



1 Multifactor colorimetric analysis on pH-indicator papers: an

2 optimized approach for direct determination of ambient

3 aerosol pH

- 4 Guo Li¹, Hang Su^{1*}, Nan Ma², Guangjie Zheng^{1†}, Uwe Kuhn¹, Meng Li¹, Thomas Klimach¹,
- 5 Ulrich Pöschl¹, Yafang Cheng^{1*}
- 6 Multiphase Chemistry Department, Max Planck Institute for Chemistry, Mainz, Germany
- 7 ² Institute for Environmental and Climate Research, Jinan University, Guangzhou, China
- 8 † Now at: Department of Energy, Environmental & Chemical Engineering, Center for Aerosol Science &
- 9 Engineering, Washington University, St. Louis, Missouri 63130, USA
- * Correspondence to: Y. Cheng (yafang.cheng@mpic.de) or H. Su (h.su@mpic.de)

11 12

13

14

15

16

17

18

19

20

21

22

23

24

25

26

Abstract

Direct measurement of the acidity (pH) of ambient aerosol particles/droplets has long been a challenge for atmospheric scientists. A novel and facile method was introduced recently by Craig et al. (2018), where the pH of size-resolved aerosol droplets was directly measured by two types of pH-indicator papers (pH ranges: 0-2.5 and 2.5-4.5) combined with RGB-based colorimetric analyses using a model of G-B (G minus B) versus pH². Given the wide pH range of ambient aerosols, we optimize the RGB-based colorimetric analysis on pH papers with a wider detection range (pH ~ 0 to 6). Here, we propose a new model to establish the linear relationship between RGB values and pH: $pH_{predict} = a \times R_{normal} + b \times G_{normal} + c \times B_{normal}$. This model shows a wider applicability and higher accuracy than those in previous studies, and is thus recommended in future RGB-based colorimetric analyses on pH papers. Moreover, we identify one type of pH paper (Hydrion® Brilliant pH dip stiks, Lot Nr. 3110, Sigma-Aldrich) that is more applicable for ambient aerosols in terms of its wide pH detection range (0 to 6) and strong anti-interference capacity. The determined minimum sample mass (~ 180 µg) highlights its potential to predict aerosol pH with a high time resolution (e.g., ≤ 1 hour). We further show that the routinely adopted way of using pH color charts to predict aerosol pH may be biased by the mismatch between the standard colors on the color charts and the real colors of investigated samples. Thus, instead of using the producer-provided color chart, we suggest an in-situ calibration of pH papers with standard pH buffers.





1 Introduction

Aerosol particles have vital impacts on atmospheric chemistry, human health and global climate (Pöschl, 2005; Baltensperger et al., 2008; Pósfai and Buseck, 2010; von Schneidemesser et al., 2015; Shiraiwa et al., 2017). Understanding the basic physicochemical properties of aerosols can provide insights into various aerosol processes in the atmosphere and may further help to establish measures against air pollution. Aerosol acidity, usually quantified by aerosol pH, is one of the most important basic properties of liquid-phase aerosols. Aerosol pH has multiple effects on the other properties of aerosols, e.g., aerosol composition (Cheng et al., 2016), reactivity (Gao et al., 2004; Iinuma et al., 2004; Northcross and Jang, 2007), toxicity (Fang et al., 2017; Chowdhury et al., 2018), phase transition (Dallemagne et al., 2016; Losey et al., 2018) and their related climatic effects (Dinar et al., 2008; Hinrichs et al., 2016; Cai et al., 2018). It also plays a critical role during secondary organic aerosol (SOA) formation (e.g., Surratt et al., 2007; Gaston et al., 2014; Han et al., 2016) and in many other chemical processes in the atmosphere (Hennigan et al., 2015; Cheng et al., 2016; Wang et al., 2016; Keene et al., 2004; Ahrens et al., 2012).

Despite its essential importance, currently there is few aerosol pH measurement data set available. One main reason is the small sizes (with an aerodynamic diameter range of 2 nm - 10 μm, see McNeill, 2017) of these atmospheric particles, rendering measurements of aerosol pH not as easy as for bulk solutions. Moreover, the non-conservative nature of H+, i.e., H+ concentrations do not scale in proportion to the dilution levels due to buffering effects and the partial dissociation of weak acids, further makes probe of aerosol pH a challenging topic (Hennigan et al., 2015). For direct measurements of aerosol pH, two types of methods have been employed: filterbased sample extraction (Koutrakis et al., 1988; Keene et al., 2002; Jang et al., 2008) and spectroscopic/microscopic analysis (Li and Jang, 2012; Dallemagne et al., 2016; Rindelaub et al., 2016; Craig et al., 2017; Wei et al., 2018). As the former method is offline, it suffers from both poor time resolution and intensive labor work (Hennigan et al., 2015). Moreover, it cannot account for the water in the aerosol droplets, and involves extraction with solvents that can shift the equilibria of present ions, leading to high uncertainties. The latter method is normally used for laboratory-generated particles with simple compositions that cannot fully represent ambient aerosols (Craig et al., 2018). Due to these limitations of direct measurements, thermodynamic equilibrium models such as ISORROPIA-II (Fountoukis and Nenes, 2007) and E-AIM (Clegg and Seinfeld, 2006a, b) have been widely used to estimate the acidity of ambient aerosol droplets, although comprehensive evaluations of the acidity are hampered by lack of observational data. Thus, developing new methods to directly measure ambient aerosol pH is imminently needed to constrain the output of thermodynamic models.

In a recent study, Craig et al. (2018) reported an intriguing way to directly measure aerosol pH using pH-indicator papers, which in the past are the most common and convenient tool to test the pH of bulk solutions. To measure aerosol pH, the generated size-resolved aqueous aerosol samples ((NH₄)₂SO₄-H₂SO₄) were firstly collected on pH-indicator papers. Then the color of the samples on pH papers was analyzed quantitatively through a colorimetric image processing program (Matlab). In this way, the standard pH color chart of the indicator papers was used as a reference to finally derive the aerosol pH. The use of pH-indicator papers and the related color processing technique introduced by Craig et al. (2018) tactfully circumvents the challenges and difficulties in aerosol pH measurements. However, Craig et al. (2018) only reported two types of pH papers with relatively high





precision for pH measurements (one with pH range 0 - 2.5 and the other 2.5 - 4.5, see Craig et al., 2018), whereas in the atmosphere the aerosol pH may vary in a wide range. Note that the authors indeed employed another type of pH paper with a larger pH range from 0 to 6 for ambient aerosol sampling, unfortunately they found that this paper was not compatible with their Matlab script for more quantitative analysis (Craig et al., 2018).

The colorimetric method used by Craig et al. (2018) was based on analyzing the red (R), green (G) and blue (B) channels of the sample images, where a linear dependence of the difference between G and B (G-B) on pH² was found. According to trichromatic theory, RGB are the three primary colors and their combination in varying proportions can generate any other specific color (Su et al., 2008). The standard RGB scale is represented by the values of R, G and B, and each has a range from 0 to 255. For example, the number [0, 0, 0], i.e., R = 0, G = 0, B = 0, corresponds to absolute black and [255, 255, 255] to true white. RGB-based image analysis has been applied in the fields of inorganic and analytical chemistry. For instance, Selva Kumar et al. (2018) found a good linearity between concentrations of Thorium ions (Th^{4+}) and the ratio of R and G (R/G), and Wan et al. (2017) reported a relation between bovine serum albumin (BSA) concentrations and the normalized values of R, G and B, respectively. In these previous studies, different RGB models (i.e., ways to interpret the RGB values) were adopted, however with few detailed explanations on the intrinsic reasons. To further enhance the reliability and comparability of the data associated with RGB analysis, a unified model/method to deal with the RGB information is needed, especially for the pH determination of aerosols where high uncertainty of measured pH values can have a huge impact on the pH-dependent multiphase chemical processes.

