Response to Interactive comment on "Best practices for precipitation sample storage for offline studies of ice nucleation" by Charlotte M. Beall et al.

The Authors wish to thank Referee 1 for their comments and discussion. We summarize our responses and changes made to the manuscript below. Note that line numbers refer to the latest, marked-up version of the manuscript:

Referee: "1. The 15 samples used as the basis of this paper seems a reasonable number; how-ever, they are all from the same location and therefore likely have a similar mix of INP types. Do the researchers have access to more diverse samples? Would these results hold if the sample was e.g. collected from a boreal forest? A desert? (see also comment on the 'correction factor' in point 3). Given the expertise in the author list, it would seem that access to a variety of samples would be possible. To emphasize this point : the title of this paper is ""Best practices for precipitation sample storage for offline studies of ice nucleation" – of universal importance. But in the Discussion "The aim of this study was to identify a storage protocol...in a coastal environment."– much more limited. The 'coastal' modifier is then repeatedly used but this isn't even universally coastal – it is a single coastal location. Either the authors should place the much more geographically restrictive information up front – title and abstract – or provide a larger diversity of samples. The latter, clearly, would be much more beneficial to the field as a whole."

We agree that a study with a greater diversity of samples and sites could be more broadly applicable, and perhaps provide greater insights. A focus on one location for demonstrating impacts of storage on precipitation samples allowed for a manageable study as one part of a PhD, and serves the purpose of highlighting that the lack of an existing standard storage protocol in the field is potentially problematic. These findings, thus, will serve as motivation for future efforts to quantify storage impacts on samples from a variety of environments, which will require either a series of field campaigns or a coordinated study between groups at different institutions/locations. To provide broader context for our dataset, we have updated Figure 1 following the assumptions of Petters and Wright, 2015 to estimate in-cloud INP concentrations from precipitation samples (i.e. 0.4 g condensed water content m^{-3}). Figure 1 shows that the INPs observed here are comparable to spectra reported previously for a wide range of marine and coastal environments, including the Caribbean, Bering Sea, East Pacific and nascent sea spray aerosol (DeMott et al., 2016). As INP spectra in this temperature regime cluster distinctly by air mass type (e.g. Figure 1-10 in Kanji et al., 2017), Fig. 1 indicates that the air masses sampled in this study were likely primarily marine. Regarding the reviewer's point "[...] this isn't even universally coastal […]", we agree that it remains to be seen how sensitivity to storage varies between sites with similar source types. This reinforces the need for future studies of the effects of storage, not only upon INPs in precipitation, but also with filter or impinger samples that are used in investigations globally.

We have made the following changes to explicitly state this assumption and reflect it consistently throughout the text:

Title: "Best practices for precipitation sample storage for offline studies of ice nucleation in marine and coastal environments"

Abstract, Line 25: "We provide the following recommendations for preservation of precipitation samples from coastal or marine environments intended for INP analysis…"

Introduction, Line 100: “Enhancements and losses of INPs according to storage protocol and treatment are reported, as well as recommendations for storage protocols that best preserve INPs in untreated, heated, and filtered precipitation samples from marine or coastal environments.”

Sec 3.1, Line 171: “ Following the assumptions in (Wright and Petters, 2015) to estimate in-cloud INP concentrations from precipitation samples (i.e. condensed water content of 0.4 g m^{-3} air), observations of INP concentrations in fresh precipitation samples are additionally compared to studies of field measurements conducted in marine and coastal environments. Figure 1 shows that atmospheric INP concentration estimates compare with INP concentrations observed in a range of marine and coastal environments, including the Caribbean, East Pacific, and Bering Sea, as well as laboratory-generated nascent sea spray aerosol (DeMott et al., 2016).”
Discussion, Line 282: “Additionally, the INP freezing temperatures and concentrations observed in this study compare with INPs observed in studies of marine and coastal environments (Fig. 1). As spectra in this regime (-5 to -20 °C and 10^{-5} to ~10^{-1} per L air, respectively) cluster distinctly by source type (see Fig. 1-10 in Kanji et al., 2017), Fig. 1 indicates that the dominant sources to air masses sampled in this study were marine. Considering that data in this study compare well with marine and coastal INPs from a variety of marine-influenced air masses (DeMott et al., 2016, Yang et al., 2019), the findings herein are likely relevant to samples from other marine and coastal environments.”

Discussion, Line 297: “If correspondence within 1 order of magnitude (or 2-3 °C) is desired, uncertainties associated with storage should also be considered in studies using samples from coastal or marine environments. Thus, uncertainty distributions provided in Tables 5-7 can be used to evaluate observed INP concentrations and responses to treatments in the context of potential changes due to storage. However, the degree to which INP sensitivity to storage varies by INP source (e.g. with soil-derived INP populations) remains to be tested.”
Conclusions, Line 370: “Based on all observations in this study, we provide the following recommendations for precipitation samples collected in coastal and marine environments for offline INP analyses...”

Referee: “2. Follow on. : At what time of year / conditions were the samples collected? Are these from the same or similar events? What is the diversity of conditions (season, meteorological, etc.)”

