively). Assuming that the ion peak assignments are correct and the measured organic acids are completely water-soluble, the carbon mass fraction of $WSOC_g$ comprised of these organic acids ranged from 7 to 100 %."

Figure 6: (a) Time series of $WSOC_g$ and the total organic carbon contributed by the measured organic acids. All the data are displayed as 1-hour averages. (b) Diurnal profiles of $WSOC_g$ and the total organic carbon contributed by the measured organic acids. Also shown are the diurnal profiles of the organic carbon contributed by the individual measured organic acids. All the concentrations represent the mean hourly averages and the standard errors are plotted as error bars. (c) Scatter plot of total organic carbon contributed by the measured organic acids with $WSOC_g$. Note that the ion peak assignment to some of these organic acids are speculative.

8. Referee comment: "*I don't understand the count rates and normalization shown in figures 6-8. How do the count rates get to be small fractions of 1 Hz? Please use a unit that is more accurate and describe how the values are determined.*"

Author response: The data for malonic, succinic and glutaric acids are presented as the ratio of their ion signals (Hz) to the instrument's sensitivity to $F_2{}^{34}SO_2$ (Hz ppb^{-1}) since these organic acids

were not calibrated. The ion signals of these organic acids are 1 to 2 orders of magnitude smaller than the sensitivity of the $F_2{}^{34}SO_2^-$ ion (study-averaged sensitivity = 2928 ± 669 Hz ppb^{-1}), resulting in these ratios to be less than 1. We have changed the units in the figures and stated more explicitly how these values were obtained in the revised manuscript:

Page 18 line 551: "The time series of ion signals (Hz) of malonic, succinic and glutaric acids normalized to the instrument's sensitivity to $F_2{}^{34}SO_2$ (Hz ppb^{-1}) are shown in Fig. S3. The ion signals of these organic acids are 1 to 2 orders of magnitude smaller than the sensitivity of the $F_2{}^{34}SO_2^-$ ion (study-averaged sensitivity = 2928 ± 669 Hz ppb^{-1}), resulting in these ratios to be less than 1."

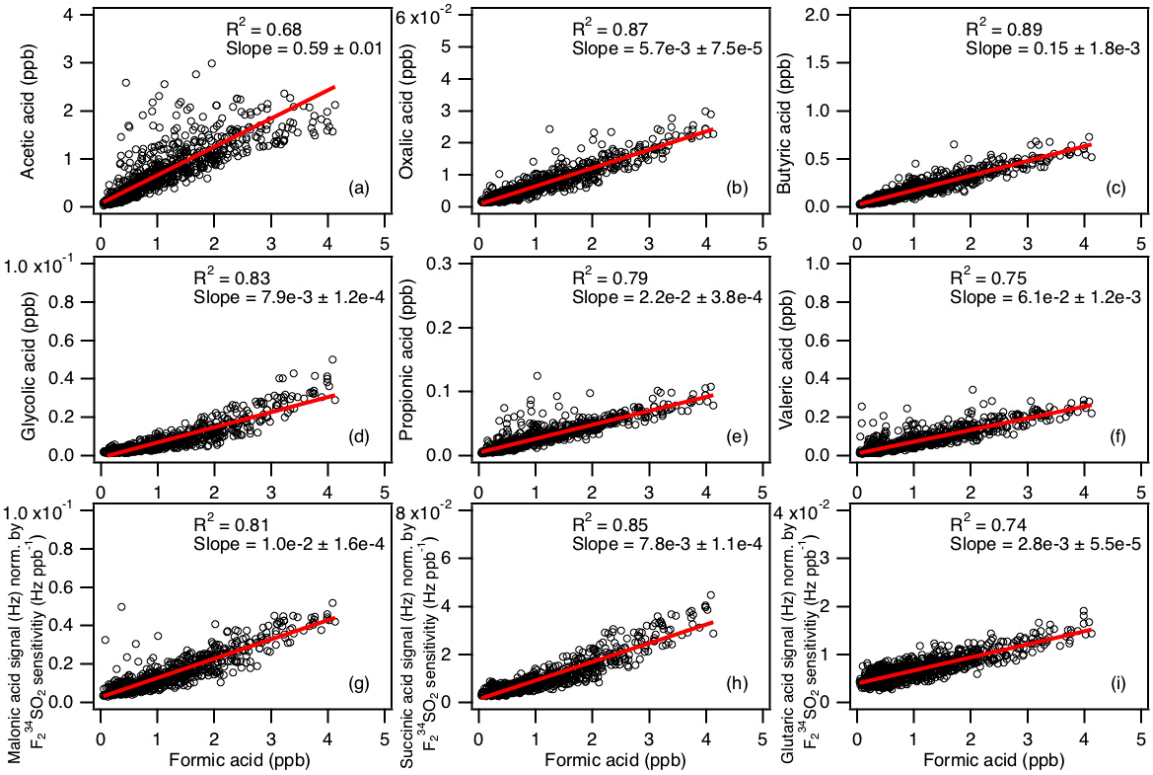

Figure 7: Scatter plots of concentrations (or signals) of (a) acetic, (b) oxalic, (c) butyric, (d) glycolic, (e) propionic, (f) valeric, (g) malonic, (h) succinic, and (i) glutaric acids with formic acid concentration. All the data are displayed as 1-hour averages. The data for malonic, succinic and glutaric acids are presented as the ratio of their ion signals (Hz) to the instrument's sensitivity to $F_2{}^{34}SO_2$ (Hz ppb^{-1}) since these organic acids were not calibrated. Red lines shown are linear fits to the data.

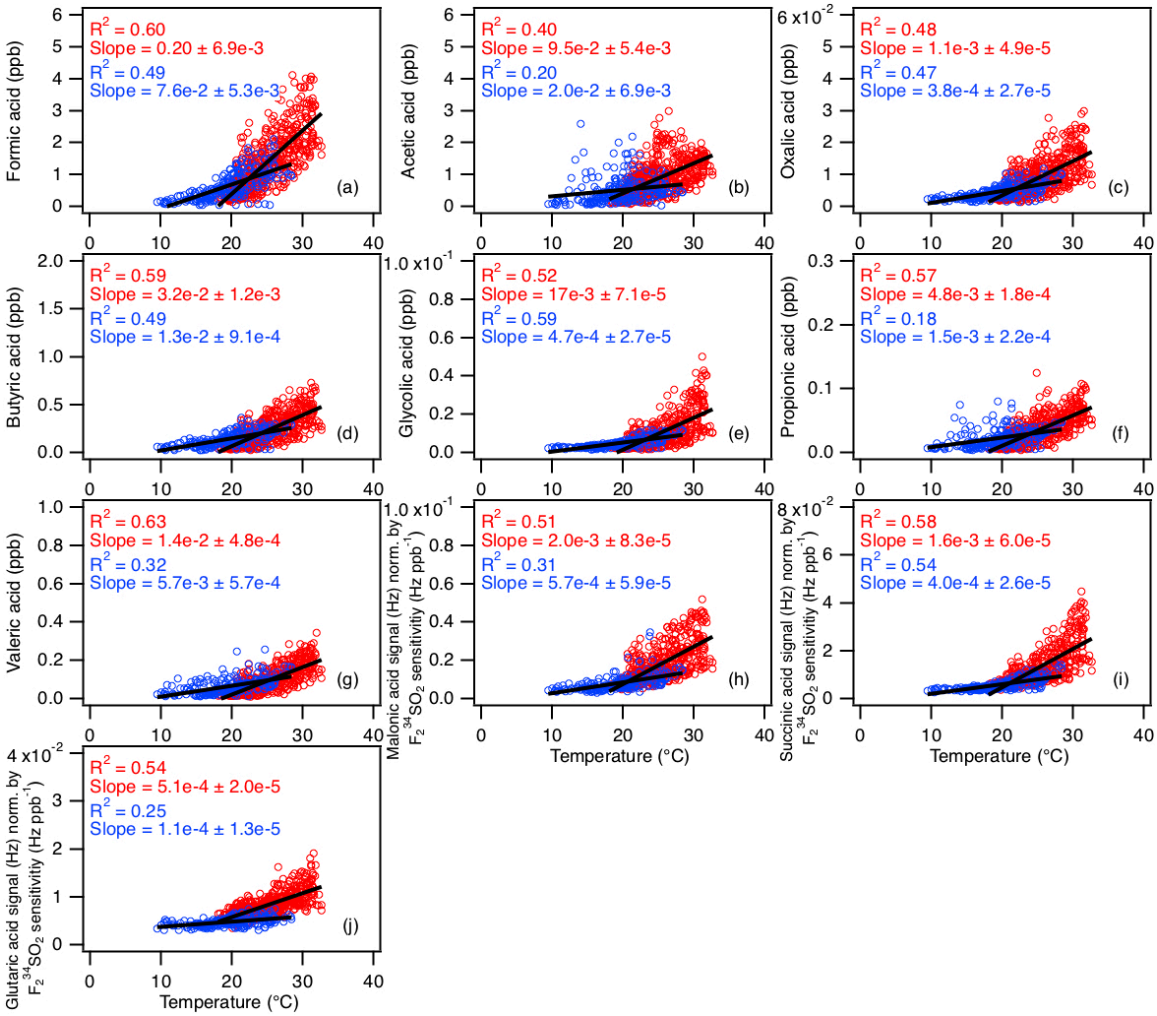

Figure 8: Scatter plots of concentrations (or signals) of (a) formic, (b) acetic, (c) oxalic, (d) butyric, (e) glycolic, (f) propionic, (g) valeric, (h) malonic, (i) succinic, and (j) glutaric acids with ambient temperature. The red symbols are data collected from 3 to 27 Sept, while the blue symbols are data collected from 28 Sept onwards. All the data are displayed as 1-hour averages. The data for malonic, succinic and glutaric acids are presented as the ratio of their ion signals (Hz) to the instrument's sensitivity to $F_2{}^{34}SO_2$ (Hz ppb^{-1}) since these organic acids were not calibrated. Black lines shown are linear fits to the datasets.

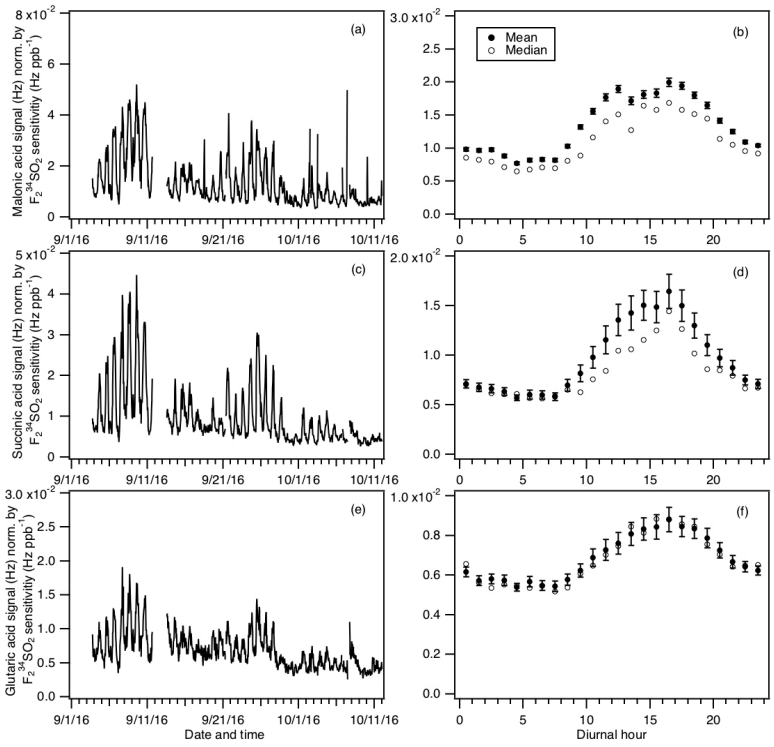

Figure S5: Time series of signals of (a) malonic, (c) succinic, and (e) glutaric acids measured during the field study. The data are displayed as 1-hour averages. Their corresponding diurnal profiles are shown in (b), (d) and (f), respectively. All the signals represent averages in 1-hour intervals and the standard errors are plotted as error bars. These organic acids were not calibrated so all the signals are presented here as Hz normalized by the instrument's sensitivity to $F_2{}^{34}SO_2$ (Hz ppb^{-1}) which was the primary calibrant used in the field study.

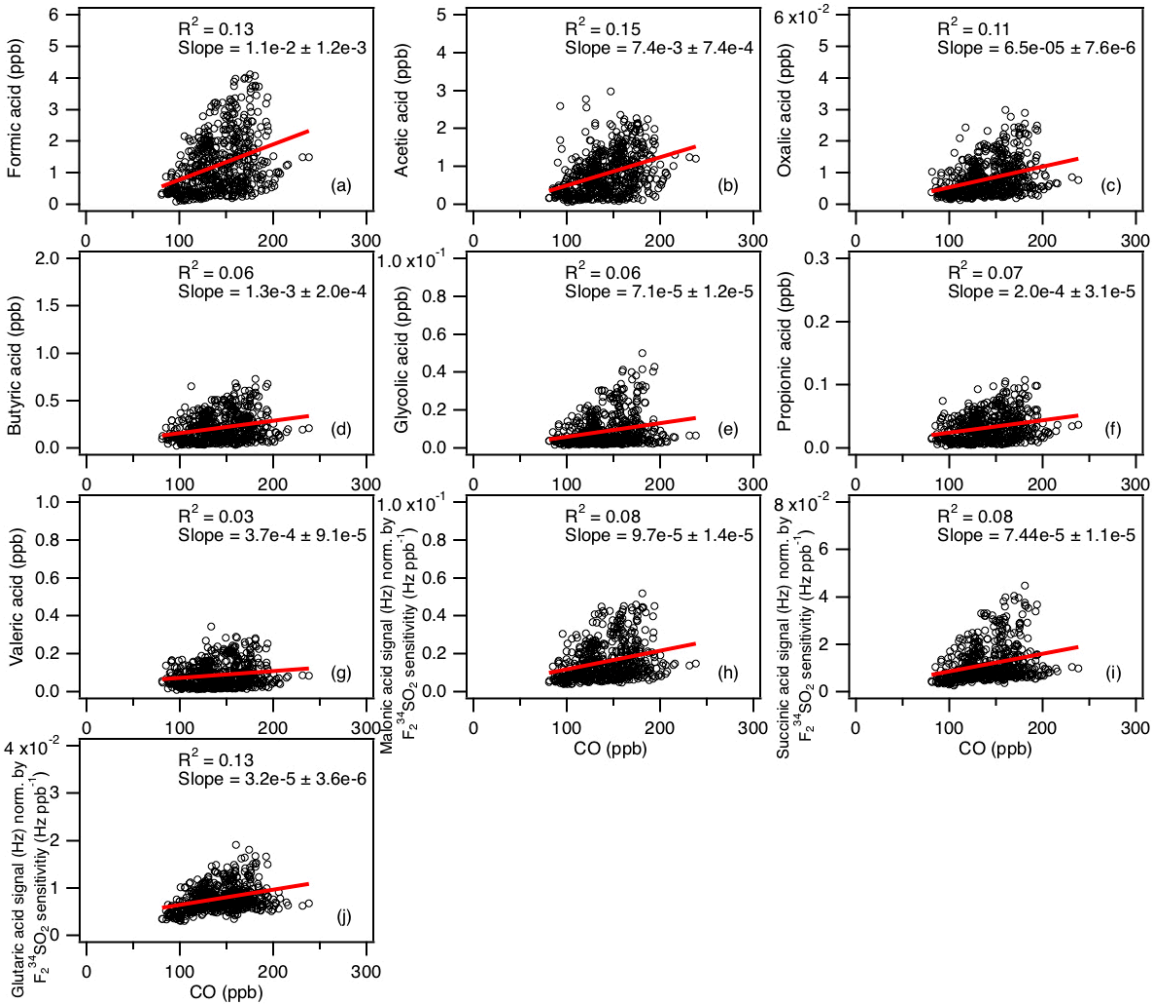

Figure S7: Scatter plots of concentrations (or signals) of (a) formic, (b) acetic, (c) oxalic, (d) butyric, (e) glycolic, (f) propionic, (g) valeric, (h) malonic, (i) succinic, and (j) glutaric acids with CO concentration. All the data are displayed as 1-hour averages. The data for malonic, succinic and glutaric acids are presented as the ratio of their ion signals (Hz) to the instrument's sensitivity to $F_2{}^{34}SO_2$ (Hz ppb^{-1}) since these organic acids were not calibrated. Red lines shown are linear fits to the data.

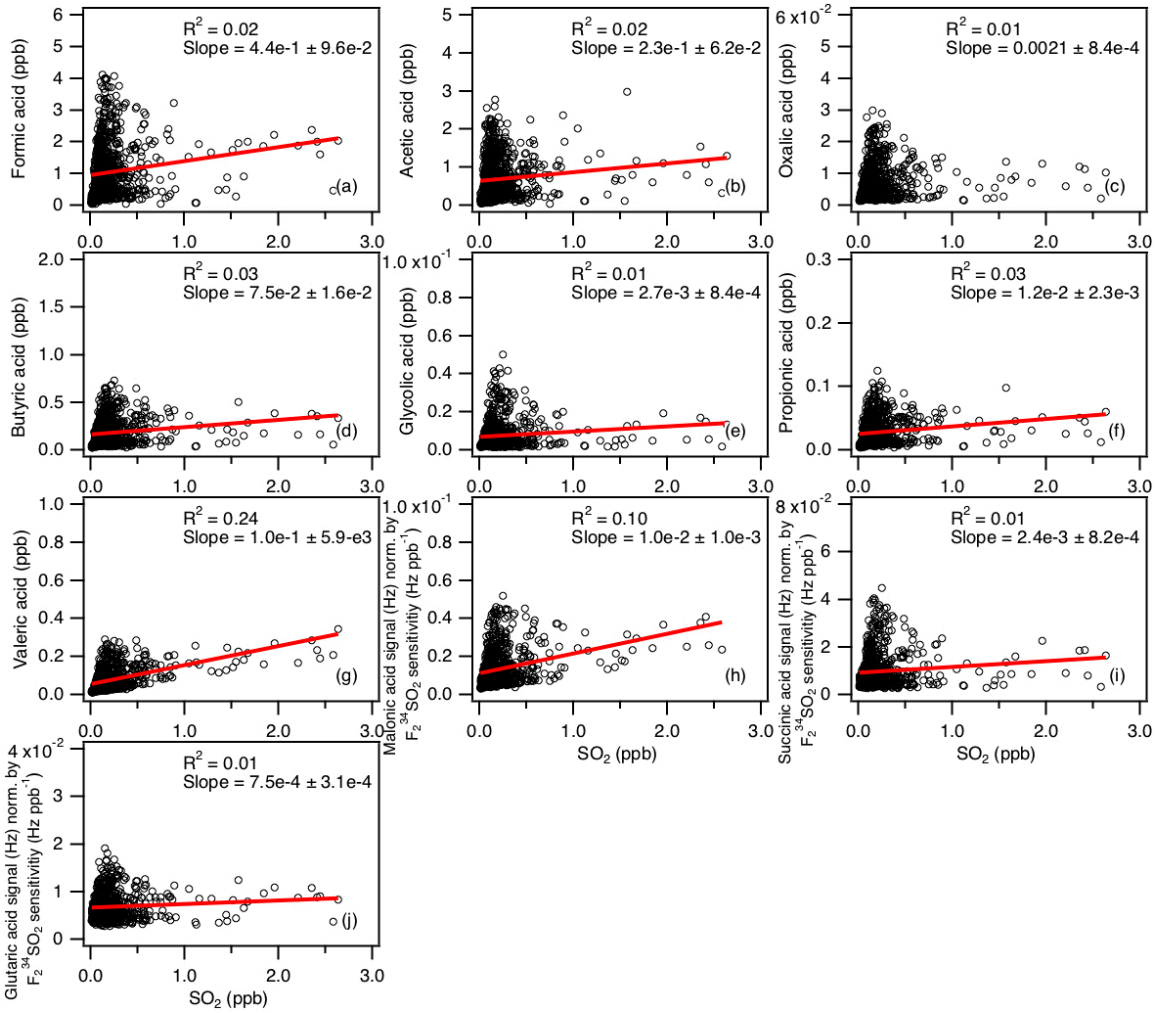

Figure S8: Scatter plots of concentrations (or signals) of (a) formic, (b) acetic, (c) oxalic, (d) butyric, (e) glycolic, (f) propionic, (g) valeric, (h) malonic, (i) succinic, and (j) glutaric acids with SO_2 concentration. All the data are displayed as 1-hour averages. The data for malonic, succinic and glutaric acids are presented as the ratio of their ion signals (Hz) to the instrument's sensitivity to $F_2{}^{34}SO_2$ (Hz ppb^{-1}) since these organic acids were not calibrated. Red lines shown are linear fits to the data.