Considering that the pH values of ambient aerosols can cover a wide range (up to ~ 6) (von Glasow and Sander, 2001; Pszenny et al., 2004; Song et al., 2018; Shi et al., 2019), the goal of the present study is to optimize the RGB-based colorimetric analysis on pH-indicator papers for direct determination of ambient aerosol pH in a wider detection range (pH ~ 0 to 6). We thus propose a new way to analyze the RGB values and establish the relationship between RGB and pH. We further compare our proposed RGB model with the models used in previous studies in terms of evaluating the established linear relationship between RGB and pH. Nine types of pH papers are tested for their potential of probing pH of ambient aerosols. In addition, the routine way of using a pH color chart to derive the pH of samples is inspected, and the results reveal some deficiencies of this method. Therefore, we suggest an optimized way to use pH papers for aerosol pH prediction with higher precision and accuracy.

2 Materials and Methods

2.1 pH-indicator papers

Nine types of pH-indicator papers were adopted in this study. Each type has a pH color chart that is accompanied with the pH papers and supposed to serve as a reference to quantify the pH of a sample through colorimetric analysis. Details about the pH paper detection ranges and the corresponded type classification used in this work can be found in Table S1. The first two types are the same as used by Craig et al. (2018), aiming to compare our results with those from Craig et al. and validate our colorimetric image processing method. The others have larger pH detection ranges covering the generally observed pH range of ambient aerosols. Note that in this study, we mainly focused on the first five types of pH papers and the remaining four types were also evaluated and compared



109

110

111

112

113

114

115

116

117

118

119

120

121

122

123

124

125

126

127

128

129

130

131 132

133

134

135

136

137

138

139

140

141

142

143

144



with the first five types in terms of their resistance to chemical interference and potential capability to measure the pH of ambient aerosols.

2.2 pH buffers and aerosol sample solutions

To examine the correlation between RGB and pH, eight standard pH buffer solutions were used as purchased and meanwhile several other buffers (with different pH values as the purchased ones) were obtained by mixing the commercial buffers with solutions of sodium hydroxide or hydrochloric acid (prepared using de-ionized water, $18.2~M\Omega$ cm). pH values of all the buffers were further checked by a pH bench meter (model: HI 2020-02, Hanna Instruments Inc., USA). Prior to the check, the pH meter was calibrated with a three-point calibration mode using the standard buffer solutions provided by Hanna Instruments Inc., USA. The measured pH values and their standard derivations are listed in Table S2, and the measured pH values show a small deviation from those specified on the buffer solution bottles, within the displayed uncertainties concomitant with these specified values. Considering that some inorganic/organic components of ambient aerosols might interfere with the dyes on pH papers and cause biased estimation of pH, salt systems with varying inorganic and/or organic acids common in aerosols and pH levels (as measured by the pH bench meter) were employed to test the applicability of different types of pH papers combined with our RGB model. Details about the composition of the tested salt systems can be found in Table S3. In general, the inorganic systems were similar to those used by Craig et al. (2018). Here, we further tested the influence of organic acids on pH paper performance by adding organic acids into the inorganic systems. As oxalic acid (C₂H₂O₄) and malonic acid (C₃H₄O₄) were frequently detected in tropospheric aerosols and found to be the dominant short dicarboxylic acids in aerosol composition (Abbatt et al., 2005; Falkovich et al., 2005), they were adopted in this study. For the solution preparation of each system, varying amounts of 1 M inorganic/organic acids were added into 30 mM inorganic salt solution to achieve different pH levels (Surratt et al., 2008; Craig et al., 2018). To prepare the inorganic and organic mixtures, the amount of added organic acids was generally two times larger than the inorganic acids and the final salt concentration could be as low as 15 mM due to the dilution effect of added acids. To prepare the solutions, all chemicals were used as purchased: NaOH (\geq 99.0%, Roth, Germany), Na₂SO₄ (\geq 99.0%, Merck, Germany), NaNO₃ (\geq 99.0%, Merck, Germany), Na₂CO₃ (≥ 99.5%, Sigma-Aldrich, USA), (NH₄)₂SO₄ (≥ 99%, Sigma-Aldrich, USA), NH₄NO₃ (≥ 98.0%, Fisher Chemical, USA), MgSO₄ (> 98%, neoFroxx GmbH, Germany), H₂SO₄ (98%, Merck, Germany), HNO₃ (65%, Merck, Germany), HCl (37%, Merck, Germany), C₂H₂O₄·2H₂O (≥ 99%, Sigma-Aldrich, USA) and C₃H₄O₄ (99%, Sigma-Aldrich, USA).

2.3 Correlation between RGB and pH

Figure 1 shows the procedure of how to use a colorimetric analysis to obtain the correlation between RGB and pH. First, $2 \mu L$ of liquid samples was dripped onto each piece of pH paper held by a clean transparent glass plate (with the other side coated by a piece of graph paper). This adopted small volume ($2 \mu L$) was based on calculation of the available amounts of liquid aerosols for aerosol sampling under a typically polluted conditions (with PM_{2.5} mass concentration around $100 \mu g \text{ m}^{-3}$) with high relative humidity ($\geq 80\%$), and assuming a sampling flow rate of several hundred liter per minute (e.g., can be achieved by a Tisch Environmental PM_{2.5} high volume air sampler, see https://tisch-env.com/high-volume-air-sampler/pm2.5) and a sampling time of a few (2 - 4) hours. Note that, to further reduce the sampling flow rate and time, we identified the minimum sample volume and mass needed to





generate a measurable color change on the suggested pH paper. The related results are shown below. Then an image of the sample was captured by a smartphone camera (Apple iPhone 5s in this study) immediately. Similar to Craig et al. (2018), the corresponding color chart of the used pH paper was included into each image to correct for potential influences of variations of light source and angle during photographing. The digital images were processed by an Adobe Photoshop software to crop a square with 100 × 100 pixels at the center of the sample (as well as each color chip on the color chart). The RGB information of the cropped square was then obtained and further analyzed by Matlab (The MathWorks, Inc. version R2018b).

2.4 RGB model

Considering that a color is represented by combination of R, G and B values, a linear combination of these three primary colors should be able to reflect the characteristics of the color and therefore represent the pH related to the color. Su et al. (2008) reported a good correlation between the linearly combined RGB and the contents of chlorophyll a and lipid, respectively in microalgae. To further account for the effect of changing light intensity on the obtained RGB values, each color channel should be normalized at first (Yadav et al., 2010). The normalization can be achieved through Eqns. (1) – (3) shown below:

160
$$R_{\text{normal}} = R/(R+G+B)$$
 (1)

161
$$G_{\text{normal}} = G/(R + G + B)$$
 (2)

162
$$B_{\text{normal}} = B/(R + G + B)$$
 (3)

- where *R*, *G* and *B* are the mean value of each primary color on the entire 100 × 100 pixels image, respectively.