A summary of the meteorological conditions associated with each sample have been added to Table 1, and the following changes have been made to the text:

Sec. 2.1, Line 117: “Satellite composites from the National Weather Service Weather Prediction Center’s North American Surface Analysis Products were used for synoptic weather analysis to generally characterize each rain event (see Table 1). Atmospheric river (AR) events were identified using the AR Reanalysis Database described in (Guan and Waliser, 2015) and (Guan et al., 2018).”

Sec. 3.1 Line 165: “Figure 1 shows INP concentrations of 15 coastal rain samples, collected in a variety of meteorological conditions including scattered, low coastal rainclouds, frontal rain, and atmospheric river events (see Table 1).”

Referee: “3. Starting in the Abstract and continuing through the paper : “...non-heat-labile INPs being generally less sensitive to storage regime...” “Non-heat-labile INPs were generally less sensitive” This seems to be an assumption; the experiment determines abundances of heat or non-head labile INPs before and after but can not directly say something was changed or not. The authors should indicate that, based on abundances, they assume that the storage process is responsible for the change but not absolutely attribute it. As an example, a constant abundance could mean that no change was caused by storage or that

there were roughly equal rates of enhancement and deactivation; the measurements made would not be able to differential this, correct?"

The referee makes a good point that there are other potential causes of changes in INPs that should be discussed. Sample handling procedures, for example, could cause apparent differences in INP concentrations, or contamination in storage containers. However, we are able to distinguish changes due to sample handling and changes due to storage by considering the differences between sample replicates. The following changes to the text have been made:

Sec 3.2, line 218: "Replicate samples were processed for each storage protocol so that impacts of sample handling can be distinguished from storage impacts. For example, if settling occurs in bulk rain samples that are then divided into smaller volumes prior to storage, INP concentrations may differ between replicates of the bulk sample. . Thus, it is assumed that INP concentration changes that are greater than differences between replicates (grey bars in Figs 2-4) can be attributed to storage impacts. We also assume that stored:fresh INP concentration ratios of 1:1 indicate insensitivity to storage, although it is possible that enhancements and losses of equal magnitude could also result in a 1:1 concentration ratio."

Referee: If the assumption that heat labile INPs are more sensitive to storage, I don't believe the authors can offer (again, point made in Abstract and continuing through paper):"correction factors are provided so that INP measurements obtained from stored samples may be used to estimate concentrations in fresh samples"– wouldn't said correction factor necessarily be a function of the ratio of non- to heat labile INPs? Therefore the correction factor would not be universal but a function of the INP mix?

Due to referee #2's point that many of the correction factors are within measurement uncertainty for droplet assay techniques, we have updated Tables 5-7 with average changes in INP concentration and 95% confidence intervals that can be used to estimate uncertainty associated with storage in samples from marine and coastal environments (see response to point #1 above). Emissions of heat-labile particles can be increased in bloom-enhanced conditions, although it is variable (McCluskey et al., 2018), and considering bloom timescales (e.g. 10 days), bloom-enhanced marine sources would not have dominated air-masses sampled in a precipitation study.

The following changes have been made to the text to reflect this update:

Abstract: Finally, the estimated uncertainties associated with the 4 storage protocols are provided for untreated, heat-treated and filtered samples for INPs between -9 and -17 °C.

Conclusion, Line 398: "2. Estimates of uncertainty attributed to storage impacts and 95% confidence intervals for INP measurements obtained from stored samples are provided (see Tables 5-7)."

Legend Table 5: **Table 5. Estimate of uncertainty associated with storage impacts for INPs with activation temperatures between -9 and -17 °C measured in stored, untreated precipitation samples.** Confidence intervals were derived from the log-normal distribution of changes observed in INP concentrations due to storage (see Fig. 2 and details in Sect. 3.2). Temperature intervals where datapoints were too few to derive confidence intervals are indicated with "NA". Changes in INP concentration corresponding to enhancements or losses greater than 1 order of magnitude (losses <= -90% or enhancements >= +900%) in bold.

Legend Table 6: **Table 6. Estimate of uncertainty associated with storage impacts for INPs with activation temperatures between -9 and -17 °C measured in stored, heat-treated precipitation samples.** Confidence intervals were derived from the log-normal distribution of changes observed in INP concentrations due to storage (see Fig. 3 and details in Sect. 3.2). Changes in INP concentration corresponding to enhancements or losses greater than 1 order of magnitude (losses <= -90% or enhancements >= +900%) in bold.

Legend Table 7**: Table 7. Estimate of uncertainty associated with storage impacts for INPs with activation temperatures between -11 and -19 °C measured in stored, filtered precipitation samples.** Confidence intervals were derived from the log-normal distribution of changes observed in INP concentrations due to storage (see Fig. 2 and details in Sect. 3.2). Temperature intervals where datapoints were too few to derive confidence intervals are indicated with "NA". Changes in INP concentration corresponding to enhancements or losses greater than 1 order of magnitude (losses <= -90% or enhancements >= +900%) in bold.

Referee: "5. Introduction "Measurements of INPs suspended in precipitation are commonly made offline using a droplet freezing assay technique, and many studies report results from samples stored prior to processing. Storage protocols vary widely, including total storage time, time between collection and storage, and temperature fluctuations between collection, shipment and storage (if these details are provided at all), yet generally samples are stored between + 4◦C and -20◦C (see Table S1)." – These two sentences follow on a paragraph on INP in clouds. They are disparate concepts and should represent two new paragraphs: (1) how are off-line INP measurements made (they are not only by drop freezing assay – that is only the technique used here)? and (2) there should be a more complete description of storage used by previous researchers, not just a statement that it varies widely / table reference."