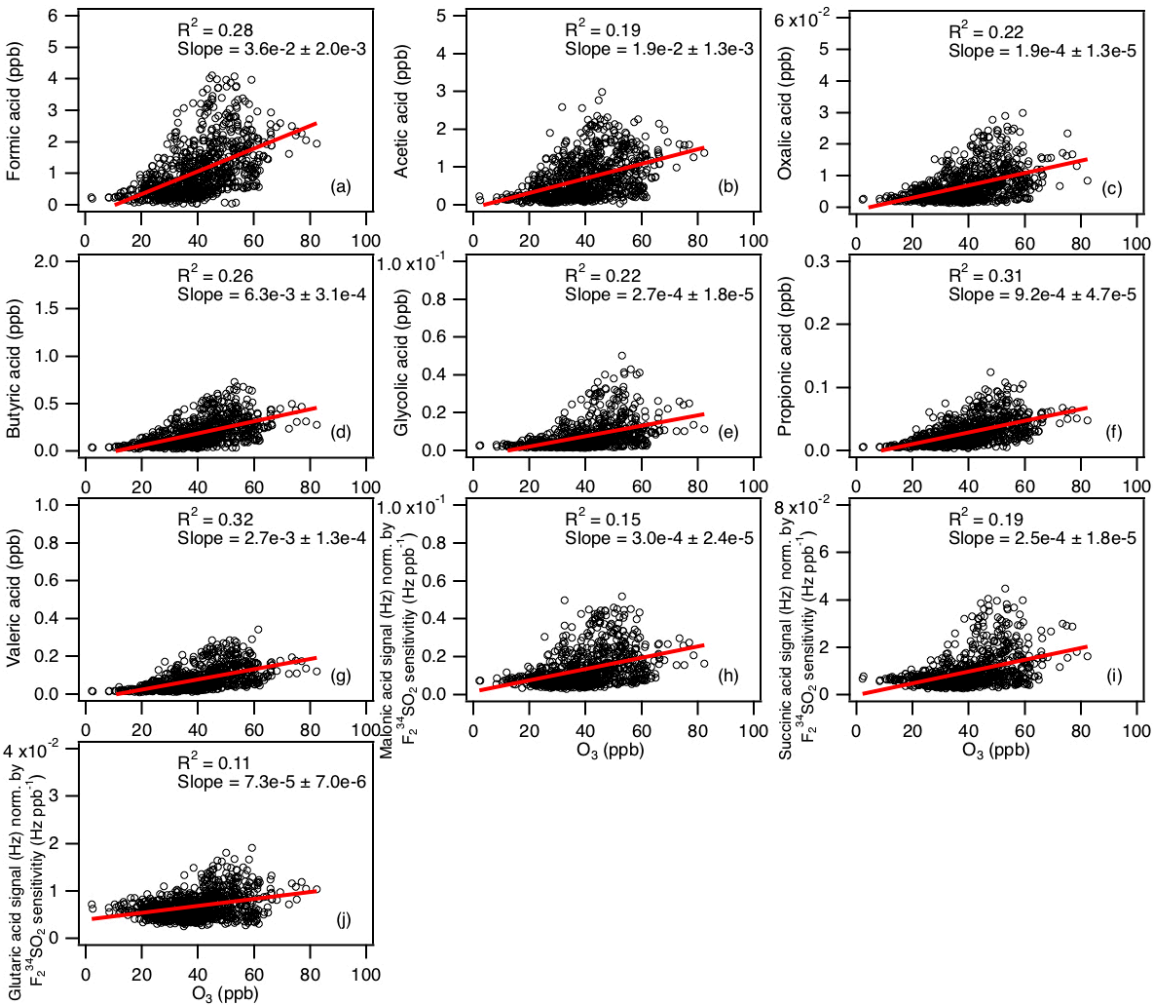

Figure S9: Scatter plots of concentrations (or signals) of (a) formic, (b) acetic, (c) oxalic, (d) butyric, (e) glycolic, (f) propionic, (g) valeric, (h) malonic, (i) succinic, and (j) glutaric acids with O_3 concentration. All the data are displayed as 1-hour averages. The data for malonic, succinic and glutaric acids are presented as the ratio of their ion signals (Hz) to the instrument's sensitivity to $F_2{}^{34}SO_2$ (Hz ppb^{-1}) since these organic acids were not calibrated. Red lines shown are linear fits to the data.

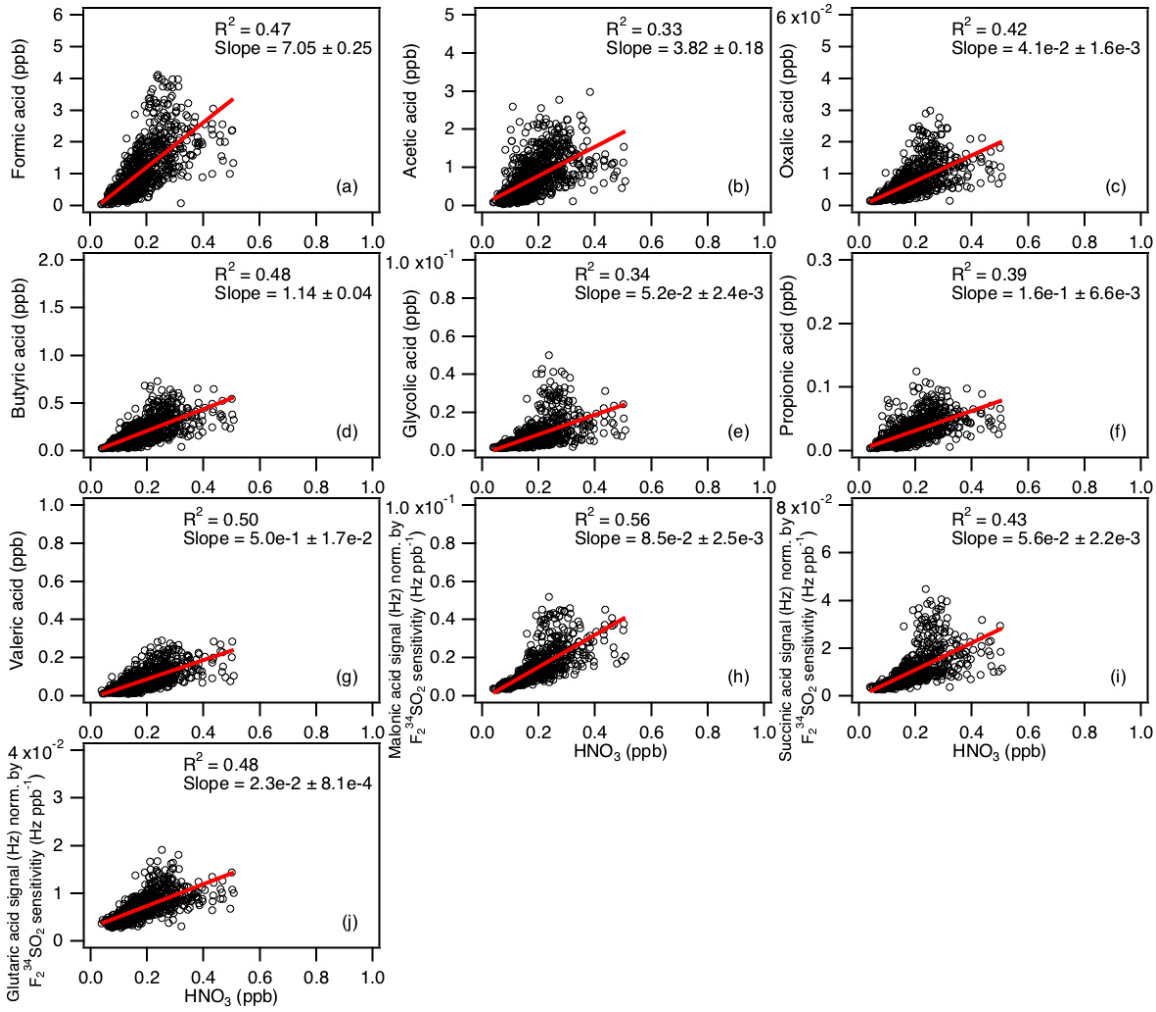

Figure S10: Scatter plots of concentrations (or signals) of (a) formic, (b) acetic, (c) oxalic, (d) butyric, (e) glycolic, (f) propionic, (g) valeric, (h) malonic, (i) succinic, and (j) glutaric acids with HNO_3 concentration. To exclude periods when the site was affected by urban or power plant emissions, data where HNO_3 > 0.5 ppb are excluded from these scatter plots. All the data are displayed as 1-hour averages. The data for malonic, succinic and glutaric acids are presented as the ratio of their ion signals (Hz) to the instrument's sensitivity to $F_2{}^{34}SO_2$ (Hz ppb^{-1}) since these organic acids were not calibrated. Red lines shown are linear fits to the data.

9. Referee comment: "*I don't understand the reagent ion signal and the use of sulfur isotopes. 34SF6- is said to be the reagent ion (line 338). Is isotopically labeled SF6 used? Why do the magnitudes of the peaks at mass 145 and 147 appear to be similar in Figure 2? Please clearly state the magnitude of the reagent ion signal. From figure 2, it appears that the reagent ion signal is approximately 400 kHz, and similar in magnitude to SF5-, CO3-. Wouldn't ion chemistry from those other ions also contribute to the reactions? A more thorough description of the reagent ion is necessary to understand how the ion chemistry can be dominated by SF6-. Figure 2 makes it appear that other ions could be substantial contributors to the ion chemistry.*"

Author response: Isotopically labeled SF_6^- was not used as the reagent ion. $^{32}SF_6^-$ (at m/z 146) is

the reagent ion. The signal of the reagent ion $^{32}SF_6^-$ was saturated for the entire field study, which led to the sharp drop in the ion signal at m/z 146 as shown in Fig. 2. Hence, we monitored the ion signal of its isotope $^{34}SF_6^-$ at m/z 148 (which was not saturated) to determine if the reaction of SF_6^- with ambient water vapor and O_3 depleted SF_6^- reagent ions. Despite fluctuations in ambient water vapor and O_3 concentrations, the $^{34}SF_6^-$ ion signal was relatively constant for the entire field study with a standard deviation of < 3%. This indicates that the reaction of SF_6^- with ambient water vapor and O_3 do not deplete $^{32}SF_6^-$ reagent ions.

The ion signal of $^{34}SF_6^-$ (at m/z 148) and the isotopic abundances of ^{32}S vs. ^{34}S can be used to estimate the theoretical ion signal of the $^{32}SF_6^-$ reagent ion at m/z 146 (i.e., ion signal obtained if no signal saturation occurred). We find that the $^{32}SF_6^-$ reagent ion would have a theoretical ion signal of ~9 x 10^6 Hz, which is 2 orders of magnitude larger than the CO_3^- signal. In addition, we expect the SF_6^- ion chemistry to dominate since SF_5^- is much more stable than SF_6^- and in any case will likely give similar product ions.

We have revised the manuscript and modified Fig. 2's caption to eliminate any confusion regarding the reagent ion used and contributions of other ions such as SF_5^- and CO_3^- to the ion chemistry:

Page 13 line 395: "Figure 2 shows a mass spectrum of ambient air. Interference peaks at m/z 39 (F^-•(HF) and CO_3^-, respectively) can be attributed to the presence of water and O_3, respectively. The reagent ion $^{32}SF_6^-$ is present at m/z 146. The $^{32}SF_6^-$ reagent ion signal was saturated, and this caused the sharp drop in the m/z 146 signal as shown in Fig. 2. Since the $^{32}SF_6^-$ reagent ion signal was saturated for the entire field study, we monitored the ion signal of its isotope $^{34}SF_6^-$ to determine if the reaction of SF_6^- with ambient water vapor (5.92 x 10^{-6} to 2.19 x 10^{-5} g cm^{-3}) and O_3 (2.1 to 82.4 ppb) depleted SF_6^- reagent ions. Figure S2a shows the time series of the $^{34}SF_6^-$ ion signal and ambient water vapor concentration for the entire field study. Despite fluctuations in ambient water vapor and O_3 concentrations, the $^{34}SF_6^-$ ion signal was relatively constant for the entire field study with a standard deviation of < 3%. This indicates that the reaction of SF_6^- with ambient water vapor and O_3 did not significantly deplete the $^{32}SF_6^-$ reagent ions during the field study."

Figure 2: Mass spectrum of ambient air and background measured in Yorkville, Georgia on 8 Sept 2016 using SF_6^-. Note that the $^{32}SF_6^-$ reagent ion signal (at m/z 146) is saturated, causing the sharp drop in its signal. As a result, the ion signal of its isotope $^{34}SF_6^-$ (at m/z 150) was monitored to determine if the reaction of SF_6^- with ambient water vapor and O_3 depleted SF_6^- reagent ions.

10. Referee comment: "*Since the major measurement technique advance here is using SF6 in high water and ozone environments, there should be greater discussion of water and ozone mixing ratios in the main text, and the necessary modifications to make the ion chemistry work in this environment. Although the supplement shows some ozone and water data, simpler discussion of the range of ozone and water mixing ratios and the effect on the ion chemistry should be included in the manuscript. Please show how formic or acetic sensitivity varies with ozone and water.*"

Author response: Ambient water vapor concentrations ranged from 5.92 x 10^{-6} to 2.19 x 10^{-5} g cm^{-3}, while ambient O_3 concentrations ranged from 2.1 to 82.4 ppb during the study. The sampled air was diluted in the CIMS flow tube and the flow tube was operated at a low pressure to minimize interferences by water vapor and O_3. Our results indicate that the reaction of SF_6^- with ambient

water vapor and O_3 did not deplete the $^{32}SF_6^-$ reagent ions. The SO_2 sensitivity showed a clear linear relationship to water vapor and varied within a factor of two for the entire field study, while the SO_2 sensitivity did not show any obvious dependence on ambient O_3 concentrations. The formic and acetic acid sensitivities did not show any obvious dependence on ambient water vapor and O_3 concentrations. Therefore, we do not expect the sensitivities of the other organic acids to depend on ambient water vapor and O_3 concentrations.

As requested, we have added more discussion about the ambient water vapor and O_3 concentrations and their effects on SF_6^- ion chemistry, the necessary sampling configurations needed to make SF_6^- ion chemistry to work in the field study, and how the formic and acetic acid sensitivity varies with water vapor and O_3:

Page 13 line 398: "Since the $^{32}SF_6^-$ reagent ion signal was saturated for the entire field study, we monitored the ion signal of its isotope $^{34}SF_6^-$ to determine if the reaction of SF_6^- with ambient water vapor (5.92 x 10^{-6} to 2.19 x 10^{-5} g cm^{-3}) and O_3 (2.1 to 82.4 ppb) depleted SF_6^- reagent ions. Figure S2a shows the time series of the $^{34}SF_6^-$ ion signal and ambient water vapor concentration for the entire field study. Despite fluctuations in ambient water vapor and O_3 concentrations, the $^{34}SF_6^-$ ion signal was relatively constant for the entire field study with a standard deviation of < 3%. This indicates that the reaction of SF_6^- with ambient water vapor and O_3 did not significantly deplete the $^{32}SF_6^-$ reagent ions during the field study.

The $F_2{}^{34}SO_2^-$ ion signal was used to monitor the CIMS SO_2 sensitivity during the field study. Figure S2b shows the time series of the $F_2{}^{34}SO_2^-/{}^{34}SF_6^-$ ion signal ratio obtained in calibration measurements. There is a noticeable increase in the $F_2{}^{34}SO_2^-/{}^{34}SF_6^-$ ion signal ratio on 28 Sept 2016, indicating an increase in the CIMS instrument sensitivity. The increase in CIMS instrument sensitivity is due to the decrease in ambient water vapor concentrations on 28 Sept 2016 (Fig. S2a). Previous laboratory and field studies showed that this was due to the hydrolysis of $F_2{}^{34}SO_2^-$, which led to the loss of this ion and diminished sensitivity at higher levels of ambient water vapor (Arnold and Viggiano, 2001; Slusher et al., 2001). However, the SO_2 sensitivity at $F_2{}^{34}SO_2^-$ only varied within a factor of two for the entire field study with a clear relationship to water vapor (Fig. S2c). The SO_2 sensitivity did not show any obvious dependence on ambient O_3 concentrations (Fig. S2d).

The formic ($HCOO^-$ at m/z 45 and $HCOO^-$•HF at m/z 65) and acetic (CH_3COO^-•HF at m/z 79) acid ions did not show any obvious dependence on ambient water vapor and O_3 concentrations during calibration measurements (Fig. S3). Therefore, we do not expect the sensitivities of the X^- and X^-•HF ions of the studied organic acids to depend on ambient water vapor and O_3 concentrations. We accounted for water vapor dependence of the $F_2{}^{34}SO_2^-$ ion signal in our post-field calibrations where the response of the CIMS acid signals were measured relative to the of the $^{34}SO_2$ sensitivity."

References:

Arnold, S. T., and Viggiano, A. A.: Turbulent ion flow tube study of the cluster-mediated reactions of SF6- with H2O, CH3OH, and C2H5OH from 50 to 500 torr, J. Phys. Chem. A, 105, 3527-3531, 10.1021/jp003967y, 2001.

Slusher, D. L., Pitteri, S. J., Haman, B. J., Tanner, D. J., and Huey, L. G.: A chemical ionization technique for measurement of pernitric acid in the upper troposphere and the polar boundary layer, Geophys. Res. Lett., 28, 3875-3878, 10.1029/2001gl013443, 2001.

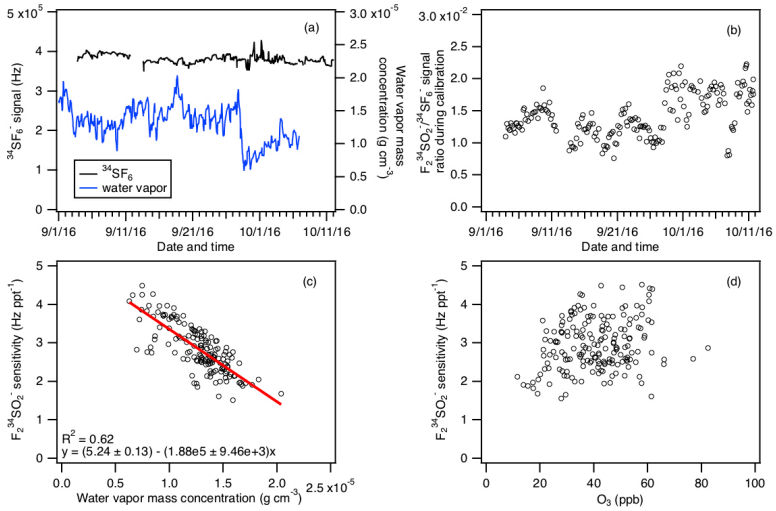

Figure S2: (a) Time series of $^{34}SF_6^-$ reagent ion signal and ambient water vapor concentration for the entire field study. The ambient water vapor mass concentrations are determined from ambient relative humidities and temperatures. (b) Time series of $F_2{}^{34}SO_2^-/{}^{34}SF_6^-$ ion signal ratio obtained during calibration measurements. Panels (c) and (d) show the $F_2{}^{34}SO_2^-$ ion sensitivity obtained from calibration measurements as a function of ambient water vapor and O_3 concentrations. Data in panels (a) to (d) are displayed as 1-hour averages.