 Note that every pixel has an RGB value vector: [R, G, B]. Then a model describing the linear combination of
- 166 RGB can be given as follows:

$$pH_{\text{predict}} = aR_{normal} + bG_{normal} + cB_{normal}$$
(4)

- where the linear combination $aR_{normal} + bG_{normal} + cB_{normal}$ essentially represents the color information and here can be treated as equivalent to the predicted pH ($pH_{predict}$) based on RGB analysis; a, b and c are the coefficients, which can be determined by linear regression analysis through Matlab. The linear regression function is expressed
- which can be determined by linear regression analysis through Matlab. The linear regression function is expressed
- 173 as

$$174 Y = aX_1 + bX_2 + cX_3 (5)$$

where Y is the dependent variable vector, X_1 , X_2 and X_3 are independent variable vectors. These vectors can be achieved from a standard color chart or a series of buffer samples (with known pH values) on pH papers: Y is the series of pH values (i.e., reference pH, $pH_{reference}$) shown on the color chart or of buffer samples (as shown in Fig. 1, the pH papers with different pH buffer solutions are collected together to form a pH series); X_1 , X_2 and X_3 are the normalized average of R, G and B respectively, based on analysis on the detected colors. As a color chart is normally used as a reference for pH measurements using pH papers, a linear regression analysis on the color chart can provide the coefficient vector [a, b, c] as an answer. Then the same set of coefficient vector (i.e., [a, b, c]) are





used to predict the pH (i.e., $pH_{predict}$) of aerosol samples using Eqn (4). Thus, with this RGB model, a linear relationship between RGB-predicted pH ($pH_{predict}$) and reference pH ($pH_{reference}$) is expected for the calibration (as shown in Fig. 1), in order to finally predict sample aerosol pH on a pH paper.

3 Results

3.1 Validation of the new RGB model

As the RGB model (i.e., G-B vs pH²) used by Craig et al. (2018) produced good linear correlations for the two types of pH papers that the authors adopted, we first examined the validity of this RGB model against the first five types of pH papers used in this work. Note that here the first two types of pH papers are the ones used and recommended by Craig et al. (2018). Figure S1 shows the relationship between average G-B and pH² derived from the color charts of these five types of pH papers, respectively. Relatively good linear correlations can be found for the first three types, which is consistent with Craig et al. (2018). However, non-monotonic correlations are encountered for the last two types of pH papers, which are the ones with wider pH detection ranges (0.5 - 5.5 and 0 - 6, respectively). These results indicate a limited feasibility of the RGB model proposed by Craig et al. (2018).

Thus, the validity of our new RGB model was further checked through the five types of pH papers. The colors on the color chart for each type of pH paper were firstly analyzed through our RGB model and then the calculated $pH_{predict}$ were compared with the reference pH shown on the color chart. As shown in Fig. 2a-e (the 'color chart' column on the left-hand side), good linearity between $pH_{predict}$ and $pH_{reference}$ can be observed for all these pH paper types.

As aforementioned, besides the RGB model used by Craig et al. (2018), other models have also been adopted to create a linear correlation between RGB and concentrations of the chemicals of interest in previous colorimetric analyses (Su et al., 2008; Yadav et al., 2010; Wan et al., 2017; Selva Kumar et al., 2018). However, few comparisons have been made regarding the goodness of the established linearity using these RGB models. Here we further compared our model with the other two models (i.e., R/G vs pH and G-B vs pH²) proposed by Selva Kumar et al. (2018) and Craig et al. (2018) respectively, in terms of evaluating their correlation coefficient, R^2 . Figure S2 (the 'color chart' panel on the left-hand side) displays the R^2 of the established linear correlation between $pH_{predict}$ and $pH_{reference}$ when the three models are used for the five types of pH papers, respectively. For the color-chart-derived linear correlation, the model G-B vs pH² presents poor goodness-of-fit for type IV and V pH papers (consistent with the results shown in Fig. S1). The model R/G vs pH shows relatively high R^2 for all the pH paper types. Nevertheless, this RGB model still underperforms our model. Overall, our RGB model could provide a high R^2 (> 0.95) for all the five types of pH papers, which demonstrates the universal validity of our RGB model.

3.2 Calibration with standard buffer solutions

A good linearity, however, may not always be obtained from the color chart of some types of pH papers in some pH ranges. For example, in the 'color chart' column of Fig. 2, the pH_{predict} present small but discernable deviations from pH_{reference} for types I, III and IV pH papers. And the type V pH paper shows even larger differences at both





ends of the pH range. Similar phenomenon was also observed in the study of Craig et al. (2018) with their RGB model, where they argued that the pH paper dye became less effective at the limits of the pH paper range, due to the pK_a values of the dye were normally at the middle of the pH range. But it may also originate from some color bias due to the differences between the experiment conditions and the ones under which the color chart is made by the producer.

Thus, following the same procedure as for the color chart (see Fig. 1), pH papers with samples of a series of $2 \mu L$ standard buffer droplets were also measured. The pH values of the standard buffers were known beforehand and further checked with a pH meter (also denoted as ' $pH_{reference}$ ', see Table S2). Figures 2f-j (the ' $2 \mu L$ buffer' column on the right-hand side) show the comparison between $pH_{predict}$ and $pH_{reference}$ for the samples of $2 \mu L$ buffers. Much better linearity between $pH_{predict}$ and $pH_{reference}$ can be observed for all the five types of pH papers. Especially, the significant deviation of $pH_{predict}$ from $pH_{reference}$ found in the left panel (the 'color chart' column) disappear for the type I and V pH papers. This means that the deviations at the edge of the pH range in the color-chart-derived calibration curves are mainly due to the color bias of the color chart itself or caused during photographing.

Actually, even small deviations found in the color-chart-derived calibration curves (the 'color chart' column in Fig. 2) may lead to significant or non-negligible errors in measuring aerosol pH. We conducted a case study using the type IV pH paper combined with our RGB model to predict the pH of buffer samples by using the color-chart-derived coefficient vector [a, b, c], i.e., the color-chart-calibration method (Fig. 2d). The blue symbols in Fig. S3 represent pH_{predict} versus $pH_{\text{reference}}$ of the standard buffer samples. Systematical underestimation of pH_{predict} can be found at the lower $pH_{\text{reference}}$ values (i.e., $pH_{\text{reference}} = 1$, 1.68 and 2) but a slight overestimation is observed at $pH_{\text{reference}} = 5$. This significant discrepancy may be attributed to the mismatch between the reference colors on the color chart and the real colors of the samples, due to the differences between our experiment conditions and the ones under which the color chart is made by the producer. This gives us a hint that the coefficient vector derived from the color chart is not suitable for predicting the pH of aerosol samples.

For the established linear relationship using 2 μ L standard buffers, the performances of different RGB models were further compared and the results are shown in Fig. S2 (the '2 μ L buffer' panel on the right-hand side). Our RGB model still outperforms the other two models for all the five types of pH papers employed in this work. Overall, the good agreement between $pH_{predict}$ and $pH_{reference}$ for all these tested pH papers verifies the wide applicability of our RGB model to the pH paper calibration using standard buffers. In the following section, we will examine the quality of predicting samples pH with the standard-buffer-calibration method.