This paragraph has been updated with the suggested structure and content.

Introduction, Paragraph beginning Line 56: A number of online (real-time) and offline (processed post-collection) techniques exist for measurement of INPs for each ice nucleation mechanism, including condensation, deposition, immersion and contact freezing. However, as some simulations have shown that immersion mode freezing is the dominant mode of primary freezing in the atmosphere between 1000 and 200 hPa (Hoose et al, 2010), most techniques target immersion freezing. Despite the lack of time resolution, offline techniques enable measurement of INPs at modest supercooling (e.g. up to -5 °C) and temperature regimes where concentrations typically fall below detection limits of online instruments (DeMott et al., 2017). Offline instruments capable of immersion mode INP measurement include a number of droplet assays, in which sample suspensions are distributed among an array of droplets that are then cooled and frozen (e.g. Budke and Koop, 2015, Harrison et al., 2018, Hill et al., 2014, Whale et al., 2015) as well as other systems in which water is condensed onto particles collected on substrates prior to cooling and freezing (e.g. Mason et al., 2015). As they are designed for analysis of liquid suspensions, droplet freezing assay techniques are commonly used for measurement of INPs suspended in precipitation (e.g. Creamean et al., 2019, Rangel-Alvarado et al., 2015, Michaud et al., 2015, Stopelli et al., 2014, Wright et al., 2014).

Many studies report results from samples stored prior to processing. Storage protocols vary widely, including total storage time, time between collection and storage, and temperature fluctuations between collection, shipment and storage (if these details are provided at all, see summary Table S1). Storage temperatures range from -80 ºC (Vali et al., 1971) to +4 ºC (e.g. Petters and Wright, 2015, Failor et al., 2017, Joyce et al., 2019), yet generally samples are stored between +4 ºC and -20 ºC. Reported storage intervals range between hours (Schnell et al. 1977; Christner et al., 2008) to 48 years (Vasebi et al. 2019).

Referee: "6. Discussion, last paragraph starts "Significant enhancements in INP concentrations occurred less frequently than losses. Again, changes in the total particle size distribution could explain some of the observed INP concentration enhancements." – an important conclusion. However, the paragraph then changes topics to the impact of freezing on IN-active (biological) molecules. This is neither consistent with the topic of the paragraph nor is it part of the research outlined in the paper. Lines 259-269, as currently constituted, should be removed."

These lines have been removed from the last paragraph of the Discussion.

Abstract ": : :likely and an additional uncertainty in INP concentrations: : :" remove and?

Corrected.
Abstract, Line 25: "We provide the following recommendations for preservation of precipitation samples from coastal or marine environments intended for INP analysis: that samples be stored at -20 ºC to minimize storage artifacts, that changes due to storage are likely an additional uncertainty in INP concentrations…"

"Significant insights have been obtained: : :" 'highly uncertain" : please eliminate nonobjective terms like 'significant' (throughout paper) – these are reader dependent, not quantitative.

These terms have been removed from the paper, except for instances referring to Fisher's Exact Test. A statement to clarify this has been added to the Results section 3.2.

Sec. 3.2, Line 229: "The term "significant" henceforth is intended to describe INP losses or enhancements that correspond to frozen well fractions that are determined to be significantly different from corresponding fresh sample frozen well fractions, according to Fisher's Exact Test (i.e. filled markers in Figs. 2-4)."

Introduction, Line 78: "The understanding of storage effects on INPs suspended in precipitation is limited (Petters and Wright, 2015)…"

Responses to *Interactive comment on* "Best practices for precipitation sample storage for offline studies of ice nucleation" by Charlotte M. Beall et al.

The Authors would like to thank Gabor Vali for his helpful suggestions and discussion. A summary of our responses is below. Line numbers refer to the latest marked-up version of the manuscript.

Referee: "There is no indication in the paper of the kind of precipitation that was sampled. Presumably - vaguely deduced from the variations in the lengths of the sampling periods -
a variety of precipitation types are included. Probably, some light rain to more showery situations were involved. Cases with all warm-rain processes and cases with ice origin may have been involved. This may justify the choice of 'precipitation' in the title rather than 'rain'. If all events were from clouds with no ice-phase, a change in the title would be warranted to indicate so. This point isn't very important to the main theme of the

paper, but it could possibly make a difference for considerations of how the present results might apply to other situations."

Thank you for this suggestion. A summary of the types of precipitation events that were sampled has been added to Table 1. Meteorological conditions associated with precipitation ranged from AR events to warm, low altitude rain clouds. It is true that the choice of "precipitation" in the title and throughout the text was motivated by ambiguity regarding the ice or liquid origin of the samples. Although all samples were collected as liquid at ground-level, it is possible that ice-processes were involved in the precipitating clouds (judging by the low cloud-top temperatures in the NWS satellite composite analysis).

The following updates have been made to the text:

Sec. 2.1, Line 117: Satellite composites from the National Weather Service Weather Prediction Center's North American Surface Analysis Products were used for synoptic weather analysis to generally characterize each rain event (see Table 1). Atmospheric river (AR) events were identified using the AR Reanalysis Database described in (Guan and Waliser, 2015) and (Guan et al., 2018).