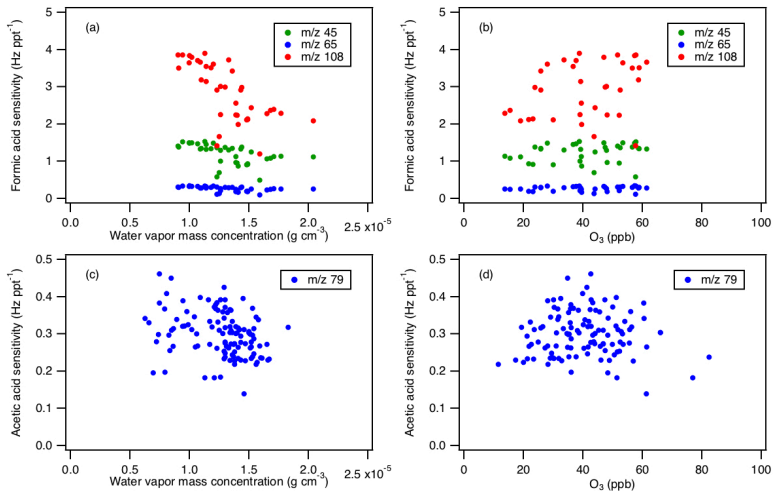

Figure S3: Panels (a) and (b) show the sensitivities of formic acid ions ($HCOO^-$ at m/z 45, $HCOO^-$•HF at m/z 65, and SF_4^- at m/z 108) obtained from calibration measurements as a function of ambient water vapor and O_3 concentrations. Panels (c) and (d) show the acetic acid sensitivity (CH_3COO^-•HF at m/z 79) obtained from calibration measurements as a function of ambient water vapor and O_3 concentrations.

11. Referee comment: "*One overlooked feature in the results is the differing diurnal variations of*

acetic and formic acids in figures 3-4. Rather than focus on organic acid budgets that rely on speculative assignments and suffer from unexamined interferences, a study of the diurnal variability of acetic and formic acids could prove interesting."

Author response: We do not know what caused the observed differing diurnal variations of formic and acetic acids. We are currently employing laboratory and modeling studies to explain these field observations. Results of these studies will be the subject of future papers.

Response to Referee 2 (Referees' comments are italicized)

1. Referee comment: "*The authors make a strong statement (line 95) that new techniques for the real-time measurement of gas-phase organic acids are need due to deficiencies in existing CIMS based ion chemistry (acetate and iodide CIMS). The authors cite issues with acetate CIMS in the detection of acetic acid and the wide range of sensitivities to different organic acids in iodide CIMS. After reading this paper it was not clear to me that SF6- has an advantage over these techniques. It was shown that interferences due to O3 hinder detection of acetic acid and there is an order of magnitude spread in sensitivity to various organic acids in Table 1. I think the authors need to better articulate how this technique is an advance over existing ion chemistry or acknowledge that it is a parallel approach to existing ion chemistry.*"

Author response: The SF_6^--CIMS technique serves as alternative approach to existing CIMS techniques in the detection of organic acids. The major advantage that SF_6^- has over I^- and $CH_3CO_2^-$ is that it allows for the detection of acetic acid and SO_2. CF_3O^- has a similar chemistry to SF_6^- but it also has issues due to hydrolysis and the ion precursor is not commercially available. This information has been added in the revised manuscript:

Page 4 line 107: "The sulfur hexafluoride (SF_6^-) anion has been used as a CIMS reagent ion to measure atmospheric inorganic species such as sulfur dioxide (SO_2), nitric acid (HNO_3) and peroxynitric acid (HO_2NO_2) (Slusher et al., 2001; Slusher et al., 2002; Huey et al., 2004; Kim et al., 2007). SF_6^- commonly reacts with most acidic gases at the collision rate by either proton or fluoride transfer reactions (Huey et al., 1995). The SF_6^- ion chemistry is selective to acidic species, which can simplify the mass spectral analysis of organic acids. However, SF_6^- is reactive to both ozone (O_3) and water vapor, which can lead to interfering reactions that limit its applicability to many species in certain environments (Huey et al., 2004). For these reasons, this work is focused on assessing the ability of SF_6^- to measure a series of organic acids in ambient air. The major advantage that SF_6^- has over I^- and $CH_3CO_2^-$ is that it allows for the detection of acetic acid and SO_2. CF_3O^- has a similar chemistry to SF_6^- but it also has issues due to hydrolysis and the ion precursor is not commercially available."

References:

Huey, L. G., Hanson, D. R., and Howard, C. J.: Reactions of SF6- and I- with Atmospheric Trace Gases, Journal of Physical Chemistry, 99, 5001-5008, 10.1021/j100014a021, 1995.

Huey, L. G., Tanner, D. J., Slusher, D. L., Dibb, J. E., Arimoto, R., Chen, G., Davis, D., Buhr, M. P., Nowak, J. B., Mauldin, R. L., Eisele, F. L., and Kosciuch, E.: CIMS measurements of

HNO3 and SO2 at the South Pole during ISCAT 2000, Atmospheric Environment, 38, 5411-5421, 10.1016/j.atmosenv.2004.04.037, 2004.

Kim, S., Huey, L. G., Stickel, R. E., Tanner, D. J., Crawford, J. H., Olson, J. R., Chen, G., Brune, W. H., Ren, X., Lesher, R., Wooldridge, P. J., Bertram, T. H., Perring, A., Cohen, R. C., Lefer, B. L., Shetter, R. E., Avery, M., Diskin, G., and Sokolik, I.: Measurement of HO2NO2 in the free troposphere during the intercontinental chemical transport experiment - North America 2004, Journal of Geophysical Research-Atmospheres, 112, 10.1029/2006jd007676, 2007.

Slusher, D. L., Pitteri, S. J., Haman, B. J., Tanner, D. J., and Huey, L. G.: A chemical ionization technique for measurement of pernitric acid in the upper troposphere and the polar boundary layer, Geophys. Res. Lett., 28, 3875-3878, 10.1029/2001gl013443, 2001.

Slusher, D. L., Huey, L. G., Tanner, D. J., Chen, G., Davis, D. D., Buhr, M., Nowak, J. B., Eisele, F. L., Kosciuch, E., Mauldin, R. L., Lefer, B. L., Shetter, R. E., and Dibb, J. E.: Measurements of pernitric acid at the South Pole during ISCAT 2000, Geophys. Res. Lett., 29, 10.1029/2002gl015703, 2002.

2. Referee comment: "*Given the focus of the journal, I think more emphasis on the experimental configuration of the CIMS should be given. Direct comparisons to existing measurements are always welcome, but the focus should remain on describing the details of the ion chemistry and or instrument operation.*"

Author response: We have added more details on the ion chemistry and instrument operation and more discussions on interferences in the revised manuscript. Please refer to our responses to comments on experimental details and interferences from referee 1 and 2.

3. Referee comment: "*Line 46: I would cite Molina et al. (2004), or Vlasenko et al. (2008) for the heterogeneous source of organic acids from the chemical aging of organic aerosol.*"

Author response: These references have been added.

4. Referee comment: "*Line 166: Please elaborate on how the lower pressure (13 mbar) minimizes interferences in reactions of SF6- with water vapor.*"

Author response: SF_6^- reacts with water vapor to form $F^-\bullet(HF)_n$ cluster ions. Arnold and Viggiano (2001) showed previously that the formation of these $F^-\bullet(HF)_n$ cluster ions are enhanced at high flow tube pressures due to increased reactive collisions between SF_6^- and water vapor. We maintained the flow tube at a low pressure to reduce reactive SF_6^-/water vapor collisions in the flow tube, thus minimizing interferences from SF_6^- reaction with water vapor.

We made the following changes in the revised manuscript:

Page 6 line 174: "Arnold and Viggiano (2001) showed that the formation of $F^-\bullet(HF)_n$ cluster ions from the reaction of SF_6^- and water vapor is enhanced at high flow tube pressures. Since these $F^-\bullet(HF)_n$ cluster ions could interfere with mass spectral analysis, the flow tube was maintained at a low pressure (~13 mbar, 0.5 % uncertainty) in this study to reduce both the water vapor concentration and reaction time in the flow tube, thus minimizing interferences

from SF_6^- reaction with water vapor."

5. Referee comment: "*Line 171: Rather than reporting voltages, it would be better to report electric fields or relative electric fields (E/N).*"

Author response: The electric field of the CDC is reported in the revised manuscript:

Page 6 line 184: "For this study, the CDC was operated at a relatively high electric field (~113 V cm^{-1}) to efficiently dissociate cluster ions."

6. Referee comment: "*Line 182: Why was the background measurement period so long (~4 minutes). It would be helpful to show one of these in time. I would have expected that the background measurement period could be significantly shorter and still capture the baseline, unless there are inlet equilibration issues.*"

Author response: We chose a 4 min background measurement time to give the scrubber (during background measurements) and flow tube ample time to equilibrate when the three-way PFA Teflon valve was switched between ambient and background modes, and to obtain good averaging statistics. This is stated explicitly in the revised manuscript:

Page 7 line 204: "Each background and calibration measurement period lasted ~4 and ~3.5 min, respectively, which not only gave the scrubber (during background measurements) and flow tube ample time to equilibrate when the three-way PFA Teflon valve was switched between ambient and background modes, but also allowed us to obtain good averaging statistics during background and calibration measurements."

We also added a section (3.1.3) and a figure (Fig. S4) discussing instrument time response during background and calibration measurements in the revised manuscript. Please refer to our response to comment 4 from referee 1.

7. Referee comment: "*Line 183: What was the 1.12 ppm SO2 standard diluted to? Presumably calibrations were not done at this mixing ratio.*"

Author response: 1.85 ppb of $^{34}SO_2$ was used in calibration measurements. This information has been added to the revised manuscript:

Page 7 line 209: "1.85 ppb of $^{34}SO_2$ was added to sampled air flow during calibration measurements."

8. Referee comment: "*Line 184: Again, it would be more helpful to present as the concentration of FA or AA that is delivered instead of the permeation tube emission rates.*"

Author response: As requested, this information has been added to the revised manuscript:

Page 7 line 216: "6.75 ppb of formic acid and 5.87 ppb of acetic acid was added to sampled air flow during calibration measurements."

9. Referee comment: "*Line 308: For reactions 1a-c, should one think of these as separate reaction channels governed by ion-molecule kinetics or does every reaction proceed through 1a and the electric field strength of the CDC sets the ratio of the observed products. This may lead to strong*

deviations in the observed products based on instrument configuration."

Author response: Every reaction of SF_6^- with organic acids (HX) proceeds through reactions 1a to 1c, where the ratio of the observed product ions can be controlled by the field strength of the CDC. This is stated explicitly in the revised manuscript:

Page 11 line 331: "CIMS measurements of atmospheric constituents use ion-molecule reactions to selectively ionize compounds of interest in the complex matrix of ambient air and produce characteristic ions. The reactions of SF_6^- with the organic acids (HX) proceed through reactions 1a to 1c, and gave similar products to those reported previously for SF_6^- reactions with inorganic acids (Huey et al., 1995): SF_5^-, X^- and X^-•HF where X^- is the conjugate base of the organic acid (reactions 1a-c).

$$SF_6^- + HX \rightarrow X^-\bullet HF + SF_5 \quad \textbf{(1a)}$$

$$SF_6^- + HX \rightarrow X^- + HF + SF_5 \quad \textbf{(1b)}$$

$$SF_6^- + HX \rightarrow SF_5^- + HF + X \quad \textbf{(1c)}$$

The effective branching ratios of the SF_5^-, X^- and X^-•HF product ions can be impacted by the field strength of the CDC."

References:

Huey, L. G., Hanson, D. R., and Howard, C. J.: Reactions of SF6- and I- with Atmospheric Trace Gases, Journal of Physical Chemistry, 99, 5001-5008, 10.1021/j100014a021, 1995.

10. Referee comment: "*Line 334: Can the mass (or molar) dilution constant be reported here instead of the inlet flow? This would help the reader understand how much ambient O3 and H2O are reduced by the sampling geometry. Also, perhaps it would be helpful to more explicitly state how a reduction in ion-molecule reaction time is helpful. This would not help in sensitivity (assuming all reactions are at the collision limit), but presumably would help minimize secondary ion chemistry, correct?*"

Author response: The ratio of the sampled air to the N_2/SF_6 mixture introduced into the CIMS flow tube is approximately 1:13. The referee is correct in pointing out that the reduction in ion-molecule reaction time helps minimizes interfering secondary ion chemistry. We reduced the ion-molecule reaction time by sampling only 0.3 L min^{-1} of air through the variable orifice into the flow tube and maintaining the flow tube at a low pressure (~13 mbar). This information is stated explicitly in the revised manuscript.

Page 13 line 384: "These reactions can deplete SF_6^- as well as form a variety of potentially interfering ions from secondary reactions (e.g., $F^-\bullet(HF)_n$ and CO_3^- ions) that depend on more abundant atmospheric species. For these reasons, efforts were made to minimize interferences by limiting reaction times and the flow sampled into the CIMS. This was accomplished by sampling only 0.3 L min^{-1} of air through the variable orifice into the flow tube and maintaining the flow tube at a low pressure (~13 mbar). The 0.3 L min^{-1} sampled air flow is diluted by 3.7 slpm of N_2/SF_6 flow in the flow tube. The ratio of the sampled air flow to the N_2/SF_6 flow introduced into the flow tube is approximately 1:13. While the high

N_2/SF_6 flow (3.7 slpm) passed through the radioactive source into the flow tube increased the SF_6^- reagent ion signal, the high dilution of the sampled air flow in the flow tube reduced the CIMS instrument sensitivity by decreasing the number density of the analytes."

11. Referee comment: "*Line 335: What is the uncertainty in the IMR and CDC pressure? Are these pressures also controlled?*"

Author response: The uncertainty in the flow tube pressure (the so-called IMR) is 0.5 %, while the uncertainty in the CDC pressure is 10 %. These pressures are controlled by a variable orifice. This information has been added to the revised manuscript:

Page 6 line 176: "Since these $F^-•(HF)_n$ cluster ions could interfere with mass spectral analysis, the flow tube was maintained at a low pressure (~13 mbar, 0.5 % uncertainty) in this study to minimize interferences from SF_6^- reaction with water vapor. The analyte ions exited the flow tube and were accelerated through the collisional dissociation chamber (CDC), which was maintained at ~0.8 mbar (10 % uncertainty). The molecular collisions in the CDC served to dissociate weakly bound cluster ions into their core ions to simplify mass spectral analysis. Flow tube and CDC pressures were controlled by the automatic variable orifice."

<u>The following are additional minor changes the authors have made to the manuscript:</u>

1. We corrected the model series of the CIMS turbo pumps:

Page 6 line 168: "These sections were evacuated by a scroll pump (Edward nXDS 20i), a drag pump (Adixen MDP 5011) and two turbo pumps (Varian Turbo-V301), respectively."

2. We corrected the uncertainty in nitric acid ambient concentrations:

Page 10 line 283: "For nitric acid which was calibrated in post-field laboratory work using a permeation tube and UV optical absorption, the uncertainty in its ambient concentrations was estimated to be 13 % based on uncertainties in UV absorption measurements (10 %) and one standard deviation of the acid's UV absorption signals (3 %), $^{34}SO_2$ sensitivity (3 %) and acid's calibrant ion signals (8 %)."

Real-time measurements of gas-phase organic acids using SF_6^- chemical ionization mass spectrometry

Theodora Nah,[1] Yi Ji,[1,2] David J. Tanner,[1] Hongyu Guo,[1] Amy P. Sullivan,[3] Nga Lee Ng,[1,2] Rodney J. Weber[1] and L. Gregory Huey[1*]

[1]*School of Earth and Atmospheric Sciences, Georgia Institute of Technology, Atlanta, GA, USA*
[2]*School of Chemical and Biomolecular Engineering, Georgia Institute of Technology, Atlanta, GA, USA*
[3]*Department of Atmospheric Science, Colorado State University, Fort Collins, CO, USA*

** To whom correspondence should be addressed: greg.huey@eas.gatech.edu*

Abstract

The sources and atmospheric chemistry of gas-phase organic acids are currently poorly understood due in part to the limited range of measurement techniques available. In this work, we evaluated the use of SF_6^- as a sensitive and selective chemical ionization reagent ion for real-time measurements of gas-phase organic acids. Field measurements are made using a chemical ionization mass spectrometer (CIMS) at a rural site in Yorkville, Georgia from September to October 2016 to investigate the capability of this measurement technique. Our measurements demonstrate that SF_6^- can be used to measure a range of organic acids in the atmosphere. 1-hour averaged ambient concentrations of organic acids ranged from a few parts per trillion by volume (ppt) to several parts per billion by volume (ppb). All the organic acids displayed similar strong diurnal behaviors, reaching maximum concentrations between 5 and 7 pm local time. The organic acid concentrations are dependent on ambient temperature, with higher organic acid concentrations being measured during warmer periods.

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Introduction

Organic acids are ubiquitous and important species in the troposphere. They are major contributors of free acidity in precipitation (Galloway et al., 1982; Keene et al., 1983; Keene and Galloway, 1984), and can also affect the formation of secondary organic aerosols (SOA) (Zhang et al., 2004; Carlton et al., 2006; Sorooshian et al., 2010; Yatavelli et al., 2015). As end products of oxidation, organic acids can also serve as useful tracers of air mass history (Sorooshian et al., 2007; Sorooshian et al., 2010). Organic acids are found

in urban, rural and remote marine environments in the gas, aqueous and particle phases. While organic acids are emitted directly from biogenic sources (e.g., microbial activity, vegetation and soil) and anthropogenic activities (e.g., fossil fuel combustion, vehicular emissions and biomass burning) (Kawamura et al., 1985; Talbot et al., 1988; Chebbi and Carlier, 1996; Talbot et al., 1999; Seco et al., 2007; Veres et al., 2010; Paulot et al., 2011; Veres et al., 2011; Millet et al., 2015), they can also be formed from photooxidation of non-methane volatile organic compounds and aqueous-phase photochemistry of semi-volatile organic compounds (Chebbi and Carlier, 1996; Hansen et al., 2003; Orzechowska and Paulson, 2005; Carlton et al., 2006; Sorooshian et al., 2007; Ervens et al., 2008; Paulot et al., 2011; Millet et al., 2015). The chemical aging of organic aerosols has also been proposed as a major source of organic acids (Molina et al., 2004; Vlasenko et al., 2008; Paulot et al., 2011). The relative importance of primary and secondary sources of organic acids are currently poorly constrained though their emissions likely depend on the magnitude of biogenic and anthropogenic activities and the meteorological conditions. Wet and dry deposition are the primary sinks of organic acids in the atmosphere (Chebbi and Carlier, 1996).

Formic and acetic acids are the dominant gas-phase monocarboxylic acids in the troposphere (Chebbi and Carlier, 1996). Due to their high vapor pressures, the gas-phase concentrations of formic and acetic acids are usually 1 to 2 orders of magnitudes higher than their particle-phase concentrations. Some field studies report strong correlations between formic and acetic acids, suggesting that these two organic acids have similar sources (Nolte et al., 1997; Souza and Carvalho, 2001; Paulot et al., 2011). A recent modeling study suggested that the dominant sources of formic acid in the southeastern U.S. are primarily biogenic in nature (Millet et al., 2015). These sources include direct emissions from vegetation and soil and photochemical production from biogenic volatile organic compounds (BVOCs). Currently, atmospheric formic and acetic acid concentrations are higher than those predicted by models, indicating that present model estimates of source and sink magnitudes are incorrect (Paulot et al., 2011; Millet et al., 2015). In the case of formic acid, deposition and secondary photochemical production via mechanisms such as photooxidation of isoprene and reaction of stabilized criegee intermediates need to be better constrained in models. Given that formic and acetic acids are major trace gases in

the atmosphere, there is a need to resolve the discrepancy between measurements and model predictions to close the atmospheric reactive carbon budget and improve our overall understanding of VOC chemistry in the atmosphere.