3.3 pH estimation for aerosol surrogates ((NH₄)₂SO₄-H₂SO₄) with the type IV and V pH papers

In order to test the feasibility of pH papers with larger pH detection ranges for pH prediction of aerosols, we further used the type IV and V pH papers to estimate the pH of lab-prepared aerosol surrogates ($(NH_4)_2SO_4$ - H_2SO_4). To minimize the effect of varying photographing conditions (e.g., angle, light variation) on the colors of the captured image, experiments were carried out in a cupboard with a constant light source. In addition, the pH paper samples as well as the smartphone were fixed on a small glass plate and a metal holder respectively, to keep their position unchanged throughout the experiment. Note that applying the standard-buffer-derived coefficients





(Fig. 2i and 2j) for pH prediction of samples required the same constant light source conditions for sample imagingprocesses as for standard buffers.

 pH_{predict} versus $pH_{\text{reference}}$ for the samples on the type IV pH paper are shown in Fig. S4. Generally, the pH_{predict} by the type IV pH paper are comparable with the $pH_{\text{reference}}$ at a lower pH range (i.e. $pH_{\text{reference}} = 0.46$, 1.52 and 3.0). However, an obvious outlier with 1.5 unit of overestimation in pH_{predict} can be found at $pH_{\text{reference}}$ around 4 (outlier highlighted by the arrow in Fig. S4). This overestimation was proved to be reproducible by our six replicate experiments and it was even found for samples of diluted H_2SO_4 solutions with reference pH around 4 on the type IV pH paper. Such overestimation may be due to the chemical interferences caused by the samples or the low buffering levels of the samples. Thus, the type IV pH paper is not recommended for future pH measurements of aerosols. However, it may still work well for the other sample types, such as found for our self-prepared phosphate buffers (Fig. S4). On the other hand, the type V pH paper shows decent agreements between pH_{predict} and $pH_{\text{reference}}$ within the examined pH range, as shown in Fig. 3a. Moreover, the pH_{predict} are also compared with the results by Craig et al. (2018). The orange and blue bars in Fig. 3a represent the measured pH ranges for the lab-generated (NH₄)₂SO₄ - H₂SO₄ aerosols with particle sizes larger than 2.5 μm using pH papers (the same as the type I and II

275 4 Discussion

4.1 Chemical interference

pH papers used here) and Raman spectroscopy, respectively.

As aforementioned, aerosol samples with different compositions may have interferences on the indicating color of a pH paper and thereby cause its poor performance for aerosol pH prediction, e.g., the overestimation of pH of aerosol surrogates ((NH₄)₂SO₄-H₂SO₄) with the Type IV pH paper. To test the capability of chemical resistance of the Type V pH paper, we further tested its performance of predicting the pH of lab-prepared aerosol surrogates with varying inorganic/organic compositions commonly exist in ambient aerosols.

Figure 4 displays $pH_{predict}$ versus $pH_{reference}$ for our lab-prepared samples under different pH levels using the Type V pH paper. As shown in Fig. 4a, systematic divergences between $pH_{predict}$ and $pH_{reference}$ (i.e., overestimation of $pH_{predict}$ when $pH_{reference}$ is in the range of 2.5 – 3.5 whereas underestimation of $pH_{predict}$ when $pH_{reference} > \sim 4.5$) can be found for these tested inorganic systems. Interestingly these mismatches disappear when the organic acids are introduced into these inorganic systems (Fig. 4b), and also for the cases when the inorganic acids are replaced by organic acids (Fig. S5). In Fig.4b, this good agreement for $pH_{predict}$ versus $pH_{reference}$ is observed not only for systems containing oxalic acid ($C_2H_2O_4$, solid markers) but also for those having malonic acid ($C_3H_4O_4$, hollow markers) with an average deviation (of $pH_{predict}$ from $pH_{reference}$) < 0.5 unit. The fact that the existence of organic acids significantly improves the quality of $pH_{predict}$ may be attributed to the enhanced buffering levels of the samples (Fillion et al., 1999; Li et al., 2016). Actually, good agreement between $pH_{predict}$ and $pH_{reference}$ is found for both the inorganic and organic phosphate systems (Fig. S6) based on our further tests, which is probably due to the high buffering levels of these systems maintained by the phosphate itself (Hourant, 2004). Nevertheless, the solvent effect of the added organics on acid dissociation equilibria may also play a role (Padró et al., 2012). The detailed mechanisms may need to be explored in future studies. Given the large contribution of organics





(Jimenez et al., 2009) and the well-known dominance of both organic acids (i.e., oxalic acid and malonic acid) in ambient aerosols (Abbatt et al., 2005; Falkovich et al., 2005), the potential interferences found for the inorganic systems can be expected to become vanished when organics are concomitant under ambient conditions. Additionally, the interference check was also performed for the other pH paper types (type III and VI-IX) that have larger pH detection ranges. Similar to the type IV pH paper, significant deviations of $pH_{predict}$ from $pH_{reference}$ (≥ 1.5 unit) were observed for these types (see SI text and Fig. S7).

4.2 Identification of the needed minimum sample amount for the type V pH paper

The pH of ambient aerosols can be changing due to the varying atmospheric composition (e.g., some important trace gases like SO2, NO2, NH3 and organic acids) and physical characteristics (e.g., ambient relative humidity and temperature). Thus, reflecting the temporal evolution of aerosol pH requires a pH measurement method with a high time resolution. As aforementioned, to collect $2 \mu L$ of liquid aerosol samples, a sampling time of 2-4hours is needed even using a high-volume air sampler with a sampling flow rate of several hundred liter per minute. Here, in order to have a higher time resolution and/or a lower sampling flow rate, we further identified the minimum sample volume needed to generate a measurable color change on the type V pH paper. Figure 3b shows the results for 0.1µL of lab-prepared aerosol samples. Similar to the RGB analysis procedure used for the 2 µL samples (e.g., in Fig. 3a), the pH_{predict} in Fig. 3b are calculated with the averaged coefficient vector [a, b, c] derived from three replicate calibration experiments with 0.1 μ L standard buffers (Fig. S8). Generally, $pH_{predict}$ agrees well with $pH_{\text{reference}}$, with biases (averaged pH_{predict} versus $pH_{\text{reference}}$) within 0.5 unit. Note that to avoid fast water exchange between the lab air and our samples (aerosols/buffers) as well as potential interfering effects (absorption/reaction) caused by the lab air, the 0.1 µL samples were transferred (through a pipette) directly onto the pH paper surface and each sample was immediately photographed (≤ ~ 3 seconds) after it got contact with the pH paper dye. Due to this extremely small sample volume, the influence of lab air on sample pH could become prominent because a significant sample color change was frequently observed after the sample was exposed to the lab air for $> \sim 5$ seconds.

These results confirm the feasibility of the type V pH paper as well as our RGB model for pH estimation of the liquid aerosol samples with a volume even down to $0.1~\mu$ L. This tiny volume corresponds to a sample mass of ~ 180 µg assuming an effective density of $1.8~g~cm^{-3}$ for ambient aerosols (Sarangi et al., 2016; Geller et al., 2006), which is comparably low to the needed minimum particulate masses in Craig et al. (2018), i.e., ~ 65 µg to ~ 2.5 mg for PM_{2.5} or larger particles with pH from 0-2.5 to 2.5-4. According to our calculation, under typically polluted conditions (with PM_{2.5} mass concentration around $100~\mu$ g m⁻³) with high relative humidity ($\geq 80\%$) to collect aerosol samples with this even smaller mass/volume can be achieved by an air sampler with a sampling flow rate of ~ 22 liter per minute and with a sampling time of ~ 1 hour. Alternatively, the sample time can be further reduced to less than 1 hour (e.g., 0.5 hour) by adopting an air sampler with a relatively higher sampling flow rate (e.g., ~ 45 liter per minute). The needed low sampling mass and the large pH detection range of the type V pH paper highlight its potential for future development of real-time aerosol pH measurements. Moreover, instead of using a color chart to calibrate pH papers for each sample (Craig et al., 2018), our results demonstrate that the in-situ calibration method of using standard buffers (independent of different samples) can derive an averaged coefficient vector [a, b, c] which can be uniformly applied to pH prediction of different samples provided





the photographing conditions are kept constant. This unique feature further facilitates the application of the type

V pH paper under ambient cases.