Sec. 3.1, Line 165: Figure 1 shows INP concentrations of 15 coastal rain samples, collected in a variety of meteorological conditions including scattered, low coastal rainclouds, frontal rain, and atmospheric river events (see Table 1). Observations generally fall within bounds of previously reported INP concentrations from precipitation and cloud water samples (grey shaded region, adapted from Petters and Wright, 2015). Observed freezing temperatures ranged from -4.0 to -18.4 ºC, with concentrations up to the limit of testing at 10^5 INP L^{-1} precipitation. AIS measurement uncertainties are represented with 95% binomial sampling intervals (Agresti and Coull, 1998).

Referee: "The main constraint mentioned in the paper and explicitly stated in the conclusions is that the results refer to coastal environments. This is not as helpful as could be, since precipitation and aerosol sources on the coasts may still include a very broad variety."

To provide broader context for our dataset, we have updated Figure 1 following the assumptions of Petters and Wright, 2015 to estimate in-cloud INP concentrations from precipitation samples (i.e. 0.4 g condensed water content m^{-3}). The updated Figure 1 shows that the INPs observed here are comparable to spectra reported previously for a wide range of marine and coastal environments, including the Caribbean, Bering Sea, East Pacific and nascent sea spray aerosol (DeMott et al., 2016). As INP spectra in this temperature regime cluster distinctly by air mass type (e.g. Figure 1-10 in Kanji et al., 2017), Fig. 1 indicates that the air masses sampled in this study were likely primarily marine.

The following updates have been made to the text:

Title: "Best practices for precipitation sample storage for offline studies of ice nucleation in marine and coastal environments**"**

Sec. 3.1, Line 171: " Following the assumptions in (Wright and Petters, 2015) to estimate in-cloud INP concentrations from precipitation samples (i.e. condensed water content of 0.4 g m^{-3} air), observations of INP concentrations in fresh precipitation samples are additionally compared to studies of field measurements conducted in marine and coastal environments. Figure 1

shows that atmospheric INP concentration estimates compare with INP concentrations observed in a range of marine and coastal environments, including the Caribbean, East Pacific, and Bering Sea, as well as laboratory-generated nascent sea spray aerosol (DeMott et al., 2016)."
Sec. 3.1, Line 282: "Additionally, the INPs in this study compare with INPs observed in studies of marine and coastal environments (Fig. 1). As spectra in this regime (-5 to -20 °C and 10^{-5} to ~10^{-1} per L air, respectively) cluster distinctly by source type (see Fig. 1-10 in Kanji et al., 2017), Fig. 1 indicates that the dominant sources to air masses sampled in this study were marine. Considering that data in this study are characteristic of marine and coastal INPs previously reported over a wide range of marine environments (DeMott et al., 2016), we assume that the findings herein are relevant to samples from other marine and coastal environments.

Sec. 3.1: Line 294: If correspondence within 1 order of magnitude (or 2-3 °C) is desired, uncertainties associated with storage should also be considered in studies using samples from coastal or marine environments. Thus, uncertainty distributions provided in Tables 5-7 can be used to evaluate observed INP concentrations and responses to treatments in the context of potential changes due to storage. However, the degree to which INP sensitivity to storage varies by INP source (e.g. with soil-derived INP populations) remains to be tested.

Referee: "Separating measurement variability from actual changes is important. Figures 2-4 include indications of measurement reproducibility with the gray bars adjacent to the data
point clusters. All of these bars are indicating values greater than unity. The caption to
Fig. 2 says that the bars represent the 'average difference between replicates'. How is
this to be interpreted? What conclusion can drawn from these data?"

The following changes have been made in Sec 3.2 to describe how grey bars are to be interpreted and conclusions that may be drawn from the data. This section has also been updated to address the referee's 1st comment on sample handling in "Minor points":

Sec. 3.2, Line 218: Replicate samples were processed for each storage protocol so that impacts of sample handling can be distinguished from storage impacts. For example, if settling occurs in bulk rain samples that are then divided into smaller volumes prior to storage, INP concentrations may differ between replicates of the bulk sample. Thus, it is assumed that INP concentration changes that are greater than differences between replicates (grey bars in Figs 2-4) can be attributed to storage impacts.

Referee: "A lingering uncertainty in the paper about whether these treatments were
applied to the fresh sample before division and storage, or just prior to INP measurement, is disconcerting. The discussion in lines following 218 seem to indicate that
filtering was done before freezing for storage. It would be good to have the sequence
better described."

Thank you for bringing this to our attention. The following change has been made to Sec. 2.2 Storage Protocols:
Line 136: Heat treatments and filters were applied to samples just prior to processing (i.e. treatments were not applied to samples prior to storage).

Referee: "The overall effects of the treatments are given as, on the average,
59% of INPs were found resistant to heat and 69% passed the filters. These numbers

are overly vague, as dependence of temperature can be expected as well as variations
from sample to sample. While such detail will not alter the data, it is relevant to possible explanations of the results."