Currently, research on gas-phase organic acids has focused primarily on formic and acetic acids (Andreae et al., 1988; Talbot et al., 1988; Grosjean, 1991; Hartmann et al., 1991; Talbot et al., 1995; Talbot et al., 1999). This is due, in part, to the analytical difficulties in measuring gas-phase > C_2 organic acids and oxidized organic acids (i.e., containing more than 2 oxygen atoms) in real time. These organic acids have low vapor pressures and are generally present in low concentrations in the gas phase. For example, dicarboxylic acids typically have vapor pressures that are 2 to 4 orders of magnitude lower than their analogous monocarboxylic acids (Chebbi and Carlier, 1996), and are present mainly in the particle and aqueous phases. Rapid and accurate measurements of gas-phase > C_2 organic acids and oxidized organic acids are necessary for constraining the regional and global SOA budget since these acids can partition readily between the gas and particle and aqueous phases and subsequently affect SOA formation (Zhang et al., 2004; Carlton et al., 2006; Ervens et al., 2008; Sorooshian et al., 2010; Yatavelli et al., 2015).

Chemical ionization mass spectrometry (CIMS) is commonly used to selectively measure atmospheric trace gases in real-time with high sensitivity. CIMS measurements rely on reactions between reagent ions and compounds of interest present in the sampled air to produce analyte ions that are detected by a mass spectrometer. The subset of molecular species detected is determined by the reagent ion employed since the specificity of the ionization process is governed by the ion-molecule reaction mechanism. CIMS is a popular tool for atmospheric measurements since it is versatile and has high time resolution and sensitivity. It is also often a soft ionization technique with minimal ion fragmentation, thus preserving the parent molecule's elemental composition and allowing for molecular speciation. Recent developments in chemical ionization methods and sources have greatly improved our ability to measure atmospheric acidic species. Some of the CIMS reagent ions that have been used to measure atmospheric organic acids include acetate ($CH_3CO_2^-$), iodide (I^-) and CF_3O^- anions (Crounse et al., 2006; Veres et al., 2008; Lee et al., 2014; Brophy and Farmer, 2015; Nguyen et al., 2015). However, each of these CIMS reagent

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ions has its drawbacks, which are generally related to their selectivity and sensitivity towards different atmospheric species. For example, acetic acid is difficult to measure with $CH_3CO_2^-$ as the CIMS reagent ion due to interferences from the reagent ion chemistry that complicates the desired ion-molecule reactions. In addition, while many organic acids can be detected using I^- as a reagent ion, its sensitivity to different acids can vary by orders of magnitude (Lee et al., 2014).

The sulfur hexafluoride (SF_6^-) anion has been used as a CIMS reagent ion to measure atmospheric inorganic species such as sulfur dioxide (SO_2), nitric acid (HNO_3) and peroxynitric acid (HO_2NO_2) (Slusher et al., 2001; Slusher et al., 2002; Huey et al., 2004; Kim et al., 2007). SF_6^- commonly reacts with most acidic gases at the collision rate by either proton or fluoride transfer reactions (Huey et al., 1995). The SF_6^- ion chemistry is selective to acidic species, which can simplify the mass spectral analysis of organic acids. However, SF_6^- is reactive to both ozone (O_3) and water vapor, which can lead to interfering reactions that limit its applicability to many species in certain environments (Huey et al., 2004). For these reasons, this work is focused on assessing the ability of SF_6^- to measure a series of organic acids in ambient air. The major advantage that SF_6^- has over I^- and $CH_3CO_2^-$ is that it allows for the detection of acetic acid and SO_2. CF_3O^- has a similar chemistry to SF_6^- but it also has issues due to hydrolysis and the ion precursor is not commercially available. We present ambient measurements of gas-phase organic acids conducted in a mixed forest-agricultural area in Georgia in early fall of 2016 to evaluate the performance of a SF_6^- CIMS technique. Gas-phase organic acid measurements are compared to gas-phase water-soluble organic carbon ($WSOC_g$) measurements performed during the field study to estimate the fraction of $WSOC_g$ that is comprised of organic acids at this rural site. Laboratory experiments are conducted to measure the sensitivity of SF_6^- with a series of organic acids of atmospheric relevance.

2. Methods

2.1. Field site

Real-time ambient measurements of gas-phase organic acids were obtained using a chemical ionization mass spectrometer from 3 Sept to 12 Oct 2016 at the SouthEastern

Aerosol Research and Characterization (SEARCH) site located in Yorkville, Georgia. A detailed description of the field site has been provided by Hansen et al. (2003). Briefly, the Yorkville field site (33.931 N, 85.046 W) was located ~55 km northwest of Atlanta, and was on a broad ridge in a large pasture where there were occasionally grazing cattle. The field site was surrounded by forest and agricultural land. There were no major roads near the field site and nearby traffic emissions were negligible. The sampling period was characterized by moderate temperatures (24.0 °C average, 32.6 °C max, 9.5 °C min) and high relative humidities (68.9% RH average, 100% RH max, 21.6% RH min). The study-averaged diurnal trends of relative humidity, temperature and solar radiance are shown in Fig. S1. Data reported are displayed in EDT. Volumetric gas concentrations reported are at ambient temperature and relative humidity.

2.2. SF_6^- CIMS

2.2.1. CIMS instrument and air sampling inlet

The CIMS instrument was housed in a temperature controlled trailer during the field study. The inlet configuration and CIMS instrument used in this study is shown in Fig. 1. Since HNO_3 and organic acids may condense on surfaces, an inlet configuration with a minimal wall interaction was used. This inlet configuration was previously described by Huey et al. (2004) and Nowak et al. (2006); hence, only a brief description will be provided here. The inlet was a 7.6 cm ID aluminum pipe that extended ~40 cm into the ambient air through a hole in the trailer's wall. This positioned the inlet ~2 m above the ground. A donut-shaped ring was attached to the ambient sampling port of the pipe to reduce the influence of crosswinds on the pipe's flow dynamics. This ring was wrapped with a fine wire mesh to prevent insects from being drawn through the pipe. A flow of ~2800 L min^{-1} was maintained in the pipe using a regenerative blower (AMETEK Windjammer 116637-03). Part of this flow (7 L min^{-1}) was sampled through a custom-made three-way PFA Teflon valve, which connected the pipe's center to the CIMS sampling orifice. The valve was maintained at a temperature of 40 °C in an insulated aluminum oven and could be switched automatically between ambient and background modes. In ambient mode, ambient air was passed through a 25 cm long, 0.65 cm ID Teflon tube into the CIMS. In background mode, ambient air was first drawn through an activated

charcoal scrubber before being delivered into the CIMS. A small flow of ambient air (~0.05 L min^{-1}) was continuously passed through the scrubber to keep it at equilibrium with ambient humidity levels. Most of the sampled air flow (6.7 L min^{-1}) was exhausted using a small diaphragm pump. The rest of the sampled air flow (0.3 L min^{-1}) was introduced into the CIMS instrument through an automatic variable orifice, which was used to maintain a constant sample air mass flow.

The CIMS instrument was comprised of a series of differentially pumped regions: a flow tube, a collisional dissociation chamber, an octopole ion guide, a quadrupole mass filter and an ion detector. These sections were evacuated by a scroll pump (Edward nXDS 20i), a drag pump (Adixen MDP 5011) and two turbo pumps (Varian Turbo-V301), respectively. Ambient air was drawn continuously into the flow tube. A flow of 3.7 slpm of N_2 containing a few ppm of SF_6 (Scott-Marrin Inc.) was passed through a ^{210}Po ion source into the flow tube. SF_6^- anions, which were produced via associative electron attachment in the ^{210}Po ion source, reacted with the sampled ambient air in the flow tube to generate analyte ions. Arnold and Viggiano (2001) showed that the formation of $F^-•(HF)_n$ cluster ions from the reaction of SF_6^- and water vapor is enhanced at high flow tube pressures. Since these $F^-•(HF)_n$ cluster ions could interfere with mass spectral analysis, the flow tube was maintained at a low pressure (~13 mbar, 0.5 % uncertainty) in this study to reduce both the water vapor concentration and reaction time in the flow tube, thus minimizing interferences from SF_6^- reaction with water vapor. The analyte ions exited the flow tube and were accelerated through the collisional dissociation chamber (CDC), which was maintained at ~0.8 mbar (10 % uncertainty). The molecular collisions in the CDC served to dissociate weakly bound cluster ions into their core ions to simplify mass spectral analysis. Flow tube and CDC pressures were controlled by the automatic variable orifice. For this study, the CDC was operated at a relatively high electric field (~113 V cm^{-1}) to efficiently dissociate cluster ions. The resulting ions were then passed into the octopole ion guide (maintained at ~6 x 10^{-3} mbar), which collimated the ions and transferred them into the quadrupole mass spectrometer (maintained at ~10^{-5} mbar) for mass selection and detection.

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Ions monitored during the field study included m/z 45, 59, 65, 73, 75, 79, 82, 87, 89, 101, 102, 103, 108, 117, 131 and 148. The assignment of these ions will be discussed in section 3. The dwell time for each m/z ion was set to 0.5 s and measurements of these ions were obtained every ~13 s, which resulted in a ~4 % (= 0.5/13 x 100 %) duty cycle for each ion monitored. The data presented in this paper was averaged to 1-hour intervals unless stated otherwise.

2.2.2. Background and calibration measurements during field study

Background measurements were performed every 25 min during the field study. During each background measurement, the sampled air flow was passed through an activated charcoal scrubber prior to delivery into the CIMS. The scrubber removed > 99 % of the targeted species in ambient air. Calibration measurements were performed every 5 h during the field study through standard additions of $^{34}SO_2$ and either formic or acetic acid to the sampled air flow. Each background and calibration measurement period lasted ~4 and ~3.5 min, respectively, which not only gave the scrubber (during background measurements) and flow tube ample time to equilibrate when the three-way PFA Teflon valve was switched between ambient and background modes, but also allowed us to obtain good averaging statistics during background and calibration measurements. A 1.12 ppm $^{34}SO_2$ gas standard was used as the source of the sulfur standard addition. 1.85 ppb of $^{34}SO_2$ was added to sampled air flow during calibration measurements. The formic and acetic acid calibration sources were permeation tubes (VICI Metronics) with emission rates of 91 and 110 ng min^{-1}, respectively. The emission rates were measured by scrubbing the output of the permeation tube in deionized water via a gas impinge immersed in water, which was then analyzed for formate and acetate using ion chromatography (Thermo Fisher Scientific). Eight samples of each acid were analyzed over the course of the field study and the standard deviations of the permeation rates were ≤ 6 %. 6.75 ppb of formic acid and 5.87 ppb of acetic acid was added to sampled air flow during calibration measurements. The CIMS instrument sensitivity measured by the $F_2{}^{34}SO_2^-$ ion signal (m/z 104) was similarly applied to all the other measured species (except for formic and acetic acids) using relative sensitivities determined in laboratory studies. The $F_2{}^{34}SO_2^-$ calibrant ion

signals were also used to calibrate ambient $F_2{}^{32}SO_2^-$ ion signals and determine ambient SO_2 concentrations as discussed in section 3.2.5.

2.2.3. Laboratory calibration

HNO_3, oxalic, butyric, glycolic, propionic and valeric acid standard addition calibrations were performed in post-field laboratory work. The response of the CIMS acid signals were measured relative to the sensitivity of $^{34}SO_2$ in these calibration measurements. The HNO_3 calibration source was a permeation tube (KIN-TEK) with a permeation rate of 39 ng min^{-1}, which was measured using UV optical absorption (Neuman et al., 2003). Solid or liquid samples of oxalic (Sigma Aldrich, ≥ 99 %), butyric (Sigma Aldrich, ≥ 99 %), glycolic (Sigma Aldrich, 99 %), propionic (Sigma Aldrich, ≥ 99.5 %) and valeric (Sigma Aldrich, ≥ 99 %) acids were used in calibration measurements. The acid sample was placed in a glass impinger, which was immersed in an ice bath to provide a constant vapor pressure. A flow of 6 to 10 mL min^{-1} of N_2 was passed over the organic acid in the glass impinger. This organic acid air stream was then diluted with varying flows of N_2 (1 to 5 L min^{-1}) to achieve different mixing ratios of the organic acid. Mixing ratios were calculated from either the acid's emission rate from the impinger or the acid's vapor pressure. The emission rate of gas-phase oxalic acid from the impinger was measured by scrubbing the output in deionized water using the same method for calibrating the formic and acetic acid permeation tubes, followed by ion chromatography analysis for oxalate. Three samples were analyzed and the emission rate was determined to be 14 ng min^{-1} with a standard deviation of < 5 %. The vapor pressures of butyric and propionic acids at 0 ˚C were measured using a capacitance manometer (MKS Instruments). The vapor pressures of glycolic and valeric acids at 0 ˚C were estimated using their literature vapor pressures at 25 ˚C and enthalpies of vaporization (Daubert and Danner, 1989; Lide, 1995; Acree and Chickos, 2010).

Attempts to generate a calibration plot for pyruvic acid using its liquid sample (Sigma Aldrich, 98 %) and the setup described above were unsuccessful as this acid was found to interact very strongly with surfaces. Glyoxylic acid calibrations were not performed due to the presence of impurities in the glyoxylic acid monohydrate solution used (Sigma Aldrich, 98 %), which resulted in the appearance of ions not attributed to

glyoxylic acid. We attempted to generate calibration plots for malonic (Sigma Aldrich, ≥ 99.5 %), succinic (Sigma Aldrich, 99 %) and glutaric (Sigma Aldrich, 99 %) acids by passing N_2 over their solid samples at room temperature. However, it was not possible to generate large enough gas phase concentrations for calibration since these organic acids have very low vapor pressures.

2.2.4. Detection limits and measurement uncertainties

The detection limits of the organic acids were estimated as 3 times the standard deviation values (3σ) of the ion signals measured during background mode. Although each background measurement period lasted ~4 min, ion signals of the different organic acids took up to 1.5 min to stabilize during the switch between ambient, calibration and background measurements during the field study. Thus, ion signals measured during the first 1.5 min were not included in the calculation of the average and standard deviation of ion signals measured during background mode. Table 1 summarizes the average detection limits of the organic acids for 2.5 min averaging periods which corresponds to the length of a background measurement with a 4 % duty cycle for each m/z. The mean difference between successive background measurements ranged from 1 to 40 ppt for the different organic acids. Future work will focus on reducing the instrument background, and therefore improving the detection limits of these organic acids.

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The uncertainties (1σ) in our ambient measurements of formic, acetic and oxalic acid concentrations originated from CIMS and IC calibration measurements. The IC measurement uncertainty was estimated to be 10 %. For formic and acetic acids, which were calibrated during the field study using permeation tubes, their CIMS measurement uncertainties were estimated to be 6 and 7 %, respectively, based on one standard deviation of the acids' calibrant ion signals. For oxalic acid which was calibrated in post-field laboratory work, the CIMS measurement uncertainty was estimated to be 9 % based on one standard deviation of the $^{34}SO_2$ sensitivity (3 %), the acid's calibrant ion signals (7 %) and linear fit of the calibration curve (5 %). Hence, the uncertainties in our ambient measurements of formic, acetic and oxalic acid concentrations were estimated to be 12, 12 and 14 %, respectively.

For nitric acid which was calibrated in post-field laboratory work using a permeation tube and UV optical absorption, the uncertainty in its ambient concentrations was estimated to be 13 % based on uncertainties in UV absorption measurements (10 %) and one standard deviation of the acid's UV absorption signals (3 %), $^{34}SO_2$ sensitivity (3 %) and acid's calibrant ion signals (8 %). For butyric and propionic acids which were calibrated in post-field laboratory work using vapor pressures measured by a capacitance manometer, the uncertainties in their ambient concentrations were estimated to be 14 % based on the vapor pressure measurement uncertainty (10 %) and one standard deviation of the $^{34}SO_2$ sensitivity (3 %), the acids' calibrant ion signals (8 %) and linear fits of the acids' calibration curves (3 %). For glycolic and valeric acids which were calibrated in post-field laboratory work using vapor pressures estimated from literature vapor pressures at 25 ˚C and enthalpies of vaporization, the uncertainties in their ambient concentrations were likely significantly larger compared to the other measured organic acids due to uncertainties in their estimated vapor pressures. We estimate the uncertainties in ambient concentrations of glycolic and valeric acids to be 22 % based on an assumed vapor pressure uncertainty of 20 % and one standard deviation of the $^{34}SO_2$ sensitivity (3 %), the acids' calibrant ion signals (8 %) and linear fits of the acids' calibration curves (2 %).

2.3. $WSOC_g$ measurements

$WSOC_g$ was measured with a MIST chamber coupled to a total organic carbon (TOC) analyzer (Sievers 900 series, GE Analytical Instruments). Ambient air first passed through a Teflon filter (45 mm diameter, 2.0 μm pore size, Pall Life Sciences) to remove particles in the air stream. This filter was changed every 3 to 4 days. The particle-free air was then pulled into a glass Mist Chamber filled with ultrapure deionized water at a flow rate of 20 L min^{-1}. The MIST chamber scrubbed soluble gases with Henry's law constants greater than 10^3 M atm^{-1} into deionized water (Spaulding et al., 2002). The resulting liquid samples from the MIST chamber were analyzed by the TOC analyzer. The TOC analyzer converted the organic carbon in the liquid samples to carbon dioxide using UV light and chemical oxidation. The carbon dioxide formed was then measured by conductivity. The amount of organic carbon in the liquid samples is proportional to the measured increase in conductivity of the dissolved carbon dioxide. Each $WSOC_g$ measurement lasted 4 min.

Background $WSOC_g$ measurements were performed for 45 min every 12 h by stopping the sample air flow and rinsing the sampling lines with deionized water. The TOC analyzer was calibrated using different concentrations of sucrose (as specified by the instrument manual) before and after the field study. The limit of detection was 0.4 μgC m^{-3}. The measurement uncertainty was estimated to be 10 % based on uncertainties in the sample air flow, liquid flow and TOC analyzer uncertainty. The MIST chamber and upstream particle filter were located in an air-conditioned building so were generally below ambient temperature. Hence, evaporation of collected particles (which will lead to positive artifacts in $WSOC_g$ measurements) are not expected to be significant.

2.4. Supporting gas measurements

Supporting gas measurements were provided by a suite of instruments operated by the SEARCH network. A non-dispersive infrared spectrometer (Thermo Fisher Scientific) provided hourly CO measurements. A UV absorption analyzer (Thermo Fisher Scientific) provided hourly O_3 measurements. A gas chromatography-flame ionization detector (GC-FID, Agilent Technologies) provided hourly VOC measurements.