5 Conclusions

338

339 We proposed a new model to establish the correlation between the color of samples on pH-indicator papers and 340 their measured pH. The model was based on RGB analysis of the images of samples. Comparison of our model 341 and another two RGB models verified the high reliability of our model. Using our RGB model, good agreement 342 between the model-predicted pH ($pH_{predict}$) and reference pH ($pH_{reference}$) for pH paper color charts as well as standard buffers were observed for all the tested types of pH papers. Different types of pH papers with larger pH 343 344 detection ranges were further examined for their performance to predict the pH of lab-prepared aerosol samples 345 with varying inorganic/organic compositions common in ambient aerosols. The results suggest that only the type 346 V pH paper (with a pH range of ~ 0 - 6) deserves practical applications for pH measurements of ambient aerosols. 347 The minimum liquid sample mass/volume needed for the type V pH paper is identified as ~ 180 µg/0.1 µL, which 348 means that this type of pH paper can be applied to collect ambient aerosols with a high time resolution (e.g., ≤ 1 349 hour). Whereas the other types may suffer from some chemical interferences during pH measurements and 350 therefore can generate large biases for the measured pH of aerosols. The routine procedure of using pH papers to 351 estimate a sample pH was also examined in a case study using the type IV pH paper. The results show that 352 referring to the color chart for pH estimation (i.e. the color-chart-calibration method) may cause a bias of the 353 predicted pH. To use the pH papers in a more proper way, here we further demonstrated that the in-situ calibration 354 method of using standard buffers (independent of different samples) could derive an averaged coefficient vector 355 [a, b, c], which can be uniformly applied to pH prediction of different samples provided the photographing 356 conditions are kept constant.

Data availability

357

- The underlying research data and Matlab code can be accessed upon contact with Guo Li (guo.li@mpic.de),
- Yafang Cheng (yafang.cheng@mpic.de) or Hang Su (h.su@mpic.de).

360 Supplement

The supplement is available in a separate file.

362 Author contributions

- 363 Y.C. and H.S. conceived and led the study. G.L. performed experiments and data analysis. Y.C., H.S., U.P., U.K.,
- 364 N.M., G.Z., M.L., T.K. discussed the results. G.L. and Y.C. wrote the manuscript with inputs from all co-authors.

365 Competing interests

366 The authors declare that they have no conflict of interest.





367 Acknowledgement

- 368 We acknowledge the National Natural Science Foundation of China (grant no. 91644218) and the National Key
- 369 Research and Development Program of China (grant no. 2017YFC0210104). This study was supported by the
- 370 Max Planck Society (MPG). G. L. acknowledges the financial support from the China Scholarship Council (CSC).
- 371 G. L. also would like to thank Ping Zhang, Jinqian Zhai and Yunkun Lang for their very helpful discussions
- 372 concerning the GRB model development.

373 References

- 374 Abbatt, J. P. D., Broekhuizen, K., and Pradeep Kumar, P.: Cloud condensation nucleus activity of internally
- mixed ammonium sulfate/organic acid aerosol particles, Atmospheric Environment, 39, 4767-4778,
- 376 <u>https://doi.org/10.1016/j.atmosenv.2005.04.029</u>, 2005.
- 377 Ahrens, L., Harner, T., Shoeib, M., Lane, D. A., and Murphy, J. G.: Improved Characterization of Gas-Particle
- 378 Partitioning for Per- and Polyfluoroalkyl Substances in the Atmosphere Using Annular Diffusion Denuder
- 379 Samplers, Environmental Science & Technology, 46, 7199-7206, 10.1021/es300898s, 2012.
- 380 Baltensperger, U., Dommen, J., Alfarra, M. R., Duplissy, J., Gaeggeler, K., Metzger, A., Facchini, M. C.,
- 381 Decesari, S., Finessi, E., Reinnig, C., Schott, M., Warnke, J., Hoffmann, T., Klatzer, B., Puxbaum, H., Geiser,
- 382 M., Savi, M., Lang, D., Kalberer, M., and Geiser, T.: Combined Determination of the Chemical Composition
- 383 and of Health Effects of Secondary Organic Aerosols: The POLYSOA Project, Journal of Aerosol Medicine and
- 384 Pulmonary Drug Delivery, 21, 145-154, 10.1089/jamp.2007.0655, 2008.
- 385 Cai, J., Zhi, G., Yu, Z., Nie, P., Gligorovski, S., Zhang, Y., Zhu, L., Guo, X., Li, P., He, T., He, Y., Sun, J., and
- 386 Zhang, Y.: Spectral changes induced by pH variation of aqueous extracts derived from biomass burning
- 387 aerosols: Under dark and in presence of simulated sunlight irradiation, Atmospheric Environment, 185, 1-6,
- 388 <u>https://doi.org/10.1016/j.atmosenv.2018.04.037</u>, 2018.
- 389 Cheng, Y., Zheng, G., Wei, C., Mu, Q., Zheng, B., Wang, Z., Gao, M., Zhang, Q., He, K., Carmichael, G.,
- 390 Pöschl, U., and Su, H.: Reactive nitrogen chemistry in aerosol water as a source of sulfate during haze events in
- 391 China, Science Advances, 2, e1601530, 10.1126/sciadv.1601530, 2016.
- Chowdhury, P. H., He, Q., Lasitza Male, T., Brune, W. H., Rudich, Y., and Pardo, M.: Exposure of Lung
- 393 Epithelial Cells to Photochemically Aged Secondary Organic Aerosol Shows Increased Toxic Effects,
- Environmental Science & Technology Letters, 5, 424-430, 10.1021/acs.estlett.8b00256, 2018.
- 395 Clegg, S. L., and Seinfeld, J. H.: Thermodynamic Models of Aqueous Solutions Containing Inorganic
- 396 Electrolytes and Dicarboxylic Acids at 298.15 K. 1. The Acids as Nondissociating Components, The Journal of
- 397 Physical Chemistry A, 110, 5692-5717, 10.1021/jp056149k, 2006a.
- 398 Clegg, S. L., and Seinfeld, J. H.: Thermodynamic Models of Aqueous Solutions Containing Inorganic
- 399 Electrolytes and Dicarboxylic Acids at 298.15 K. 2. Systems Including Dissociation Equilibria, The Journal of
- 400 Physical Chemistry A, 110, 5718-5734, 10.1021/jp056150j, 2006b.
- 401 Craig, R. L., Nandy, L., Axson, J. L., Dutcher, C. S., and Ault, A. P.: Spectroscopic Determination of Aerosol
- 402 pH from Acid-Base Equilibria in Inorganic, Organic, and Mixed Systems, The Journal of Physical Chemistry
- 403 A, 121, 5690-5699, 10.1021/acs.jpca.7b05261, 2017.
- 404 Craig, R. L., Peterson, P. K., Nandy, L., Lei, Z., Hossain, M. A., Camarena, S., Dodson, R. A., Cook, R. D.,
- 405 Dutcher, C. S., and Ault, A. P.: Direct Determination of Aerosol pH: Size-Resolved Measurements of
- 406 Submicrometer and Supermicrometer Aqueous Particles, Analytical Chemistry, 90, 11232-11239,
- 407 10.1021/acs.analchem.8b00586, 2018.