This is a good point. The figures quoted above were calculated at the temperature of the next to last freezing event of the corresponding fresh sample (beyond which the data is not meaningful), again using the cumulative distribution. I recalculated the ratios of heat-treated and filtered to untreated INPs in fresh samples in the temperature intervals consistent with the rest of the manuscript (-9, -11, -13, -15 etc). This new way of calculating the filtered to untreated fractions yields a different answer regarding the general sizes of the INPs, interestingly, probably because smaller particles represent larger fraction of INPs only at the colder temperatures. It is also now necessary to discuss that we observed some enhancements in INPs after heating fresh samples (5 of the 15 samples). We have added the following detail and discussion:

Sec 3.1, Line 178: "In 5 of the 15 heat-treated samples, INP concentrations were increased by 1.9 – 13X between -9 and -11 °C (see Discussion). Excluding these 5 samples, the fraction of heat-resilient INPs varied between samples and generally increased with decreasing temperature. Geometric means and standard deviations of heat-treated:untreated INP ratios were 0.40 ×/÷ 1.9, 0.51 ×/÷ 2.0, and 0.62 ×/÷ 2.1 at -11 , -13, and -15 °C respectively.

Fractions of INPs < 0. 45 µm also varied between samples, with geometric means and standard deviations of 0.48 ×/÷ 1.73, 0.30 ×/÷ 3.4 and 0.37 ×/÷ 1.9 at -11 , -13, and -15 °C respectively. Mean values of heat-resilient INP fractions and INPs < 0.45 µm were calculated using the geometric mean, which is more appropriate than the arithmetic mean for describing a distribution of ratios (Fleming and Wallace, 1986)."

Discussion, Line 272: "The fractions of INPs < 0.45 µm observed in this study varied between 52 and 63% at -11 and -15 °C, respectively. Excluding the five heat-treated samples in which INP concentrations were enhanced (e.g. 1.9 - 13X between -9 and -11 °C), the average fraction of non-heat-labile INPs varied between 40 and 62% at -11 and -15 °C, respectively. INP enhancements in heat-treated samples are unexpected, as heat-treatments are typically applied assuming that heat destroys proteinaceous (e.g. biological) INPs. The causes of INP enhancements in heat-treated samples are unknown and have only been reported in coastal precipitation samples (Martin et al., 2017) and nascent sea spray aerosol (McCluskey et al., 2018).

Discussion, Line 305: "Despite the range of enhancements and losses of heat-sensitive INPs observed in fresh samples, non-heat-labile INPs were generally less sensitive to storage than the total INP population., and with the exception of samples stored at room temperature, all techniques yielded similar results with fewer enhancements or losses. "

Discussion, Line 330: In this study, a large fraction (30% to 48%, on average) of INPs observed in fresh precipitation samples were < 0.45 µm. Considering this and that INPs < 0.45 µm exhibit significant losses across all storage types, there is a risk that filter-treatments on stored samples in this study would lead to the underestimation of INPs < 0.45 µm.

Referee: On the level of internal consistency in the paper, it is worth asking how justified is the statement underlying conclusion 6 (line 280). Significantly greater losses are said to occur in storage for filtered samples. This is not really evident from a comparison of Fig. 2 with Fig. 4, or from the figures in Table 5 versus Table 7. Greater variability (larger 95% range) is found only for 'refrigeration' and 'freezing', while 'room temperature' and 'flash freezing' have narrower ranges and smaller standard deviations in Table 7 than in Table 5. Perhaps the claimed effect was clear for selected samples but not for the combined sample set.

This conclusion is based on the increased frequency of significant (Fisher's, $p<0.01$) data points on Figs 2 and 4. After making the suggested changes (see response to comment below), the 95% confidence intervals span losses > 1 order of magnitude across all protocols and most temperature intervals. We have added the following text to explain how Tables 5-7 may be interpreted:

Discussion, Line 291: While mean INP changes are within a factor of ~2 or less of fresh sample INP concentrations for all protocols except "Room temperature" (Table 5), none of the 4 storage protocols prevented significant losses or enhancements of INP concentrations in all samples (Fig. 2), indicating that INP concentration measurements on fresh precipitation are superior to measurements on stored samples. 95% confidence intervals in Table 5 span losses > 1 order of magnitude in all protocols across multiple temperature intervals. As uncertainties < 1 order of magnitude are necessary for the quantitative comparison between studies (DeMott et al., 2017), our results demonstrate that uncertainties associated with storage must be considered in studies of stored samples from coastal or marine environments. Thus, the uncertainty distributions provided in Tables 5-7 can be used to evaluate observed INP concentrations and responses to treatments in the context of potential changes due to storage. However, the degree to which INP sensitivity to storage varies by INP source (e.g. with soil-derived INP populations) remains to be tested.

Referee: "Tables 5-7 have some technical problems (see comment below on lines 185-189), but taking the data as is, most notable is the large range of variations for the corrections factors. Not just the 95% range, but even 50% spread: for the last line in Table 5, the 50% range is roughly 0.78 to 2.8. Experiments seldom lead to more accurate INP concentrations due to limitations in sample sizes (number of drops or vials). This reinforces the point that the results should be viewed as indications of the uncertainties associated with aging of samples during storage and not as correction factors that can usefully improve measured INP data in other studies. This argument is further supported by the potential for differences in the aging effects for precipitation at different times and locations. The current data provide help in weighing the importance of aging versus other sources of uncertainties in a given experimental design."