3. Results and discussion

3.1. General SF_6^- CIMS field performance

3.1.1. SF_6^- ion chemistry with organic acids

CIMS measurements of atmospheric constituents use ion-molecule reactions to selectively ionize compounds of interest in the complex matrix of ambient air and produce characteristic ions. The reactions of SF_6^- with the organic acids (HX) proceed through reactions 1a to 1c, and gave similar products to those reported previously for SF_6^- reactions with inorganic acids (Huey et al., 1995): SF_5^-, X^- and X^-•HF where X^- is the conjugate base of the organic acid (reactions 1a-c).

$$SF_6^- + HX \rightarrow X^-\bullet HF + SF_5 \quad (1a)$$

$$SF_6^- + HX \rightarrow X^- + HF + SF_5 \quad (1b)$$

$$SF_6^- + HX \rightarrow SF_5^- + HF + X \quad (1c)$$

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The effective branching ratios of the SF_5^-, X^- and X^-•HF product ions can be impacted by the field strength of the CDC. The SF_5^- ion (m/z 127, reaction 1c) is a common reaction product of the reactions of SF_6^- with many species and is probably thermodynamically driven by the formation of HF (Huey et al., 1995). Unfortunately, the production of SF_5^- does not allow for the selective detection of any atmospheric species. In addition, the larger the branching ratio of the SF_5^- channel, the lower the CIMS sensitivity to an individual acid since the effective rate constants for the X^- and X^-•HF channels are lower.

The reaction of SF_6^- with formic acid and oxalic acid also produced SF_4^- ions (m/z 108). These reactions are probably thermodynamically driven by the formation of CO_2 and HF:

$$SF_6^- + HC(O)OH \rightarrow SF_4^- + CO_2 + 2HF \quad (2)$$

$$SF_6^- + HO(O)CC(O)OH \rightarrow SF_4^- + 2CO_2 + 2HF \quad (3)$$

We used the X^- and/or X^-•HF ions to determine ambient organic acid concentrations since these ions are characteristic of the individual acids. For all the organic acids, the X^-•HF ion signal is substantially lower than that of the X^- ion for the conditions in this study. However, this is probably largely due to the relatively high collision energy used in the CDC which led to efficient dissociation of the fluoride adducts to form X^- ions. Consequently, only the proton transfer channel (1b) is used to quantify most of the organic acids in the field study. The exceptions are formic and acetic acid as discussed in section 3.2.1 and 3.2.2

Table 1 shows a summary of the sensitivities of X^- and X^-•HF ions of organic acids. The average sensitivities of the $HCOO^-$ (m/z 45) and $HCOO^-$•HF (m/z 65) ions of formic acid were 1.29 ± 0.22 and 0.29 ± 0.05 Hz ppt^{-1}, respectively, while the average sensitivities of the CH_3COO^- (m/z 59) and CH_3COO^-•HF (m/z 79) ions of acetic acid were 1.46 ± 0.29 and 0.30 ± 0.06 Hz ppt^{-1}, respectively. A weak ^{210}Po ion source (< 1 mCi) was used by SF_6^--CIMS instrument during the field study, hence these sensitivities will be substantially higher if a stronger radioactive source is used. Nevertheless, these sensitivities are compared to formic and acetic acid sensitivities measured by a high-resolution time-of-flight chemical ionization mass spectrometer (Aerodyne Research Inc.) that utilized I^-

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reagent ions during the field study. Although the formic acid sensitivity measured by I^--CIMS (1.33 ± 0.28 Hz ppt^{-1}) was comparable to that measured by SF_6^--CIMS, the acetic acid sensitivity measured by I^--CIMS (< 0.1 Hz ppt^{-1}) was substantially lower than that measured by SF_6^--CIMS. Previous studies have similarly reported low acetic acid sensitivity measured by I^--CIMS (Aljawhary et al., 2013; Lee et al., 2014).

3.1.2. Characterization of interferences

SF_6^- is very sensitive to many trace atmospheric species but its reactions with water vapor and O_3 when sampling ambient air can lead to issues both with selectivity and stability. For example, SF_6^- reacts nonlinearly with water vapor to form a series of $F^-•(HF)_n$ cluster ions (Huey et al., 1995; Arnold and Viggiano, 2001). SF_6^- also reacts efficiently with O_3 to form O_3^-, which is rapidly converted to CO_3^- in ambient air (Slusher et al., 2001). These reactions can deplete SF_6^- as well as form a variety of potentially interfering ions from secondary reactions (e.g., $F^-•(HF)_n$ and CO_3^- ions) that depend on more abundant atmospheric species. For these reasons, efforts were made to minimize interferences by limiting reaction times and the flow sampled into the CIMS. This was accomplished by sampling only 0.3 L min^{-1} of air through the variable orifice into the flow tube and maintaining the flow tube at a low pressure (~13 mbar). The 0.3 L min^{-1} sampled air flow is diluted by 3.7 slpm of N_2/SF_6 flow in the flow tube. The ratio of the sampled air flow to the N_2/SF_6 flow introduced into the flow tube is approximately 1:13. While the high N_2/SF_6 flow (3.7 slpm) passed through the radioactive source into the flow tube increased the SF_6^- reagent ion signal, the high dilution of the sampled air flow in the flow tube reduced the CIMS instrument sensitivity by decreasing the number density of the analytes.

Figure 2 shows a mass spectrum of ambient air. Interference peaks at m/z 39 (F^-•(HF) and CO_3^-, respectively) can be attributed to the presence of water and O_3, respectively. The reagent ion $^{32}SF_6^-$ is present at m/z 146. The $^{32}SF_6^-$ reagent ion signal was saturated, and this caused the sharp drop in the m/z 146 signal as shown in Fig. 2. Since the $^{32}SF_6^-$ reagent ion signal was saturated for the entire field study, we monitored the ion signal of its isotope $^{34}SF_6^-$ to determine if the reaction of SF_6^- with ambient water vapor (5.92 x 10^{-6} to 2.19 x 10^{-5} g cm^{-3}) and O_3 (2.1 to 82.4 ppb) depleted SF_6^- reagent ions. Figure S2a shows the time series of the $^{34}SF_6^-$ ion signal and ambient water vapor

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concentration for the entire field study. Despite fluctuations in ambient water vapor and O_3 concentrations, the $^{34}SF_6^-$ ion signal was relatively constant for the entire field study with a standard deviation of < 3%. This indicates that the reaction of SF_6^- with ambient water vapor and O_3 did not significantly deplete the $^{32}SF_6^-$ reagent ions during the field study.

The $F_2{}^{34}SO_2^-$ ion signal was used to monitor the CIMS SO_2 sensitivity during the field study. Figure S2b shows the time series of the $F_2{}^{34}SO_2^-/{}^{34}SF_6^-$ ion signal ratio obtained in calibration measurements. There is a noticeable increase in the $F_2{}^{34}SO_2^-/{}^{34}SF_6^-$ ion signal ratio on 28 Sept 2016, indicating an increase in the CIMS instrument sensitivity. The increase in CIMS instrument sensitivity is due to the decrease in ambient water vapor concentrations on 28 Sept 2016 (Fig. S2a). Previous laboratory and field studies showed that this was due to the hydrolysis of $F_2{}^{34}SO_2^-$, which led to the loss of this ion and diminished sensitivity at higher levels of ambient water vapor (Arnold and Viggiano, 2001; Slusher et al., 2001). However, the SO_2 sensitivity at $F_2{}^{34}SO_2^-$ only varied within a factor of two for the entire field study with a clear relationship to water vapor (Fig. S2c). The SO_2 sensitivity did not show any obvious dependence on ambient O_3 concentrations (Fig. S2d).

The formic ($HCOO^-$ at m/z 45 and $HCOO^-$•HF at m/z 65) and acetic (CH_3COO^- •HF at m/z 79) acid ions did not show any obvious dependence on ambient water vapor and O_3 concentrations during calibration measurements (Fig. S3). Therefore, we do not expect the sensitivities of the X^- and X^-•HF ions of the studied organic acids to depend on ambient water vapor and O_3 concentrations. We accounted for water vapor dependence of the $F_2{}^{34}SO_2^-$ ion signal in our post-field calibrations where the response of the CIMS acid signals were measured relative to the of the $^{34}SO_2$ sensitivity.

3.1.3. Background and calibration measurements

Figure S4 shows an example of the CIMS instrument response during the switch between background, calibration and ambient measurements of formic and acetic acids during the field study. The 13 s time resolution data was used to determine the CIMS instrument time response. Formic (m/z 45, 65 and 108) and acetic (m/z 79) acid ion signals took ~1.5 min to reach a steady state after switches between ambient, calibration and background measurements (Figs. S4a and S4c).

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The decays in the formic and acetic acid ion signals and times required for them to reach steady state after the removal of calibration gases during the switch from standard addition calibration to ambient sampling were used to determine the CIMS response time. The signal decays were fitted using double exponential functions. For formic acid, the m/z 45, 65 and 108 ion signals decayed to $1/e^2$ in 37 ± 2, 33 ± 2 and 32 ± 2 s, respectively (Fig. S4b). For acetic acid, the m/z 79 ion signal decayed to $1/e^2$ in 42 ± 2 s (Fig. S4d).

3.2. Ambient measurements

3.2.1. Formic acid

Figure 2 shows typical mass spectra obtained under background and measurement modes during the field study. The SF_6^- reagent ion is present at m/z 146. One of the prominent species in the mass spectrum is formic acid, which is detected as $HCOO^-$ and $HCOO^-$•HF at m/z 45 and 65, respectively. Our laboratory studies demonstrated that the reaction of formic acid with SF_6^- also produced a large fraction of SF_4^- ions at m/z 108. The reaction of SF_6^- with oxalic acid also produced SF_4^- ions, but its SF_4^- product ion yield is low and gas phase oxalic acid is not present in large concentrations. In addition, SF_4^- is present in the mass spectrum obtained under background mode but the SF_4^- background ion signals are lower than those typically observed in measurement mode at the Yorkville site. As a result, we determined the ambient formic acid concentrations using the $HCOO^-$, $HCOO^-$•HF and SF_4^- ions. Figure 3a shows a scatter plot comparing the ambient formic acid concentrations measured at Yorkville using the $HCOO^-$, $HCOO^-$•HF and SF_4^- ions. Linear regression analysis reveals that the formic acid concentrations determined by the three ions are highly correlated ($R^2 = 0.99$) with slopes exhibiting a near 1:1 correlation. The excellent correlation between these three ions and the agreement with laboratory data indicates that formic acid is selectively measured by this method.

The time series of formic acid, temperature and solar radiation measured at Yorkville are shown in Fig. 3b. Formic acid concentrations ranged from 40 ppt to 4 ppb during the field study, with strong and consistent diurnal trends. The day-to-day variability in formic acid concentrations are associated with changes in solar radiation and temperature. Higher formic acid concentrations are measured during warm and sunny days,

similar to formic acid measurements performed in Centreville, rural Alabama during the 2013 Southern Oxidant Aerosol Study (SOAS) (Brophy and Farmer, 2015; Millet et al., 2015). Figure 3c shows the study-averaged diurnal profiles of formic acid and solar irradiance. Formic acid started to increase at 7:30, which coincided with a sharp increase in solar irradiance. Concentrations continued to increase throughout the day and peaked at 18:30, which coincided with the approximate time just before solar irradiance reached zero. Formic acid then decreased continuously throughout the night.

The immediate early-morning increase in formic acid observed in this field study is similar to that seen during the SOAS study (Millet et al., 2015). However, there are some differences in the formic acid diurnal cycles measured in this field study and the SOAS study. Formic acid peaked at 15:30 during SOAS, approximately 3 hours before solar irradiance decreased to zero. In contrast, formic acid concentrations only started to decrease at sunset (at 19:30) in this study. This suggests that there may be differences in the types and/or magnitudes of formic acid sources and sinks in this two field studies. Land cover and/or land use differences may have contributed to differences in formic acid sources and sinks at the Centreville and Yorkville field sites. The area surrounding the Yorkville field site is covered primarily by hardwood mixed with farmland and open pastures. In contrast, the Centreville field site is surrounded by forests comprised of mixed oak-hickory and loblolly trees (Hansen et al., 2003). It is also possible that seasonal differences contributed to differences in formic acid sources and sinks in the two field studies. The SOAS campaign took place in the middle of summer (1 June to 15 July 2013) when biogenic emissions are typically higher while this field study took place in early fall when biogenic emissions are lower due to cooler temperatures. For example, the average concentration of isoprene (a formic acid source) in this study (1.21 ppb) is lower than that in SOAS (1.92 ppb (Millet et al., 2015)). Despite these differences, our overall results are similar to the formic acid measurements performed in SOAS in both magnitude and diurnal variability.

3.2.2. Acetic acid

Acetic acid is detected with SF_6^- as CH_3COO^- and CH_3COO^-•HF at m/z 59 and 79, respectively. However, these ions are subject to interferences from the reaction of SF_6^- with

water vapor present in the sampled ambient air. Two of these interfering ions $F^-\bullet(HF)_2$ and $F^-\bullet(HF)_3$ occur at m/z 59 and 79, respectively. As discussed earlier, we minimized the impact of these interferences by diluting the sample flow into the CIMS and running the CDC at a high collision energy to dissociate the HF cluster ions. As expected from cluster bond strengths, we found that larger HF cluster ions dissociated more easily than smaller ones. For example, at a CDC electric field of ~113 V cm^{-1} (the configuration used in this field study), virtually all of the $F^-\bullet(HF)_3$ cluster ions dissociated while very few of the $F^-\bullet(HF)$ cluster ions dissociated. This indicates that the m/z 79 channel for acetic acid is more immune to interference from water vapor than the m/z 59 channel. This is supported by the observation that the background ion signal at m/z 59 ($R^2 = 0.50$) is more highly correlated with ambient water vapor concentrations than the background ion signal of m/z 79 ($R^2 = 0.30$). In addition, the m/z 59 ion is subjected to interference from the reaction of SF_6^- with O_3 present in the sampled ambient air. SF_6^- reacts with O_3 in the presence of CO_2 to form CO_3^- at m/z 60 (Slusher et al., 2001). As shown in Fig. 2, the large CO_3^- peak at m/z 60 is a potential interference to the m/z 59 signal. As the background scrubber also removed O_3 from the ambient air, there is a large difference in the m/z 60 ion signal between the measurement and background modes (~100,000 Hz). Thus, even a few percent bleed over of m/z 60 to m/z 59 can lead to an over-estimation of ambient acetic acid concentrations. For these reasons, we used m/z 79 ($X^-\bullet HF$) to determine ambient acetic acid concentrations even though this channel has a lower sensitivity than the m/z 59 channel (X^-).

The time series of acetic acid, temperature and solar radiation measured at Yorkville are shown in Fig. 4a. Acetic acid concentrations ranged from 30 ppt to 3 ppb during the field study. The day-to-day variability in acetic acid concentrations resembled the behavior of formic acid concentrations, with higher concentrations being measured during warm and sunny days. Figure 4b shows the study-averaged diurnal profiles of acetic acid and solar irradiance. The diurnal profile of acetic acid is similar to that of formic acid with a more pronounced evening maximum. Acetic acid started to increase at 7:30 and built up through the day, peaking at 19:30 and decreased continuously overnight. In general, acetic acid concentrations are well correlated with ($R^2 = 0.67$) and comparable in magnitude (~60 % on average) to formic acid. The study-averaged formic acid/acetic acid concentration ratio (1.65) is comparable to ratios from previous field studies in rural and

526 urban environments (Talbot et al., 1988; Talbot et al., 1995; Granby et al., 1997; Khare et
527 al., 1999; Talbot et al., 1999; Baboukas et al., 2000; Singh et al., 2000; Kuhn et al., 2002;
528 Baasandorj et al., 2015; Millet et al., 2015).

529 **3.2.3. Larger organic acids**

530 In addition to formic acid and acetic acid, eight other ions were monitored during
531 the field study: m/z 73, 75, 87, 89, 101, 103, 117 and 131. These ions were chosen as they
532 had significant signals when ambient air was sampled and were not obviously formed from
533 SF_6^- reaction with water vapor or O_3. Since the CIMS utilized in this study only had unit
534 mass resolution, these ions are the sum of all organic acid isomers and isobaric organic
535 acids of the same molecular weight as well as other product ions from species that might
536 react with SF_6^-. However, real-time ion chromatography measurements of aerosol
537 composition performed during the field study demonstrated the presence of particulate
538 oxalic, malonic, succinic and glutaric acids (Nah et al., 2018). For this reason, for m/z 89,
539 103, 117 and 131 ions, we assigned them as X^- ions of oxalic, malonic, succinic and glutaric
540 acids, respectively. As these organic acids have low vapor pressures, their gas-phase
541 concentrations are expected to be lower than their particle-phase concentrations, though
542 their gas-particle ratios will depend on thermodynamic conditions (Nah et al., 2018).
543 Particulate formic acid and acetic acid were also detected by ion chromatography during
544 the field study, but were at much lower concentrations relative to the gas phase (Nah et al.,
545 2018). For simplicity, we also denoted m/z 73, 75, 87 and 101 ions as X^- ions of propionic,
546 glycolic, butyric and valeric acids, respectively, for the remainder of this paper. These
547 organic acids have previously been measured in rural and urban environments (Kawamura
548 et al., 1985; Veres et al., 2011; Brophy and Farmer, 2015). However, we note that these
549 assignments are speculative. Post-field calibration measurements were used to estimate the
550 ambient concentrations of these organic acids.

551 Figure 5 shows the time series and diurnal profiles of oxalic, butyric, glycolic,
552 propionic and valeric acids measured during the field study. Daytime concentrations of
553 these organic acids ranged from a few tens of ppt to several hundred ppt. The time series
554 of ion signals (Hz) of malonic, succinic and glutaric acids normalized to the instrument's
555 sensitivity to $F_2{}^{34}SO_2$ (Hz ppb^{-1}) are shown in Fig. S3. The ion signals of these organic

acids are 1 to 2 orders of magnitude smaller than the sensitivity of the $F_2^{34}SO_2^-$ ion (study-averaged sensitivity = 2928 ± 669 Hz ppb^{-1}), resulting in these ratios to be less than 1. Concentrations of these organic acids are not available since calibrations were not performed for these compounds. The eight organic acids displayed very similar day-to-day variability as formic and acetic acids, with higher concentrations (or ion signals) being measured on warm and sunny days. The diurnal profiles of all the measured organic acids have similar diurnal trends, with their concentrations reaching a maximum between 17:30 and 19:30 and rapidly decreasing after sunset.

3.2.4. Comparison with $WSOC_g$

$WSOC_g$ measurements were performed during the field study using a MIST chamber coupled to a TOC analyzer. The study average $WSOC_g$ was 3.6 ± 2.7 μgC m^{-3}, slightly lower than that measured during the SOAS study (4.9 μgC m^{-3}) (Xu et al., 2017), and approximately four times lower than that measured in urban Atlanta, Georgia (13.7 μgC m^{-3}) (Hennigan et al., 2009). Despite being comparable in magnitude, the diurnal profiles of $WSOC_g$ measured in this study and the SOAS study are different. $WSOC_g$ measured in the SOAS study decreased at sunset, while $WSOC_g$ measured in this study decreased 2 hours after sunset. Differences in $WSOC_g$ concentrations and diurnal profiles at the three different sites may be due to differences in emission sources as a result of different measurement periods, land use and/or land cover.

To estimate the fraction of $WSOC_g$ that is comprised of organic acids, the total organic carbon contributed by formic, acetic, oxalic, butyric, glycolic, propionic and valeric acids is compared to the $WSOC_g$ measurements. We emphasize that the ion peak assignment of some of these organic acids are speculative. Hence, this comparison primarily serves as a zeroth order check to determine if the peak assignments are plausible. Figures 6a and 6b show the time series and diurnal profiles of $WSOC_g$ and the organic carbon contributed by the measured organic acids. Formic and acetic acids comprised majority of the total organic carbon contributed by the measured organic acids (study averages of 31 and 38 %, respectively). Assuming that the ion peak assignments are correct and the measured organic acids are completely water-soluble, the carbon mass fraction of $WSOC_g$ comprised of these organic acids ranged from 7 to 100 %. Based on the orthogonal

586 distance regression slope shown in Fig. 6c, the study-averaged carbon mass fraction of
587 $WSOC_g$ comprised of the measured organic acids is 30 %. The total organic carbon
588 contributed by the measured organic acids are moderately correlated with $WSOC_g$ (R^2 =
589 0.38). This is likely due to the presence of other water-soluble gas phase species (with
590 different day-to-day variability from the organic acids) that contribute to the $WSOC_g$. This
591 is supported by slight differences in the diurnal profiles of $WSOC_g$ and the organic carbon
592 contributed by the organic acids (Fig. 6b). While the diurnal profiles of $WSOC_g$ and the
593 organic carbon contributed by the organic acids have similar general shapes, $WSOC_g$
594 peaked at 21:30, approximately 2 hours after the solar irradiance have decreased to zero.
595 In contrast, the organic carbon contributed by the organic acids start to decrease at sunset
596 (at 19:30).