- 408 Dallemagne, M. A., Huang, X. Y., and Eddingsaas, N. C.: Variation in pH of Model Secondary Organic Aerosol
- during Liquid-Liquid Phase Separation, The Journal of Physical Chemistry A, 120, 2868-2876,
- 410 10.1021/acs.jpca.6b00275, 2016.
- 411 Dinar, E., Anttila, T., and Rudich, Y.: CCN Activity and Hygroscopic Growth of Organic Aerosols Following
- 412 Reactive Uptake of Ammonia, Environmental Science & Technology, 42, 793-799, 10.1021/es071874p, 2008.
- 413 Falkovich, A. H., Graber, E. R., Schkolnik, G., Rudich, Y., Maenhaut, W., and Artaxo, P.: Low molecular
- 414 weight organic acids in aerosol particles from Rondônia, Brazil, during the biomass-burning, transition
- and wet periods, Atmos. Chem. Phys., 5, 781-797, 10.5194/acp-5-781-2005, 2005.
- 416 Fang, T., Guo, H., Zeng, L., Verma, V., Nenes, A., and Weber, R. J.: Highly Acidic Ambient Particles, Soluble
- 417 Metals, and Oxidative Potential: A Link between Sulfate and Aerosol Toxicity, Environmental Science &
- 418 Technology, 51, 2611-2620, 10.1021/acs.est.6b06151, 2017.
- 419 Fillion, N., Probst, A., and Probst, J.-L.: Dissolved organic matter contribution to rain water, throughfall and soil
- 420 solution chemistry, Analusis, 27, 409-413, 1999.
- 421 Fountoukis, C., and Nenes, A.: ISORROPIA II: a computationally efficient thermodynamic equilibrium model
- 422 fc
- 422 10r 423 K⁺–Ca²⁺–Mg²⁺–NH₄<sup>+</sup
- 424 >–Na⁺–SO₄^{2−}–NO₃<sup>&
- 425 minus;</sup>−</sup>−</sup>&mash;H₂O aerosols, Atmos. Chem. Phys., 7,
- 426 4639-4659, 10.5194/acp-7-4639-2007, 2007.
- 427 Gao, S., Ng, N. L., Keywood, M., Varutbangkul, V., Bahreini, R., Nenes, A., He, J., Yoo, K. Y., Beauchamp, J.
- 428 L., Hodyss, R. P., Flagan, R. C., and Seinfeld, J. H.: Particle Phase Acidity and Oligomer Formation in
- 429 Secondary Organic Aerosol, Environmental Science & Technology, 38, 6582-6589, 10.1021/es049125k, 2004.
- 430 Gaston, C. J., Riedel, T. P., Zhang, Z., Gold, A., Surratt, J. D., and Thornton, J. A.: Reactive Uptake of an
- 431 Isoprene-Derived Epoxydiol to Submicron Aerosol Particles, Environmental Science & Technology, 48, 11178-
- 432 11186, 10.1021/es5034266, 2014.
- 433 Geller, M., Biswas, S., and Sioutas, C.: Determination of Particle Effective Density in Urban Environments with
- 434 a Differential Mobility Analyzer and Aerosol Particle Mass Analyzer, Aerosol Science and Technology, 40,
- 435 709-723, 10.1080/02786820600803925, 2006.
- 436 Han, Y., Stroud, C. A., Liggio, J., and Li, S. M.: The effect of particle acidity on secondary organic aerosol
- 437 formation from α-pinene photooxidation under atmospherically relevant conditions, Atmos. Chem. Phys., 16,
- 438 13929-13944, 10.5194/acp-16-13929-2016, 2016.
- 439 Hennigan, C. J., Izumi, J., Sullivan, A. P., Weber, R. J., and Nenes, A.: A critical evaluation of proxy methods
- 440 used to estimate the acidity of atmospheric particles, Atmos. Chem. Phys., 15, 2775-2790, 10.5194/acp-15-
- 441 2775-2015, 2015.
- 442 Hinrichs, R. Z., Buczek, P., and Trivedi, J. J.: Solar Absorption by Aerosol-Bound Nitrophenols Compared to
- 443 Aqueous and Gaseous Nitrophenols, Environmental Science & Technology, 50, 5661-5667,
- 444 10.1021/acs.est.6b00302, 2016.
- 445 Hourant, P.: General Properties of the Alkaline Phosphates: Major Food and Technical Applications,
- 446 Phosphorus Research Bulletin, 15, 85-94, 10.3363/prb1992.15.0_85, 2004.
- 447 Iinuma, Y., Böge, O., Gnauk, T., and Herrmann, H.: Aerosol-chamber study of the α-pinene/O3 reaction:
- influence of particle acidity on aerosol yields and products, Atmospheric Environment, 38, 761-773,
- 449 https://doi.org/10.1016/j.atmosenv.2003.10.015, 2004.
- 450 Jang, M., Cao, G., and Paul, J.: Colorimetric Particle Acidity Analysis of Secondary Organic Aerosol Coating
- 451 on Submicron Acidic Aerosols, Aerosol Science and Technology, 42, 409-420, 10.1080/02786820802154861,
- 452 2008

© Author(s) 2019. CC BY 4.0 License.