We agree that these results are better indications of the uncertainties associated with storage. Updates have been made to the tables, table legends and text to reflect this change:

Abstract: Finally, the estimated uncertainties associated with the 4 storage protocols are provided for untreated, heat-treated and filtered samples for INPs between -9 and -17 °C.

Conclusion, Line 375: "2. Estimates of uncertainty attributed to storage impacts and 95% confidence intervals for INP measurements obtained from stored samples are provided (see Tables 5-7)."

Legend Table 5: **Table 5. Estimate of uncertainty associated with storage impacts for INPs with activation temperatures between -9 and -17 °C measured in stored, untreated precipitation samples.** Confidence intervals were derived from the log-normal distribution of changes observed in INP concentrations due to storage (see Fig. 2 and details in Sect. 3.2). Temperature intervals where datapoints were too few to derive confidence intervals are indicated with "NA". Changes in INP concentration corresponding to enhancements or losses greater than 1 order of magnitude (losses <= -90% or enhancements >= +900%) in bold.

Legend Table 6: **Table 6. Estimate of uncertainty associated with storage impacts for INPs with activation temperatures between -9 and -17 °C measured in stored, heat-treated precipitation samples.** Confidence intervals were derived from the log-normal distribution of changes observed in INP concentrations due to storage (see Fig. 3 and details in Sect. 3.2). Changes in INP concentration corresponding to enhancements or losses greater than 1 order of magnitude (losses <= -90% or enhancements >= +900%) in bold.

Legend Table 7**: Table 7. Estimate of uncertainty associated with storage impacts for INPs with activation temperatures between -11 and -19 °C measured in stored, untreated precipitation samples.** Confidence intervals were derived from the log-normal distribution of changes observed in INP concentrations due to storage (see Fig. 2 and details in Sect. 3.2). Temperature intervals where datapoints were too few to derive confidence intervals are indicated with "NA". Changes in INP concentration corresponding to enhancements or losses greater than 1 order of magnitude (losses <= -90% or enhancements >= +900%) in bold.

Minor points:
Referee: "How was the sample division done for different treatments? While this can be expected to be a step without risk of introducing discrepancies among the samples, it is not
without such a possibility. Thus, the manner it was done should be described, as well
as any tests done to assure that this step doesn't lead to artifacts."

The following updates have been made to the text:
Sec. 2.2, Line 128: Prior to storage, 25 - 50 mL bulk sample aliquots were distributed directly from collection bottles into Falcon® tubes, shaking bottles ~10 s between each distribution.

Sec. 3.2, Line 218: Replicate samples were processed for each storage protocol so that impacts of sample handling can be distinguished from storage impacts. For example, if settling occurs in bulk rain samples that are then divided into smaller volumes prior to storage, INP concentrations may differ between replicates of the bulk sample. Thus, it is assumed that INP concentration changes that are greater than differences between replicates (grey bars in Figs 2-4) can be attributed to storage impacts.

Referee: Line 85 mentions samples getting divided into 24 bottles during collection. What is
the relationship between this and the division of the samples for different treatments?

This is a description of the precipitation collection device, which changes bottles on a rotating carousel at the specified time interval. There is no consistent relationship between the bottle numbers and sample division because sometimes we combined bottles corresponding to consecutive 1-hour sampling intervals in order to have enough volume for each of the sampling protocols, replicates, treatments, etc. This is described in Sec. 2.1: "The samples were distributed via the distributor arm into one of twenty-four 1-liter polypropylene bottles on an hourly time interval. Bottles were combined when the hourly precipitation volume was insufficient for sample separation and analysis (< 50 mL)."

I have updated the last line to help clarify:
Sec. 2.1, Line 112: "Bottles corresponding to consecutive 1-hour time intervals were combined when the hourly precipitation volume was insufficient for sample separation and analysis (< 50 mL per bottle)."

Figure 1 shows points near -5◦C for one sample. This should be of special interest but the paper doesn't mention it. Was the sample unusable?

We have added the following text to acknowledge the two warm-freezing observations (this sample was used in the storage experiments).

Sec. 3.1, Line 176: "However, two of the warmest-freezing INP observations in Fig. 1 (at -4.0 and -4.75 °C) exceed temperatures commonly observed in marine-influenced atmospheres, precipitation and cloudwater samples

Referee: Perhaps Fig.1 could be made less congested by showing only the interval 0◦C to -20◦C.
This is true, however, I think that seeing the whole Wright and Petters, 2015 and now DeMott et al., 2016 composite spectrum because it provides context for the regime we are observing.

Referee: Is there more than one point included in Figs. 2-4 for a sample from the same rain event and time period? Unfortunately, one can't determine from the figures how many data points are shown for each temperature. More than the number of rain events? The number of points differs for different temperatures. Is this because of limits in the temperature range of freezing events?