597 **3.2.5. SO_2 and HNO_3 observations**

598 In addition to evaluating the field performance of the SF_6^- CIMS technique in gas-
599 phase organic acid measurements, another focus of this study was to investigate the
600 possible sources of the measured organic acids. For this reason, HNO_3 and SO_2 (two
601 common anthropogenic tracers) were also measured by SF_6^- CIMS during the field study.
602 Correlations between the concentrations of organic acids and those of HNO_3 and SO_2 were
603 then examined to determine if the organic acids were anthropogenic in nature (section 3.3).
604 While their reactions with SF_6^- have multiple product channels (Huey et al., 1995), only
605 the NO_3^-•HF (m/z 82) and $F_2SO_2^-$ (m/z 102) ions were used for quantitative purposes:

606 $$SF_6^- + HNO_3 \rightarrow NO_3^-\bullet HF + SF_5 \quad (4)$$

607 $$SF_6^- + SO_2 \rightarrow F_2SO_2^- + SF_4 \quad (5)$$

608 Figure S4 shows the time series of SO_2 and HNO_3 measured during the field study.
609 As expected at a rural site, SO_2 and HNO_3 concentrations are low most of the time (study
610 averages of 230 and 180 ppt, respectively). However, there were occasional periods when
611 the site was impacted by anthropogenic pollution. In particular, there are spikes in both
612 SO_2 and HNO_3 concentrations throughout the study that corresponded to the site being
613 impacted by power plant or urban emissions. Outside of these anthropogenic spikes, HNO_3

showed a clear diurnal profile with a maximum at approximately 12:30, consistent with local photochemical production.

3.3. Potential sources of organic acids

Correlation analysis on organic acid concentrations can provide insights on their sources. Figure 7 shows that the concentration of formic acid is strongly correlated with those of the other measured organic acids (R^2 = 0.68 to 0.89). This suggests that these organic acids have the same or similar sources at Yorkville. The sources of organic acids can be biogenic or anthropogenic in nature. To determine if the primary sources of organic acids are of biogenic or anthropogenic origin, we first examined the correlations of organic acid concentrations with those of anthropogenic pollutants CO, SO_2, O_3 and HNO_3. CO and SO_2 are common tracers for combustion sources. The organic acid concentrations are poorly correlated with CO (Fig. S5, R^2 = 0.03 to 0.15) and SO_2 (Fig. S6, R^2 = 0.01 to 0.24), indicating that primary emissions from combustion are a minor source of organic acids in Yorkville. HNO_3 and O_3 are common photochemical tracers of urban air masses. The organic acid concentrations are weakly correlated with O_3 (Fig. S7, R^2 = 0.11 to 0.32) and HNO_3 (Fig. S8, R^2 = 0.33 to 0.56). In addition, there is no noticeable increase in organic acid concentrations during periods of elevated CO, SO_2, O_3 and HNO_3 concentrations when the site was impacted by pollution plumes. Together, these results indicate that the primary sources of organic acids in Yorkville are likely not anthropogenic in nature.

Diurnal profiles of the measured organic acids suggest that their sources are linked to higher daytime temperatures and/or photochemical processes. Figure 8 compares the concentrations of organic acids against ambient temperatures measured during the study. Since there was a noticeable decrease in mean ambient temperatures starting on 28 Sept 2016, we grouped the datasets into two time periods (3 to 27 Sept and 28 Sept to 12 Oct) to better evaluate the effect of temperature on organic acid concentrations. The average temperature in the first time period (3 to 27 Sept) is 24.8 °C (32.6 °C max, 18.1 °C min), while the average temperature in the second time period (28 Sept to 12 Oct) is 19.5 °C (28.4 °C max, 9.5 °C min). We find that organic acid concentrations are on average higher and more highly correlated with temperatures in the warmer first time period (R^2 = 0.40 to

643 0.63) compared to the cooler second time period (R^2 = 0.18 to 0.59). These observations
644 can be explained by temperature-dependent emissions of organic acids and their BVOC
645 precursors. Previous studies have shown that emissions of organic acids and their BVOC
646 precursors depend strongly on light and temperature, with substantially lower
647 concentrations being emitted in the dark and/or at low temperatures (Kesselmeier et al.,
648 1997; Kesselmeier, 2001; Sindelarova et al., 2014). We find that the concentration of
649 isoprene, which was the dominant BVOC in Yorkville, has a somewhat similar diurnal
650 profile as the organic acids and decreased with temperature on 28 Sept 2016 (Fig. S9). In
651 addition, the concentrations of formic and acetic acids are moderately correlated with that
652 of isoprene (R^2 = 0.42 and 0.40, respectively) (Fig. S10).

653 Multiphase photochemical aging of ambient organic aerosols can also be a source
654 of gas-phase organic acids (Eliason et al., 2003; Ervens et al., 2004; Molina et al., 2004;
655 Lim et al., 2005; Park et al., 2006; Walser et al., 2007; Sorooshian et al., 2007; Vlasenko
656 et al., 2008; Pan et al., 2009; Sorooshian et al., 2010). Organic acids may be formed in the
657 particle phase during organic aerosol photochemical aging, with subsequent volatilization
658 into the gas phase. Real-time ion chromatography measurements of aerosol composition
659 demonstrated the presence of particulate formic, acetic, oxalic, malonic, succinic and
660 glutaric acids (Nah et al., 2018). However, since the ratios of gas-phase formic and acetic
661 acid mass concentration to the total organic aerosol mass concentration are large (study
662 averages of 40 and 35 %, respectively) (Nah et al., 2018), it is unlikely that organic aerosol
663 photochemical aging is a large source of formic and acetic acids. In contrast, the ratios of
664 gas-phase oxalic, malonic, succinic and glutaric acids mass concentration to the total
665 organic aerosol mass concentration are small, suggesting that organic aerosol
666 photochemical aging may be an important source of these gas-phase organic acids.

667 In summary, the temperature dependence and diurnal profile of organic acid
668 concentrations combined with poor correlations between organic acid concentrations and
669 those of anthropogenic pollutants CO, SO_2, O_3 and HNO_3 strongly suggest that the primary
670 sources of gas-phase organic acids at Yorkville are biogenic in nature. However, our data
671 alone does not allow us to determine if the organic acids are a result of direct emissions or
672 photochemical oxidation of other BVOC emissions and/or organic aerosols. Partitioning

673 of these organic acids between the gas and particle phases will be discussed in a
674 forthcoming paper (Nah et al., 2018).

675 **4. Summary**

676 SF_6^- reacted with all of the studied organic acids to produce product ions that were
677 characteristic of the individual acids (i.e., X^- or X^-•HF). These reactions all occurred at less
678 than the maximum collisional rate due to significant yields of SF_5^- and SF_4^-, which reduced
679 the sensitivity of the method. For the conditions employed in this study, the sensitivities of
680 X^- and X^-•HF ions of the organic acids relative to that of the $F_2{}^{34}SO_2^-$ ion (study-averaged
681 sensitivity 2928 ± 669 Hz ppb^{-1}) ranged from 0.04 to 2.18. The detection limits of the
682 organic acids were approximated from 3 times the standard deviation values (3σ) of the ion
683 signals obtained during background measurements. Reasonable limits of detection for 2.5
684 min integration periods (1 to 60 ppt) were obtained for all the organic acids studied. Water
685 vapor and O_3 can lead to interferences with this method but for the conditions employed in
686 this study, they were largely limited to acetic acid measurements at m/z 59. However,
687 fluctuations in ambient water vapor can also lead to changes in sensitivity for the detection
688 of some species (e.g., SO_2). Uncertainties in organic acid concentrations originate primarily
689 from calibration measurements and ranged from 9 to 22 %. Overall, the tractable mass
690 spectra obtained by the SF_6^- CIMS method coupled with reasonable limits of detection and
691 the high correlations observed between the individual organic acids demonstrated the
692 potential of this method. Obvious next steps for the SF_6^- CIMS method are to compare it
693 to other measurement methods for organic acids and to deploy the SF_6^- ion chemistry to a
694 higher resolution time-of-flight mass spectrometer to reduce the potential for interferences.

695 The SF_6^- CIMS method was deployed for measurements of gas phase organic acids
696 in a mixed forest-agricultural area in Yorkville, Georgia from Sept to Oct 2016. The
697 organic acids measured in the field study were formic and acetic acids. In addition,
698 measurements tentatively assigned to oxalic, butyric, glycolic, propionic, valeric, malonic,
699 succinic and glutaric acids were performed. Ambient concentrations of these organic acids
700 ranged from a few ppt to several ppb. All the organic acids exhibited similar strong diurnal
701 trends. Organic acid concentrations built up throughout the day, peaked between 17:30 and
702 19:30 before decreasing continuously overnight. Strong correlations between organic acid

concentrations indicated that these organic acids likely have the same or similar sources at Yorkville. We concluded that the organic acids were likely not due to anthropogenic emissions since they were poorly correlated with anthropogenic pollutants and their concentrations were not elevated when the site was impacted by pollution plumes. Higher organic acid concentrations were measured during warm and sunny days. Organic acid concentrations were strongly correlated with temperature during the first month of the study when ambient temperatures were high. Together, our results suggested that the primary sources of organic acids at Yorkville were biogenic in nature. Direct biogenic emissions of organic acids and/or their BVOC precursors were likely enhanced at high ambient temperatures, resulting in the observed variability of organic acid concentrations. Another potential source is the production of organic acids in the particle phase from the multiphase photochemical aging of organic aerosols followed by evaporation to the gas phase, though this source is likely not a large source of formic and acetic acids. However, given the inability of current models and photochemical mechanisms to explain formic acid observations in the Southeastern U.S. (Millet et al., 2015), it is unlikely that our observations of formic acid and larger organic acids can be explained as well. Further work (i.e., laboratory, field and modeling studies) is needed to determine how organic acids are formed in the atmosphere.

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7. Competing financial interests

The authors declare no competing financial interests.

8. Data availability

Data can be accessed by request (greg.huey@eas.gatech.edu).

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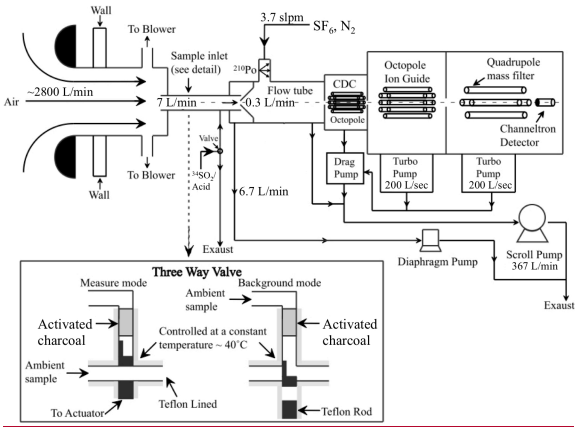

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972 **Figure 1:** The CIMS instrument and inlet configuration used in the field study. The
973 automated three-way sampling valve is shown in the inset. The figure was adapted from
974 Liao et al. (2011).

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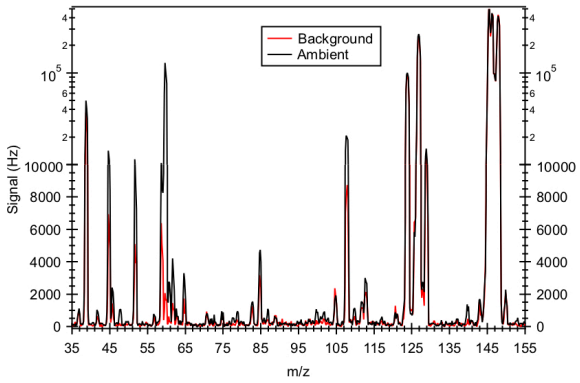
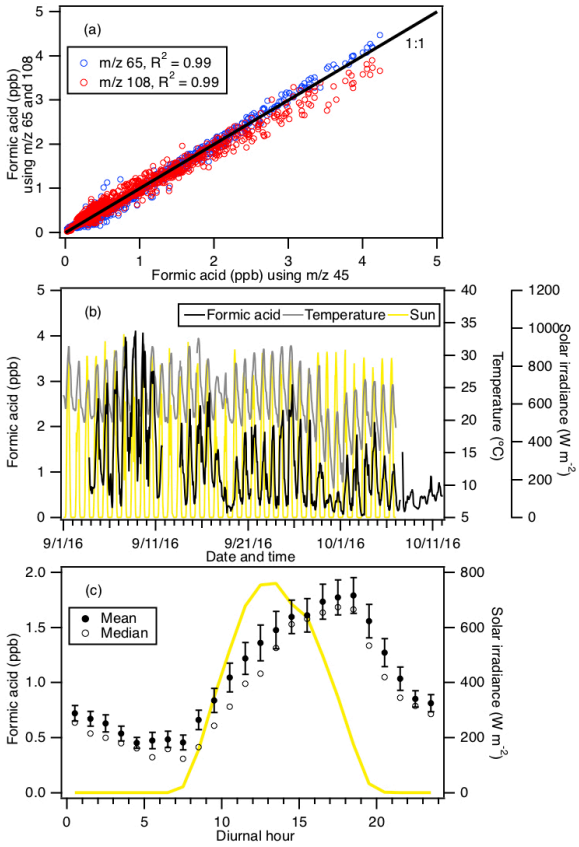


Figure 2: Mass spectrum of ambient air and background measured in Yorkville, Georgia on 8 Sept 2016 using SF_6^-. Note that the $^{32}SF_6^-$ reagent ion signal (at m/z 146) is saturated, causing the sharp drop in its signal. As a result, the ion signal of its isotope $^{34}SF_6^-$ (at m/z 150) was monitored to determine if the reaction of SF_6^- with ambient water vapor and O_3 depleted SF_6^- reagent ions.



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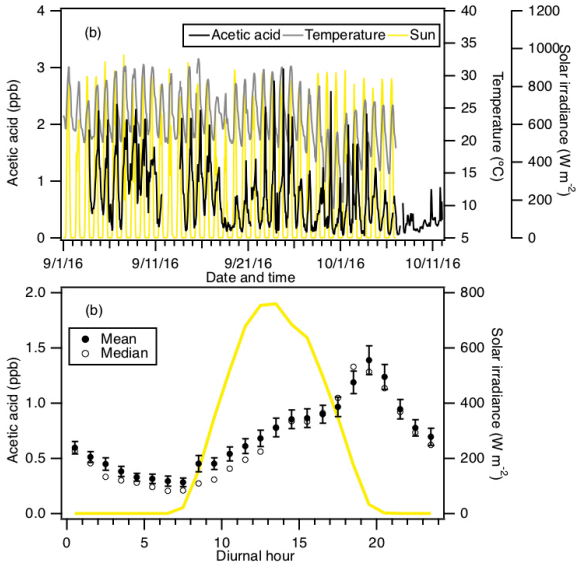
989 **Figure 3:** (a) Scatter plot comparison of ambient formic acid concentrations determined
990 using mass peaks m/z 45, 65 and 108. The three datasets correlated well with one another
991 (R^2 = 0.99). Linear regression of the data gave slopes of 1 (for m/z 65) and 0.95 (for m/z
992 108), indicating that all three mass peaks can be used to determine the formic acid
993 concentration. (b) Time series of formic acid concentration, temperature and solar
994 irradiance. All the data are displayed as 1-hour averages. (c) Diurnal profiles of formic acid
995 concentration (symbols) and solar irradiance (yellow line). All the concentrations represent
996 averages in 1-hour intervals and the standard errors are plotted as error bars.



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998 **Figure 4:** (a) Time series of acetic acid concentration, temperature and solar irradiance.
999 All the data are displayed as 1-hour averages. (c) Diurnal profiles of acetic acid (symbols)
1000 and solar irradiance (yellow line). All the concentrations represent averages in 1-hour
1001 intervals and the standard errors are plotted as error bars.

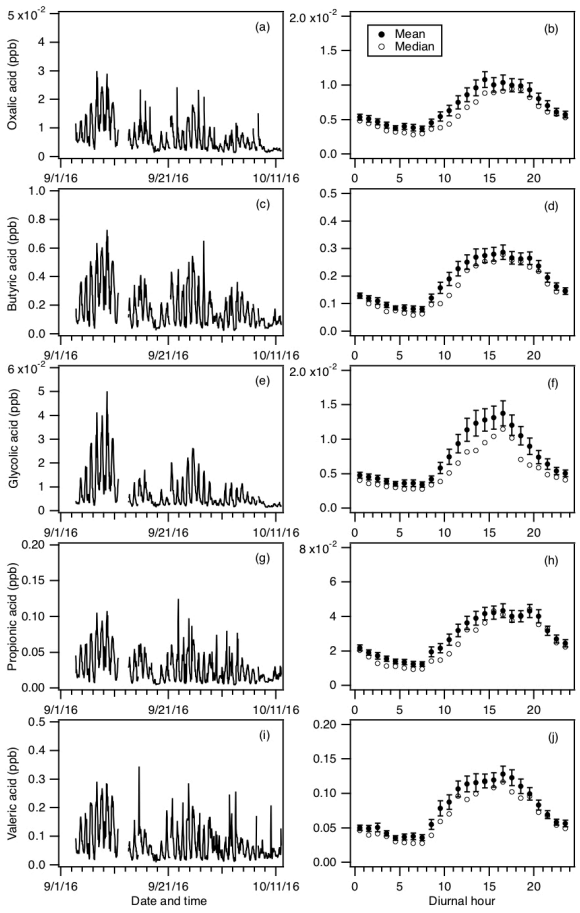
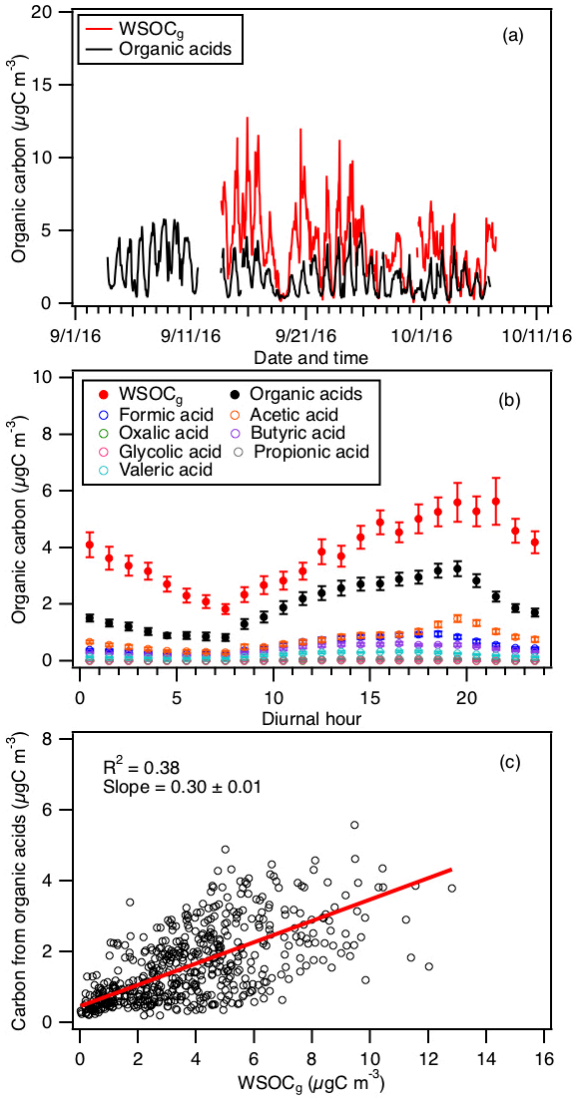


Figure 5: Time series of concentrations of (a) oxalic, (c) butyric, (e) glycolic, (g) propionic, and (i) valeric acids measured during the field study. All the data are displayed as 1-hour averages. Their corresponding diurnal profiles are shown in (b), (d), (f), (h) and (j), respectively. The diurnal profile concentrations represent averages in 1-hour intervals and the standard errors are plotted as error bars.