- 453 Jimenez, J. L., Canagaratna, M. R., Donahue, N. M., Prevot, A. S. H., Zhang, Q., Kroll, J. H., DeCarlo, P. F.,
- 454 Allan, J. D., Coe, H., Ng, N. L., Aiken, A. C., Docherty, K. S., Ulbrich, I. M., Grieshop, A. P., Robinson, A. L.,
- Duplissy, J., Smith, J. D., Wilson, K. R., Lanz, V. A., Hueglin, C., Sun, Y. L., Tian, J., Laaksonen, A.,
- 456 Raatikainen, T., Rautiainen, J., Vaattovaara, P., Ehn, M., Kulmala, M., Tomlinson, J. M., Collins, D. R.,
- 457 Cubison, M. J., Dunlea, J., Huffman, J. A., Onasch, T. B., Alfarra, M. R., Williams, P. I., Bower, K., Kondo, Y.,
- 458 Schneider, J., Drewnick, F., Borrmann, S., Weimer, S., Demerjian, K., Salcedo, D., Cottrell, L., Griffin, R.,
- Takami, A., Miyoshi, T., Hatakeyama, S., Shimono, A., Sun, J. Y., Zhang, Y. M., Dzepina, K., Kimmel, J. R.,
- 460 Sueper, D., Jayne, J. T., Herndon, S. C., Trimborn, A. M., Williams, L. R., Wood, E. C., Middlebrook, A. M.,
- 461 Kolb, C. E., Baltensperger, U., and Worsnop, D. R.: Evolution of Organic Aerosols in the Atmosphere, Science,
- 462 326, 1525-1529, 10.1126/science.1180353, 2009.
- 463 Keene, W. C., Pszenny, A. A. P., Maben, J. R., and Sander, R.: Variation of marine aerosol acidity with particle
- 464 size, Geophysical Research Letters, 29, 5-1-5-4, 10.1029/2001gl013881, 2002.
- 465 Keene, W. C., Pszenny, A. A. P., Maben, J. R., Stevenson, E., and Wall, A.: Closure evaluation of size-resolved
- 466 aerosol pH in the New England coastal atmosphere during summer, Journal of Geophysical Research:
- 467 Atmospheres, 109, 10.1029/2004jd004801, 2004.
- 468 Koutrakis, P., Wolfson, J. M., and Spengler, J. D.: An improved method for measuring aerosol strong acidity:
- 469 Results from a nine-month study in St Louis, Missouri and Kingston, Tennessee, Atmospheric Environment
- 470 (1967), 22, 157-162, https://doi.org/10.1016/0004-6981(88)90308-3, 1988.
- 471 Li, H., Liu, F., Kang, L., and Zheng, M.: Study on the buffering capacity of wort, Journal of the Institute of
- 472 Brewing, 122, 138-142, 10.1002/jib.286, 2016.
- 473 Li, J., and Jang, M.: Aerosol Acidity Measurement Using Colorimetry Coupled With a Reflectance UV-Visible
- 474 Spectrometer, Aerosol Science and Technology, 46, 833-842, 10.1080/02786826.2012.669873, 2012.
- 475 Losey, D. J., Ott, E.-J. E., and Freedman, M. A.: Effects of High Acidity on Phase Transitions of an Organic
- 476 Aerosol, The Journal of Physical Chemistry A, 122, 3819-3828, 10.1021/acs.jpca.8b00399, 2018.
- 477 McNeill, V. F.: Atmospheric Aerosols: Clouds, Chemistry, and Climate, Annual Review of Chemical and
- 478 Biomolecular Engineering, 8, 427-444, 10.1146/annurev-chembioeng-060816-101538, 2017.
- 479 Northcross, A. L., and Jang, M.: Heterogeneous SOA yield from ozonolysis of monoterpenes in the presence of
- 480 inorganic acid, Atmospheric Environment, 41, 1483-1493, https://doi.org/10.1016/j.atmosenv.2006.10.009,
- 481 2007.
- 482 Padró, J. M., Acquaviva, A., Tascon, M., Gagliardi, L. G., and Castells, C. B.: Effect of temperature and solvent
- 483 composition on acid dissociation equilibria, I: Sequenced pssKa determination of compounds commonly used as
- 484 buffers in high performance liquid chromatography coupled to mass spectroscopy detection, Analytica Chimica
- 485 Acta, 725, 87-94, https://doi.org/10.1016/j.aca.2012.03.015, 2012.
- 486 Pöschl, U.: Atmospheric Aerosols: Composition, Transformation, Climate and Health Effects, Angewandte
- 487 Chemie International Edition, 44, 7520-7540, 10.1002/anie.200501122, 2005.
- 488 Pósfai, M., and Buseck, P. R.: Nature and Climate Effects of Individual Tropospheric Aerosol Particles, Annual
- 489 Review of Earth and Planetary Sciences, 38, 17-43, 10.1146/annurev.earth.031208.100032, 2010.
- 490 Pszenny, A. A. P., Moldanová, J., Keene, W. C., Sander, R., Maben, J. R., Martinez, M., Crutzen, P. J., Perner,
- 491 D., and Prinn, R. G.: Halogen cycling and aerosol pH in the Hawaiian marine boundary layer, Atmos. Chem.
- 492 Phys., 4, 147-168, 10.5194/acp-4-147-2004, 2004.
- 493 Rindelaub, J. D., Craig, R. L., Nandy, L., Bondy, A. L., Dutcher, C. S., Shepson, P. B., and Ault, A. P.: Direct
- 494 Measurement of pH in Individual Particles via Raman Microspectroscopy and Variation in Acidity with
- 495 Relative Humidity, The Journal of Physical Chemistry A, 120, 911-917, 10.1021/acs.jpca.5b12699, 2016.
- 496 Sarangi, B., Aggarwal, S. G., Sinha, D., and Gupta, P. K.: Aerosol effective density measurement using
- 497 scanning mobility particle sizer and quartz crystal microbalance with the estimation of involved uncertainty,
- 498 Atmos. Meas. Tech., 9, 859-875, 10.5194/amt-9-859-2016, 2016.





- 499 Selva Kumar, R., Kumar, S. K. A., Vijayakrishna, K., Sivaramakrishna, A., Brahmmananda Rao, C. V. S.,
- 500 Sivaraman, N., and Sahoo, S. K.: Development of the Smartphone-Assisted Colorimetric Detection of Thorium
- 501 by Using New Schiff's Base and Its Applications to Real Time Samples, Inorganic Chemistry, 57, 15270-
- 502 15279, 10.1021/acs.inorgchem.8b02564, 2018.
- 503 Shi, G., Xu, J., Shi, X., Liu, B., Bi, X., Xiao, Z., Chen, K., Wen, J., Dong, S., Tian, Y., Feng, Y., Yu, H., Song,
- 504 S., Zhao, Q., Gao, J., and Russell, A. G.: Aerosol pH dynamics during intense haze periods in an urban
- 505 environment in China: use of detailed, hourly, speciated observations to study the role of ammonia availability
- and secondary aerosol formation and urban environment, Journal of Geophysical Research: Atmospheres, 0,
- 507 10.1029/2018jd029976, 2019.
- 508 Shiraiwa, M., Ueda, K., Pozzer, A., Lammel, G., Kampf, C. J., Fushimi, A., Enami, S., Arangio, A. M.,
- 509 Fröhlich-Nowoisky, J., Fujitani, Y., Furuyama, A., Lakey, P. S. J., Lelieveld, J., Lucas, K., Morino, Y., Pöschl,
- 510 U., Takahama, S., Takami, A., Tong, H., Weber, B., Yoshino, A., and Sato, K.: Aerosol Health Effects from
- 511 Molecular to Global Scales, Environmental Science & Technology, 51, 13545-13567, 10.1021/acs.est.7b04417,
- **512** 2017.
- 513 Song, S., Gao, M., Xu, W., Shao, J., Shi, G., Wang, S., Wang, Y., Sun, Y., and McElroy, M. B.: Fine-particle
- 514 pH for Beijing winter haze as inferred from different thermodynamic equilibrium models, Atmos. Chem. Phys.,
- 515 18, 7423-7438, 10.5194/acp-18-7423-2018, 2018.
- 516 Su, C.-H., Fu, C.-C., Chang, Y.-C., Nair, G. R., Ye, J.-L., Chu, I.-M., and Wu, W.-T.: Simultaneous estimation
- 517 of chlorophyll a and lipid contents in microalgae by three-color analysis, Biotechnology and Bioengineering, 99,
- 518 1034-1039, 10.1002/bit.21623, 2008.
- 519 Surratt, J. D., Lewandowski, M., Offenberg, J. H., Jaoui, M., Kleindienst, T. E., Edney, E. O., and Seinfeld, J.
- 520 H.: Effect of Acidity on Secondary Organic Aerosol Formation from Isoprene, Environmental Science &
- 521 Technology, 41, 5363-5369, 10.1021/es0704176, 2007.
- 522 Surratt, J. D., Gómez-González, Y., Chan, A. W. H., Vermeylen, R., Shahgholi, M., Kleindienst, T. E., Edney,
- 523 E. O., Offenberg, J. H., Lewandowski, M., Jaoui, M., Maenhaut, W., Claeys, M., Flagan, R. C., and Seinfeld, J.
- 524 H.: Organosulfate Formation in Biogenic Secondary Organic Aerosol, The Journal of Physical Chemistry A,
- 525 112, 8345-8378, 10.1021/jp802310p, 2008.
- 526 von Glasow, R., and Sander, R.: Variation of sea salt aerosol pH with relative humidity, Geophysical Research
- 527 Letters, 28, 247-250, 10.1029/2000GL012387, 2001.
- 528 von Schneidemesser, E., Monks, P. S., Allan, J. D., Bruhwiler, L., Forster, P., Fowler, D., Lauer, A., Morgan,
- 529 W. T., Paasonen, P., Righi, M., Sindelarova, K., and Sutton, M. A.: Chemistry and the Linkages between Air
- 530 Quality and Climate Change, Chemical Reviews, 115, 3856-3897, 10.1021/acs.chemrev.5b00089, 2015.
- Wan, Z., Zhong, L., Pan, Y., Li, H., Zou, Q., Su, K., and Wang, P.: Portable Microplate Analyzer with a
- 532 Thermostatic Chamber Based on a Smartphone for On-site Rapid Detection, Analytical Sciences, 33, 1291-
- 533 1296, 10.2116/analsci.33.1291, 2017.
- Wang, G., Zhang, R., Gomez, M. E., Yang, L., Levy Zamora, M., Hu, M., Lin, Y., Peng, J., Guo, S., Meng, J.,
- 535 Li, J., Cheng, C., Hu, T., Ren, Y., Wang, Y., Gao, J., Cao, J., An, Z., Zhou, W., Li, G., Wang, J., Tian, P.,
- Marrero-Ortiz, W., Secrest, J., Du, Z., Zheng, J., Shang, D., Zeng, L., Shao, M., Wang, W., Huang, Y., Wang,
- 537 Y., Zhu, Y., Li, Y., Hu, J., Pan, B., Cai, L., Cheng, Y., Ji, Y., Zhang, F., Rosenfeld, D., Liss, P. S., Duce, R. A.,
- Kolb, C. E., and Molina, M. J.: Persistent sulfate formation from London Fog to Chinese haze, Proceedings ofthe National Academy of Sciences, 113, 13630-13635, 10.1073/pnas.1616540113, 2016.
- Wei, H., Vejerano, E. P., Leng, W., Huang, Q., Willner, M. R., Marr, L. C., and Vikesland, P. J.: Aerosol
- 541 microdroplets exhibit a stable pH gradient, Proceedings of the National Academy of Sciences, 115, 7272-7277,
- 542 10.1073/pnas.1720488115, 2018.
- Yadav, S. P., Ibaraki, Y., and Dutta Gupta, S.: Estimation of the chlorophyll content of micropropagated potato
- 544 plants using RGB based image analysis, Plant Cell, Tissue and Organ Culture (PCTOC), 100, 183-188,
- 545 10.1007/s11240-009-9635-6, 2010.