Yes, more than one point from the same sample is included in Figs. 2-4. This was motivated by the fact that a subset of the replicate samples exhibited differing sensitivity to storage (Fig. S5). However, replicates were not included in uncertainty factor calculations to avoid underweighting the small subset of samples that did not have replicates due to sample volume limitations. This is currently stated in the figure legends and table legends, and tables 2-4 show how many unique samples are represented in each figure vs how many replicates. The number of points differ due to limits of detection. There are typically fewer datapoints in the warmest and coldest temperature bin. At the warmest temperature bin, one of the samples (fresh or stored) is more likely to have 0 wells frozen, which would result in either a ratio of 0 or Inf. Ratios of zero were excluded because they are reflective of the limit of detection due to the number of droplets processed rather than a true lack of ice nucleating particles at this temperature. Similarly, datapoints tend to be fewer in the coldest temperature bin because in one or both of the samples (fresh and stored), all the wells had frozen.

The following has been added to the text:
Sec. 3.2 Line 191: "Numbers of datapoints in Figs 2-4 differ across the temperature intervals due to limits of detection (i.e. ratios were not calculated at temperatures where zero or all wells were frozen in the fresh and/or stored sample)."

Figure 2 legend: All samples were processed at one or two time intervals between 1 and 166 days post-collection (see Figs S1-S4). For samples processed at two intervals, both replicate samples are represented in the figure for a total of 14, 16, 18 and 12 samples in (a), (b), (c) and (d), respectively (see Table 2 for summary of sample and replicate numbers). Significant data are also labelled to indicate the sample number (01-15, see Table 1), and replicate ("A" or "B", and "U" indicates there were no replicates for the sample).

Figure 3 legend: All samples were processed at one or two time intervals between 1 and 166 days post-collection. For samples processed at two intervals, both replicate samples are represented in the figure for a total of 13, 16, 15 and 12 samples in (a), (b), (c) and (d), respectively (see Table 3 for summary of sample and replicate numbers).

Figure 4 legend: All samples were processed at one or two time intervals between 1 and 166 days post-collection. For samples processed at two intervals, both replicate samples are represented in the figure for a total of 13, 15, 16 and 12 samples in (a), (b), (c) and (d), respectively (see Table 4 for summary of sample and replicate numbers).
Figures 2-4 have been also been updated to show the numbers of samples represented for each protocol. Each filled data marker is now also labeled to show which sample and replicate it corresponds to.

Referee: "It would have been useful to identify by precipitation sample each data point in Figs 2-4, at least for the outliers. This would clarify, for example, if all the points with lowest values in Fig 2(a) are for the same sample or not, and if the same sample has the lowest points in Fig 2(b) and 2(c)"

This is a good idea, thank you. Figs 2-4 have been updated as suggested, with some text on each filled marker with the precipitation sample number and either the letter "A" or "B" to indicate which replicate it is. Samples without replicates are marked "U".

Referee: "Line 73: Were heat treatment and filtering applied before division for different storage temperatures, or just before INP analysis? One can assume it is the former, but the paper leaves it unspecified. Lines 101-102 still don't make clear what was done. Line 179 seems to indicate that filtration was done before storage."
Heat treatments and filtering was applied just before INP analysis, since this order is probably more common in e.g. field campaigns where samples must be frozen onsite before transport.

The following was added to Sec. 2.2, Line 136: "Heat treatments and filters were applied to samples just prior to processing (i.e. treatments were not applied to samples prior to storage)."

Referee: "Line 100: reference to 'section above' seems incorrect."
This line has been corrected in Sec 2.2, Line 133: "INP measurements were made in two or three time steps: within two hours of collection, and once or twice after storing using one of four storage protocols described above, depending on volume."

Referee: "Line 171: Is the ratio cited independent of the INP activity temperature?"
This ratio has been updated to reflect temperature dependence (see responses to referee comment p.3 beginning "The overall effects of the treatments are given as, on the average […]")

Referee: "Line 138: ' ... binned by 2◦C increments .. ' seems odd for cumulative data. More likely, values are 'determined (or calculated) at successive 2◦C intervals'. If that is not the case, please explain. The word 'binned' appears in numerous places in the text."
Thank you for bringing this to our attention. The following corrections have been made:

Sec. 3.2, Line 189: INP concentrations of stored replicate samples are compared with original fresh precipitation samples in Figures 2-4, calculated in successive 2 ºC increments between -7 and -19 ºC.

Sec 3.2, Line 194: All stored:fresh ratios were calculated from cumulative INP distributions in 2 ºC intervals, meaning that the INP concentration in each interval is inclusive of the concentration in the preceding (warmer) temperature interval. Thus,

in this study, deviations observed in a stored sample are not necessarily independent, i.e. the sensitivity of INPs to storage in one temperature interval could impact the observed changes in each of the following (colder) temperature interval. For example, in the fresh untreated precipitation samples (see Fig. 1), the contribution of INPs from the preceding 2 ºC interval ranges from 32 to 46% between -9 and -17 ºC.

Sec 3.2., Line 216: For each temperature interval containing data from at least two sets of replicate samples, the average difference in stored:fresh concentration ratios between replicates are represented with grey bars to indicate measurement variability
Sec 3.2, Line 226: "Finally, Fisher's Exact Test was applied to frozen and unfrozen well fractions between each stored sample and its corresponding fresh sample at each of the 2 ºC temperature intervals."

Sec 3.2, Line 232: "Results in Fig. 2 show that significant enhancements or losses of INPs occurred for all stored samples between -9 and -17 ºC, and that on average, stored samples exhibit INP losses (as indicated by the mean change in each temperature interval)."