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1009 **Figure 6:** (a) Time series of $WSOC_g$ and the total organic carbon contributed by the
1010 measured organic acids. All the data are displayed as 1-hour averages. (b) Diurnal profiles
1011 of $WSOC_g$ and the total organic carbon contributed by the measured organic acids. Also
1012 shown are the diurnal profiles of the organic carbon contributed by the individual measured

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organic acids. All the concentrations represent the mean hourly averages and the standard errors are plotted as error bars. (c) Scatter plot of total organic carbon contributed by the measured organic acids with $WSOC_g$. Note that the ion peak assignment to some of these organic acids are speculative.

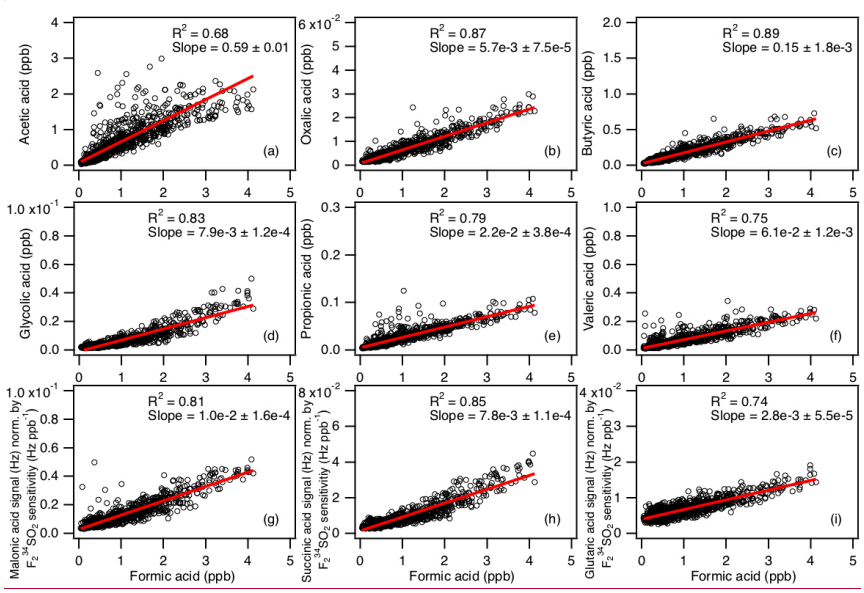

Figure 7: Scatter plots of concentrations (or signals) of (a) acetic, (b) oxalic, (c) butyric, (d) glycolic, (e) propionic, (f) valeric, (g) malonic, (h) succinic, and (i) glutaric acids with formic acid concentration. All the data are displayed as 1-hour averages. The data for malonic, succinic and glutaric acids are presented as the ratio of their ion signals (Hz) to the instrument's sensitivity to $F_2{}^{34}SO_2$ (Hz ppb^{-1}) since these organic acids were not calibrated. Red lines shown are linear fits to the data.

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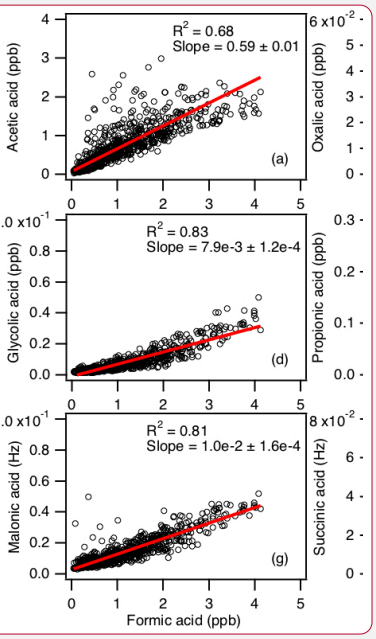

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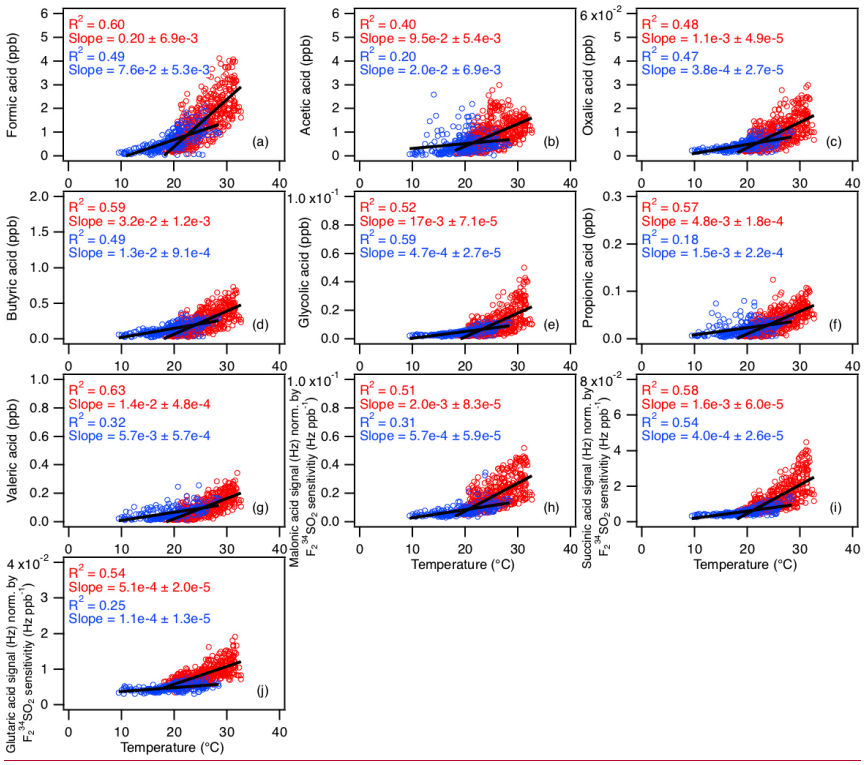

Figure 8: Scatter plots of concentrations (or signals) of (a) formic, (b) acetic, (c) oxalic, (d) butyric, (e) glycolic, (f) propionic, (g) valeric, (h) malonic, (i) succinic, and (j) glutaric acids with ambient temperature. The red symbols are data collected from 3 to 27 Sept, while the blue symbols are data collected from 28 Sept onwards. All the data are displayed as 1-hour averages. The data for malonic, succinic and glutaric acids are presented as the ratio of their ion signals (Hz) to the instrument's sensitivity to $F_2{}^{34}SO_2$ (Hz ppb^{-1}) since these organic acids were not calibrated. Black lines shown are linear fits to the datasets.

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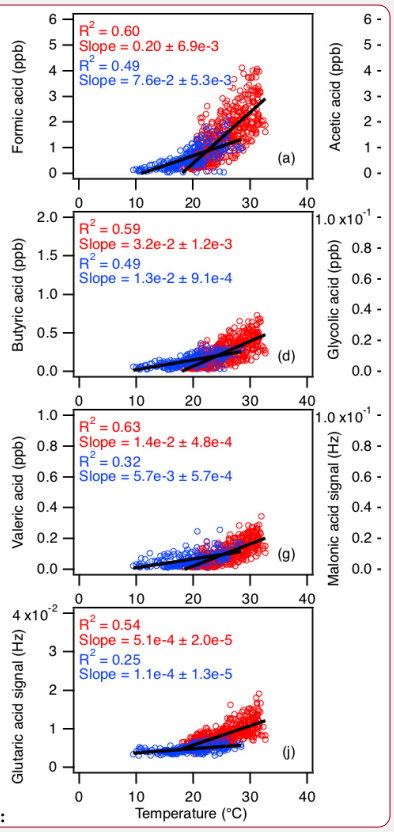

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Table 1: Summary of organic acids of interest, their detection limits and sensitivities of their X^- and X^-•HF ions[a]

Organic Acid	Detection limit (ppt)[b]	Sensitivity (Hz ppt^{-1})	
		X^-	X^-•HF
Formic acid	30	1.29 ± 0.22	0.29 ± 0.05
Acetic acid	60	1.46 ± 0.29	0.30 ± 0.06
Oxalic acid	1	6.38 ± 0.32	0.97 ± 0.05
Butyric acid	30	0.41 ± 0.01	0.12 ± 0.004
Glycolic acid	2	5.53 ± 0.11	1.64 ± 0.03
Propionic acid	6	2.05 ± 0.02	1.26 ± 0.01
Valeric acid	10	0.76 ± 0.008	0.35 ± 0.004

[a]Only organic acids with calibration measurements are shown.

[b]Detection limits are approximated from 3 times the standard deviation values (3σ) of the ion signals measured during background mode. Shown here are the average detection limits of the organic acids for 2.5 min averaging periods which corresponds to the length of a background measurement at a 4 % duty cycle for each mass.

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[c]Study-averaged sensitivity of the $F_2{}^{34}SO_2^-$ ion = 2928 ± 669 Hz ppb^{-1}

Supplementary Information:

Real-time measurements of gas-phase organic acids using SF_6^- chemical ionization mass spectrometry

Theodora Nah,[1] Yi Ji,[1,2] David J. Tanner,[1] Hongyu Guo,[1] Amy P. Sullivan,[3] Nga Lee Ng,[1,2] Rodney J. Weber[1] and L. Gregory Huey[1*]

[1]*School of Earth and Atmospheric Sciences, Georgia Institute of Technology, Atlanta, GA, USA*
[2]*School of Chemical and Biomolecular Engineering, Georgia Institute of Technology, Atlanta, GA, USA*
[3]*Department of Atmospheric Science, Colorado State University, Fort Collins, CO, USA*

** To whom correspondence should be addressed: greg.huey@eas.gatech.edu*

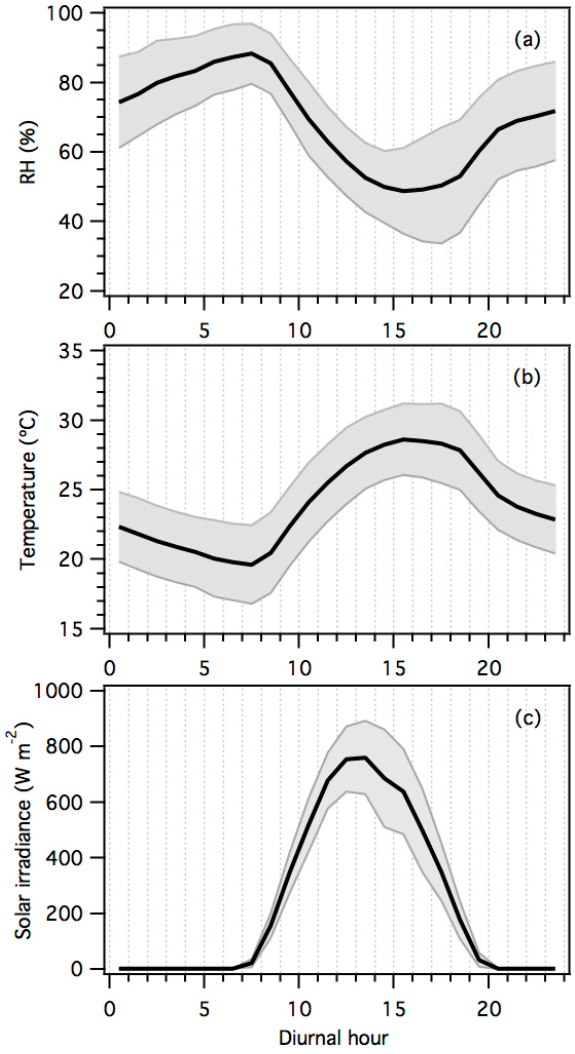

Figure S1: Diurnal trends of (a) relative humidity, (b) temperature, and (c) solar radiance. The lines within the shaded area represents the average values. The upper and lower boundaries of the shaded areas mark one standard deviation.

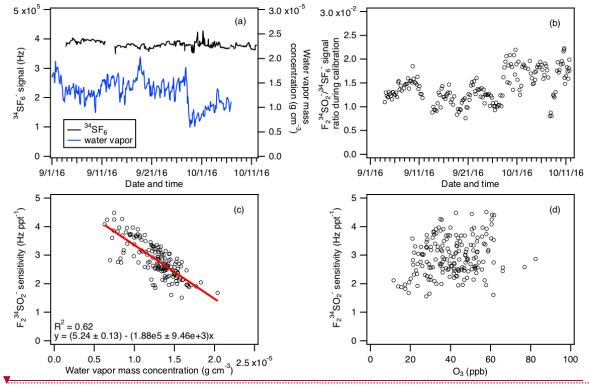

Figure S2: (a) Time series of $^{34}SF_6^-$ reagent ion signal and ambient water vapor concentration for the entire field study. The ambient water vapor mass concentrations are determined from ambient relative humidities and temperatures. (b) Time series of $F_2{}^{34}SO_2^-/{}^{34}SF_6^-$ ion signal ratio obtained during calibration measurements. Panels (c) and (d) show the $F_2{}^{34}SO_2^-$ ion sensitivity obtained from calibration measurements as a function of ambient water vapor and O_3 concentrations. Data in panels (a) to (d) are displayed as 1-hour averages.

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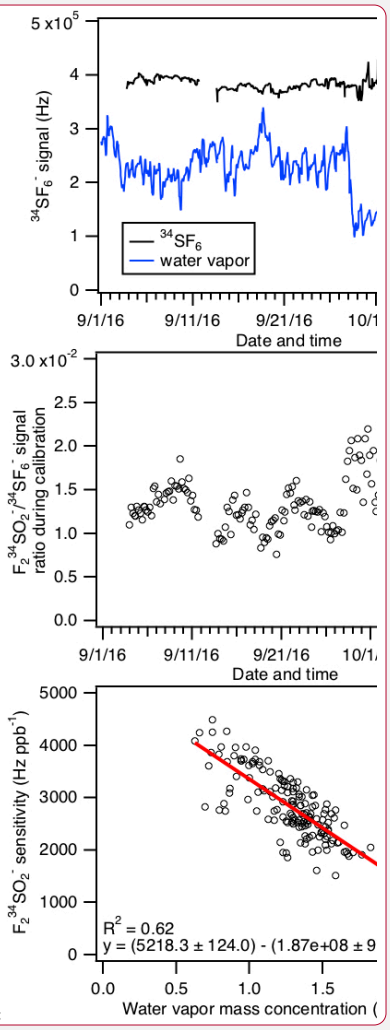


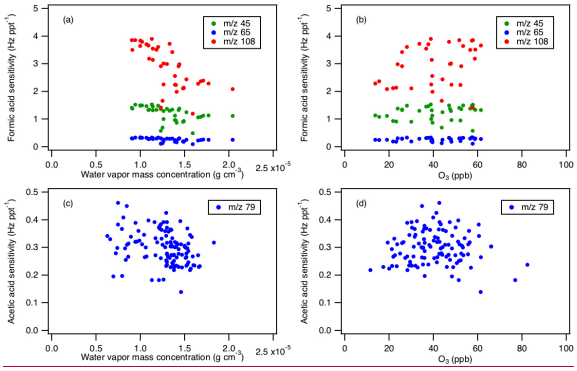


Figure S3: Panels (a) and (b) show the sensitivities of formic acid ions ($HCOO^-$ at m/z 45, $HCOO^-$•HF at m/z 65, and SF_4^- at m/z 108) obtained from calibration measurements as a function of ambient water vapor and O_3 concentrations. Panels (c) and (d) show the acetic acid sensitivity (CH_3COO^-•HF at m/z 79) obtained from calibration measurements as a function of ambient water vapor and O_3 concentrations.

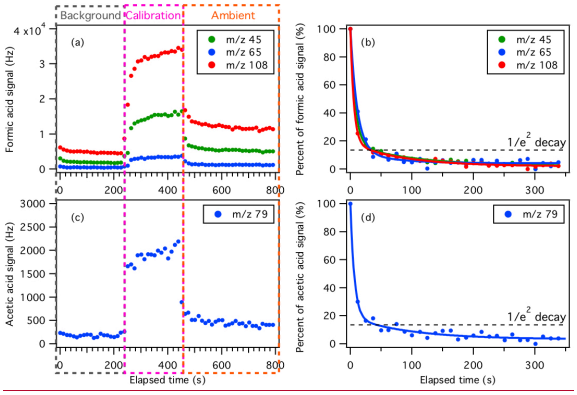
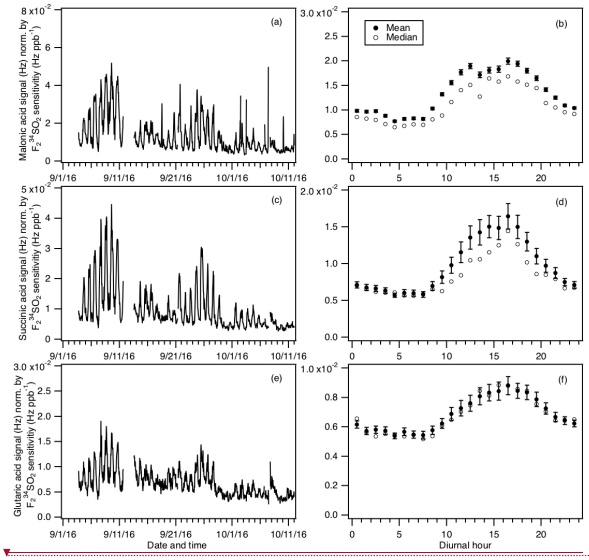


Figure S4: Example of the CIMS instrument response during switches between background, calibration and ambient measurements of (a) formic, and (c) acetic acids. Panels (b) and (d) show the percent of formic and acetic acid ion signals after the removal of a 6.75 ppb of formic acid and 5.87 ppb of acetic acid standard addition calibration as a function of time. The data shown here is 13 s time resolution data. Double exponential fits to each m/z ion are shown as colored solid lines. Black dashed lines show the times for the ions to decay to $1/e^2$.



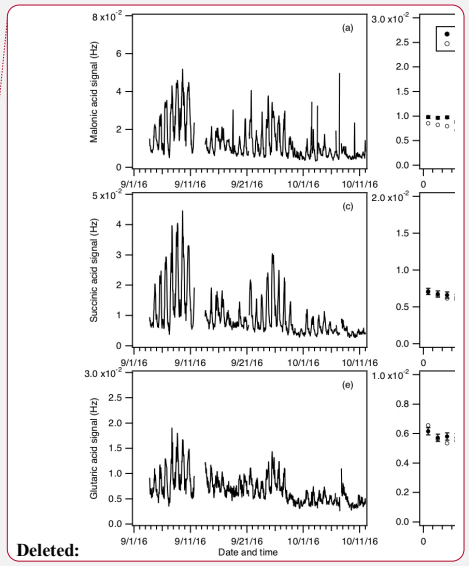


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Figure S5: Time series of signals of (a) malonic, (c) succinic, and (e) glutaric acids measured during the field study. The data are displayed as 1-hour averages. Their corresponding diurnal profiles are shown in (b), (d) and (f), respectively. All the signals represent averages in 1-hour intervals and the standard errors are plotted as error bars. These organic acids were not calibrated so all the signals are presented here as Hz normalized by the instrument's sensitivity to $F_2{}^{34}SO_2$ (Hz ppb^{-1}), which was the primary calibrant used in the field study.