546 List of Figures:

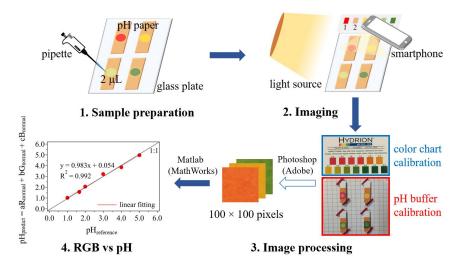
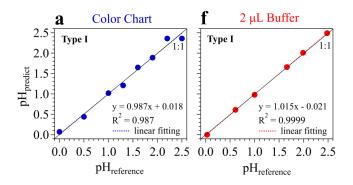
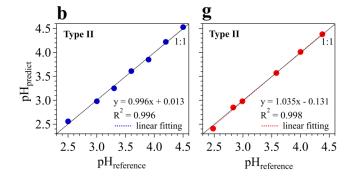


Figure 1: Schematic of using the RGB-based colorimetric method for pH estimation. For the color-chart-calibration method, both the color chart and the standard buffer samples are imaged into one digital photo for subsequent processing. For the standard-buffer-calibration method, only the standard buffer samples are used for imaging. Note that when using the standard-buffer-calibration results to predict the pH of aerosol samples, the photographing conditions for the samples are the same as those of the buffer calibration.

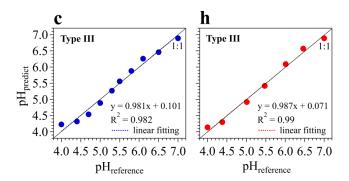




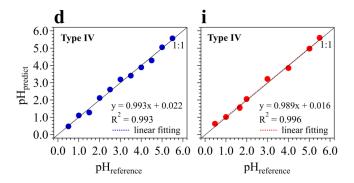




572







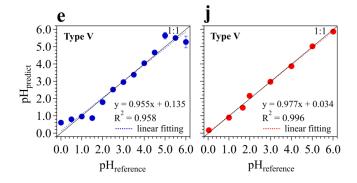
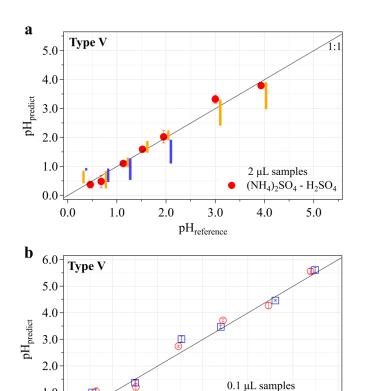


Figure 2: Predicted pH (pH_{predict}) using our RGB model versus the reference pH shown on the color chart and the pH-meter-probed-pH of the buffer samples (all denoted as pH_{reference}) respectively, for the five different pH papers: (a) and (f) Type I: 0 - 2.5, (b) and (g) Type II: 2.5 - 4.5, (c) and (h) Type III: 4.0 - 7.0, (d) and (i) Type IV: 0.5 - 5.5 and (e) and (j) Type V: 0 - 6.0. Blue symbols denote the established relationship based on color charts only. Red symbols represent the results for $2 \mu L$ of buffer droplets on pH papers. The error bars represent the standard deviation of five to six replicate experiments. Note that the error bars in most of the panels are smaller than the symbols.





597

598

599

600

601

602

603

1.0

0.0

1.0

2.0

594

Figure 3: pH estimation using the type V pH paper for samples with different volumes: (a) 2 μ L and (b) 0.1 μ L. pH_{predict} are calculated with the averaged coefficient vector [a, b, c] derived from three to six replicate experiments with the same amounts of standard buffers as of the samples under constant photographing conditions. The error bars represent the standard deviation of three to six replicate experiments. In (a), the heights of the orange and blue bars indicate the reported pH ranges measured with pH papers and Raman spectroscopy respectively, for (NH4)2SO4 - H2SO4 aerosols with particle sizes larger than 2.5 μ m in Craig et al. (2018). Each orange or blue bar has the same pH_{reference} as of the red symbol close to it. In (b), for processing the digital images of the 0.1 μ L samples, a square with 20 × 20 pixels at the center of the samples is cropped for subsequent colorimetric analyses.

3.0

 $pH_{reference}$

NH₄NO₃-HNO₃-C₃H₄O₄ (NH₄)₂SO₄-H₂SO₄-C₃H₄O₄

5.0

4.0

6.0

604 605

606 607



613

614

615

616

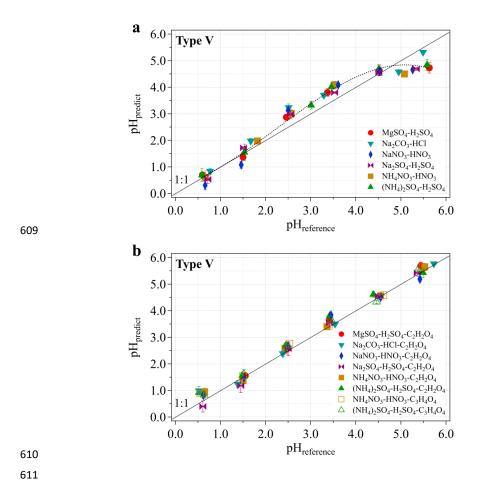


Figure 4: pH estimation using the type V pH paper for salt systems with only inorganic acids (a) and both inorganic and organic acids (b). $pH_{predict}$ are calculated with the averaged coefficient vector [a, b, c] derived from three replicate calibration experiments with standard buffers and under constant photographing conditions. The error bars represent the standard deviation of three to four replicate experiments. The dotted line in (a) is used to guide the eye.