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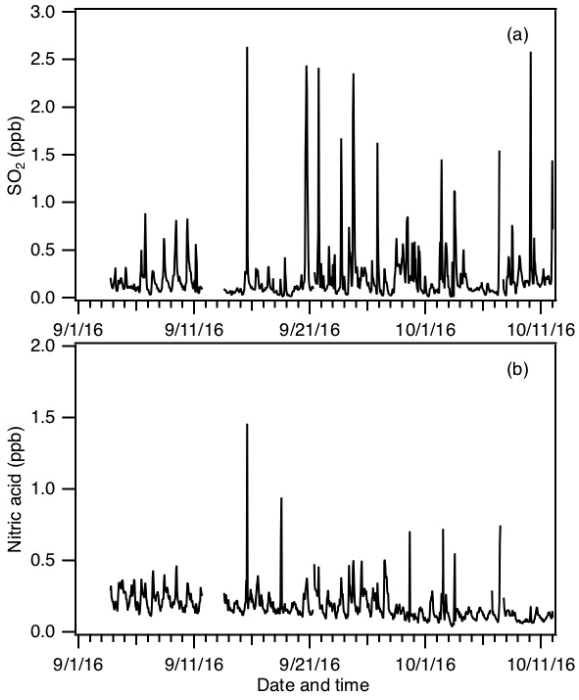

Figure S6: Time series of (a) SO_2 and (b) HNO_3 concentrations measured during the field study. All the data are displayed as 1-hour averages.

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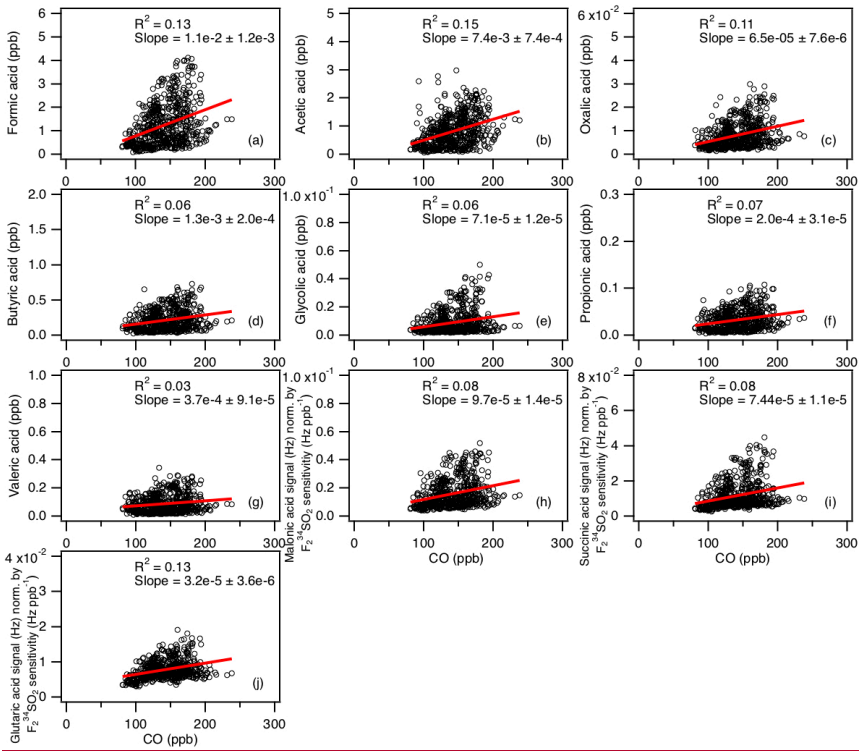

Figure S7: Scatter plots of concentrations (or signals) of (a) formic, (b) acetic, (c) oxalic, (d) butyric, (e) glycolic, (f) propionic, (g) valeric, (h) malonic, (i) succinic, and (j) glutaric acids with CO concentration. All the data are displayed as 1-hour averages. The data for malonic, succinic and glutaric acids are presented as the ratio of their ion signals (Hz) to the instrument's sensitivity to $F_2{}^{34}SO_2$ (Hz ppb^{-1}) since these organic acids were not calibrated. Red lines shown are linear fits to the data.

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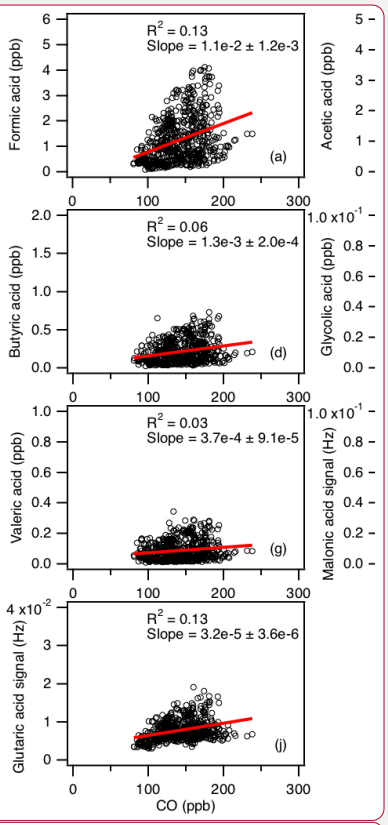

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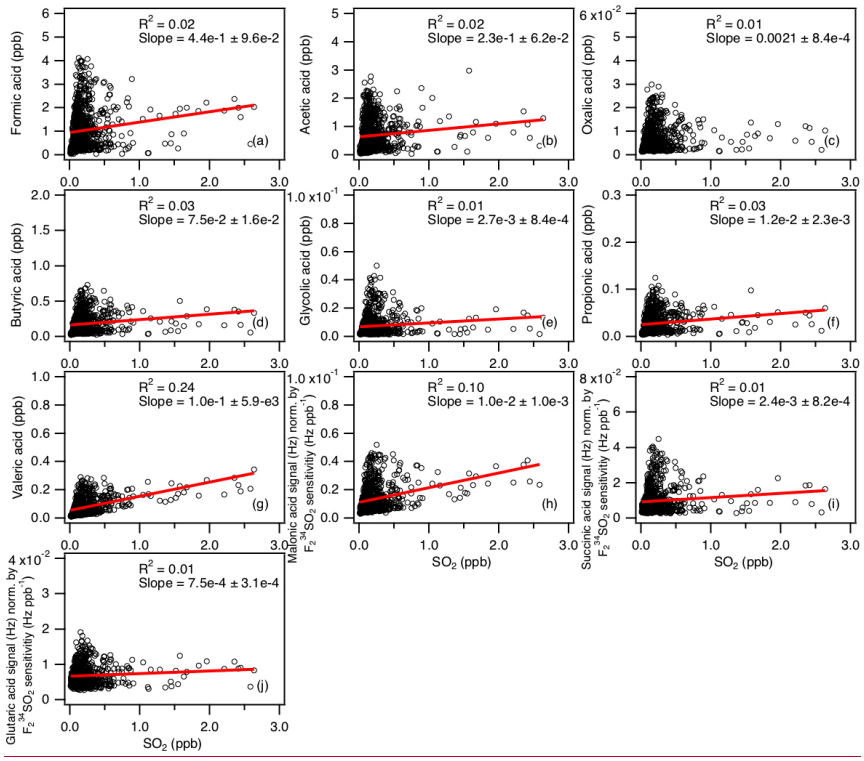
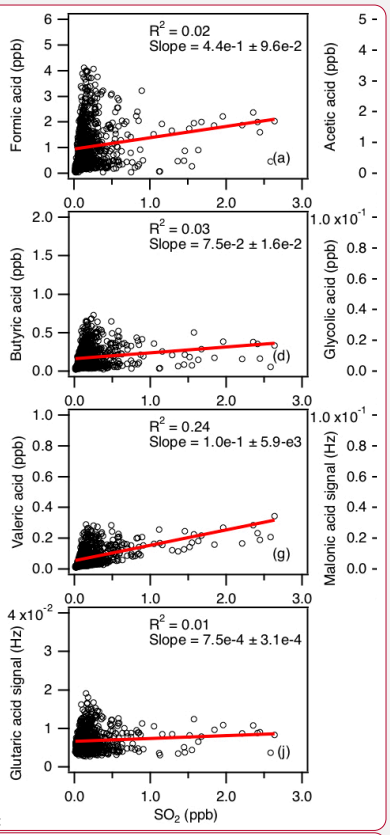


Figure S8: Scatter plots of concentrations (or signals) of (a) formic, (b) acetic, (c) oxalic, (d) butyric, (e) glycolic, (f) propionic, (g) valeric, (h) malonic, (i) succinic, and (j) glutaric acids with SO_2 concentration. All the data are displayed as 1-hour averages. The data for malonic, succinic and glutaric acids are presented as the ratio of their ion signals (Hz) to the instrument's sensitivity to $F_2{}^{34}SO_2$ (Hz ppb^{-1}) since these organic acids were not calibrated. Red lines shown are linear fits to the data.



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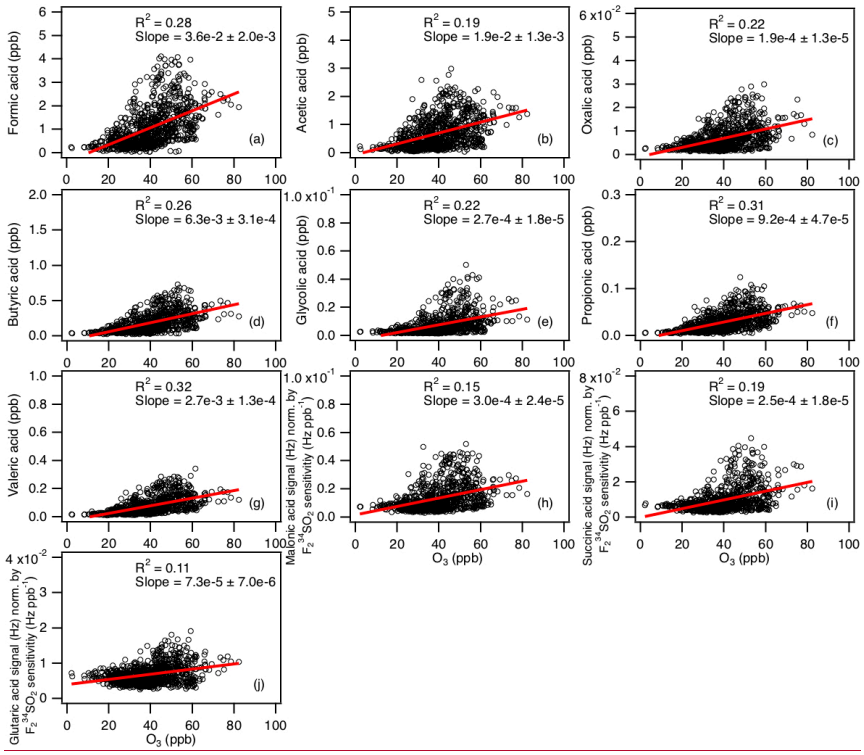
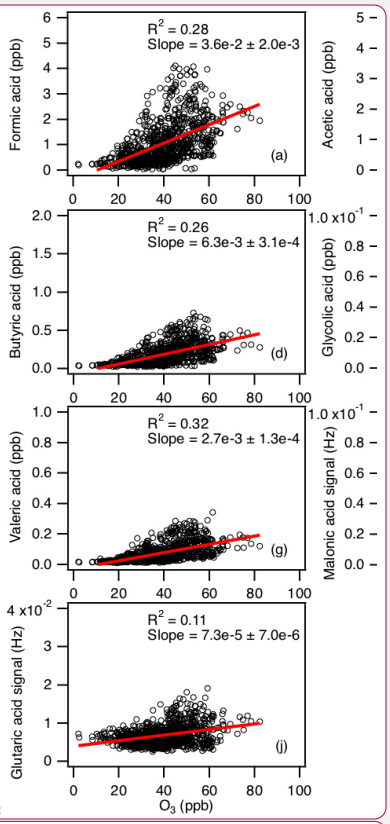


Figure S9: Scatter plots of concentrations (or signals) of (a) formic, (b) acetic, (c) oxalic, (d) butyric, (e) glycolic, (f) propionic, (g) valeric, (h) malonic, (i) succinic, and (j) glutaric acids with O_3 concentration. All the data are displayed as 1-hour averages. The data for malonic, succinic and glutaric acids are presented as the ratio of their ion signals (Hz) to the instrument's sensitivity to $F_2{}^{34}SO_2$ (Hz ppb^{-1}) since these organic acids were not calibrated. Red lines shown are linear fits to the data.



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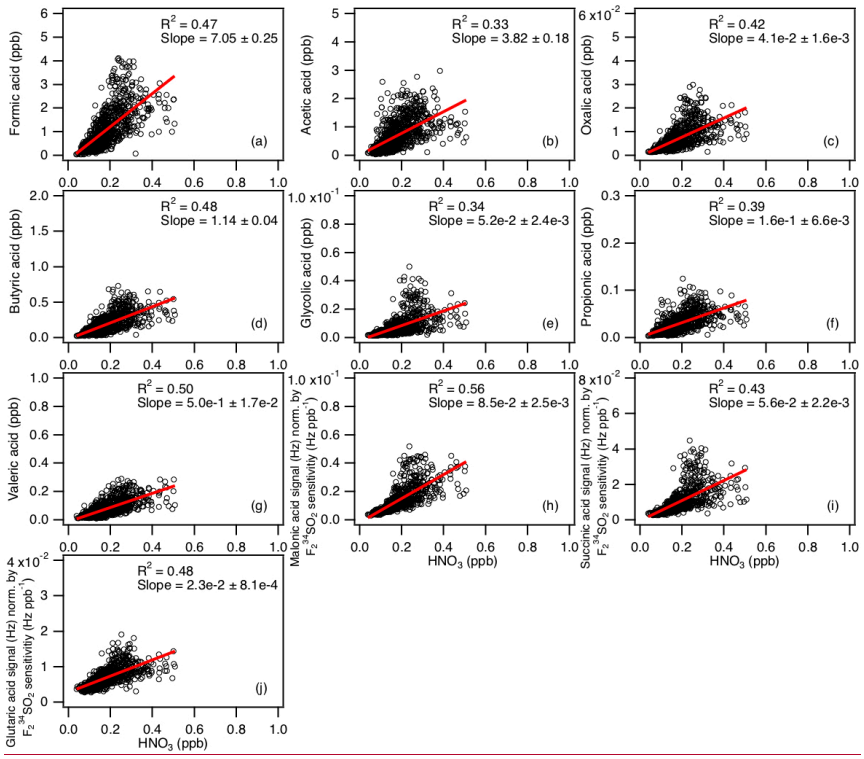
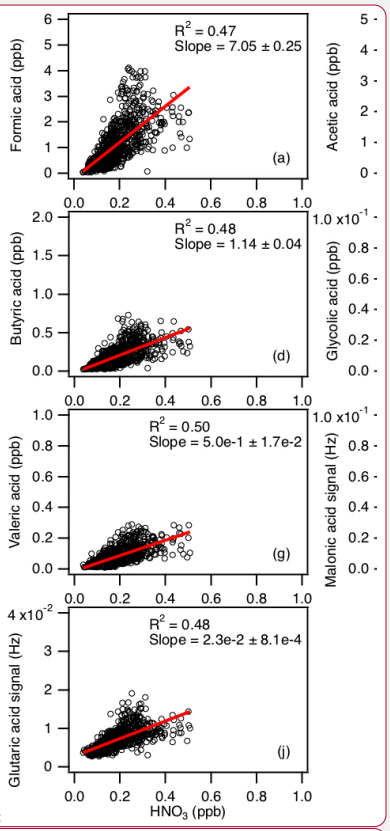


Figure S10: Scatter plots of concentrations (or signals) of (a) formic, (b) acetic, (c) oxalic, (d) butyric, (e) glycolic, (f) propionic, (g) valeric, (h) malonic, (i) succinic, and (j) glutaric acids with HNO_3 concentration. To exclude periods when the site was affected by urban or power plant emissions, data where HNO_3 > 0.5 ppb are excluded from these scatter plots. All the data are displayed as 1-hour averages. The data for malonic, succinic and glutaric acids are presented as the ratio of their ion signals (Hz) to the instrument's sensitivity to $F_2{}^{34}SO_2$ (Hz ppb^{-1}) since these organic acids were not calibrated. Red lines shown are linear fits to the data.



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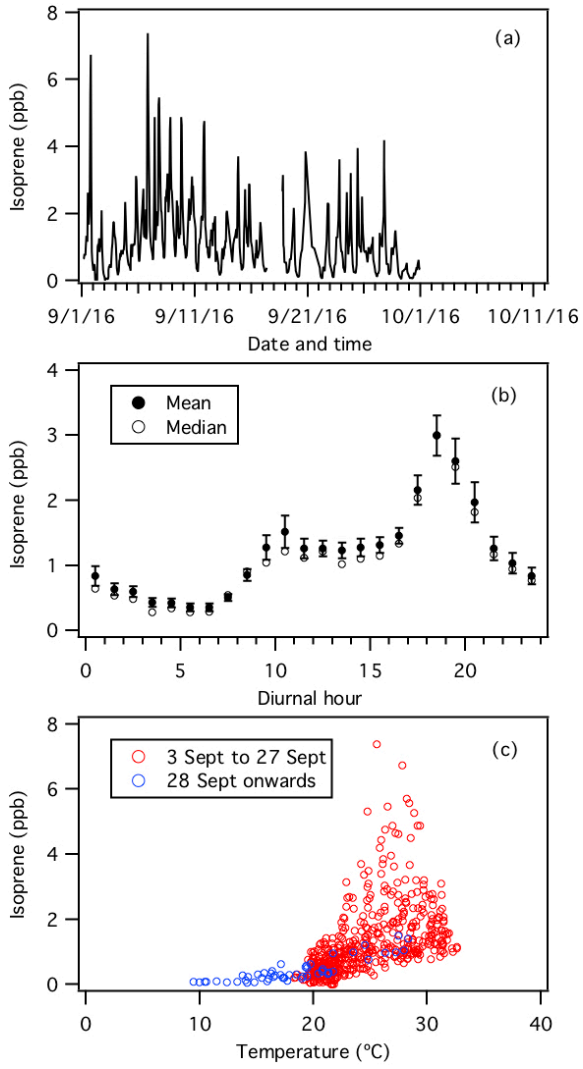

Figure S11: (a) Time series of isoprene concentration during the field study. (b) Diurnal profile of isoprene. All the concentrations represent averages in 1-hour intervals and the standard errors are plotted as error bars. (c) Scatter plot of isoprene concentration with ambient temperature. All the data are displayed as 1-hour averages.

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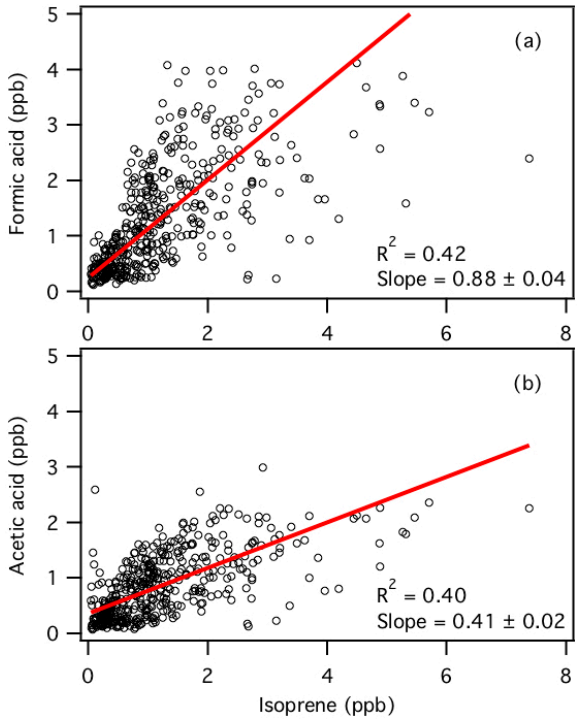

Figure S12: Scatter plots of concentrations of (a) formic and (b) acetic acids with isoprene concentration. All the data are displayed as 1-hour averages. Red lines shown are linear fits to the data.

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