Figure 2 legend: **Figure 2: Ratio of INP concentrations measured in untreated precipitation samples (stored:fresh), calculated in successive 2 ºC increments between -19 and -7 ºC.**
In temperature intervals containing stored:fresh ratios from at least two sets of replicate samples, grey bars represent the average difference between replicates.
Stored sample frozen well fractions that passed Fishers Exact Test ($p < 0.01$) for significant differences from original fresh sample frozen well fractions at each of the 5 temperatures are indicated with filled markers, and the mean change in each temperature interval is marked with a star.

Figure 3 legend: **Figure 3: Ratio of INP concentrations measured in heated precipitation samples (stored:fresh), calculated in successive 2 ºC increments between -19 and -7 ºC.**
In temperature intervals containing stored:fresh ratios from at least two sets of replicate samples, grey bars represent the average difference between replicates

Figure 4 legend: **Ratio of INP concentrations measured in filtered (0.45 µm) precipitation samples (stored:fresh), calculated in successive 2 ºC increments between -19 and -7 ºC.**
In temperature intervals containing stored:fresh ratios from at least two sets of replicate samples, grey bars represent the average difference between replicates

Referee: "Line 140: What does 'significant' refer to here? Maybe the authors meant 'measured'."
This line has been corrected in Sec 3.2, Line 190: This temperature range was chosen for the analysis because most fresh precipitation samples exhibited freezing activity between -7 and -19 ºC.

Referee: "Line 141: The cumulative values at any point are calculated by accounting for all freezing events (all frozen sample wells) at temperatures higher than the value at which the
concentrations is evaluated, not just those of the preceding value at 2◦C higher temperature. Also, in line 146, 'each' should be replaced by 'all', and line 147 should be
rephrased and clarified."
This line has been corrected in Sec 3.2, Line 194: All stored:fresh ratios were calculated from cumulative INP distributions in 2 ºC intervals, meaning that the INP concentration in each interval is inclusive of the concentration in all of the preceding

(warmer) temperature intervals.

Line 201: For example, in fresh untreated precipitation samples (see Fig. 1), 32% of the INP concentration calculated at -11 °C activated in one of the preceding (warmer) 2 °C temperature intervals. At -17 °C, this fraction is increased to 46%.

Referee: Line 157: ' .. containing data from at least two sets of replicate samples ...' seems to say that data points shown include replicates from the same rain event. This is brought up again in lines 187-188 and in the caption to Fig. 2.
Yes, this line is intended to make clear that replicates are represented in Fig. 2-4. See response to p. 6 comment beginning "Is there more than one point included in Figs. 2-4 for a sample from the same rain event and time period?"

Referee: "Lines 160-161: 'well counts' and 'well fractions' are not the same - please clarify."
This line has been corrected in Sec 3.2, Line 229: Finally, Fisher's Exact Test was applied to frozen and unfrozen well fractions between each stored sample and its corresponding fresh sample at each of the 2 ºC temperature intervals. Stored sample frozen well fractions that were significantly different ($p < 0.01$) from fresh sample frozen well fractions at each of the 5 temperatures are indicated with filled markers.

Referee: "Line 161: ' ... at each of the 5 temperatures ..' should probably be left out"

Line 228 has been corrected: "Stored sample frozen well fractions that were significantly different ($p < 0.01$) from fresh sample frozen well fractions are indicated with filled markers."

Referee: "Line 163: Here it says that all stored samples showed significant changes whereas only a few points are shaded in Fig. 2"
Line 232 in Sec 3.2. has been corrected: "Results in Fig. 2 show that significant enhancements or losses of INPs occurred in all storage protocols between -9 and -17 ºC, and that on average, stored samples exhibit INP losses (as indicated by the mean change in each temperature interval)."

Referee: "Line 179: The reference to Sect. 2.3 for detail is incorrect"
Line 248 in Sec 3.2 has been corrected: Effects of storage protocol on INP concentrations of filtered precipitation samples are shown in Figure 4 (0.45 µm syringe filter, see Sect. 2.2 for details).

Referee: Lines 185-189: Tables 5-7 indicate the range of impacts that may be expected on the basis of the data presented in this paper. The correction factors here given appear to have been derived combining data from all temperatures for given storage and treatment type. This has an inherent multiplicity problem as data at successively lower temperatures include all data from higher temperatures. Thus, a value for, say, -11◦C is also incorporated into the values at -13◦C, -15◦C etc. so that the ratio at -11◦C is given multiple, though uneven, weights when combining all the values for -11◦C and lower into calculating a mean and standard deviation for the given treatment. Also, all data were included in calculating the values in Tables 5-7, not just the cases for which the differences observed were shown to be statistically significant. One may wonder what the results would be of only those cases were included."

Thank you for bringing this to our attention. We agree that there is a multiplicity problem in combining the data from all intervals and have updated the tables to show average changes and confidence intervals for each 2 °C temperature increment. See also responses to p.4 comment beginning "Tables 5-7 have some technical problems […]" about how the tables and text have been changed to reflect that these data represent uncertainty associated with storage rather than correction factors.
I also agree that it would be interesting to recalculate these figures using the significant (Fisher's) datapoints only, but I couldn't think of a way to justify the exclusion of the insignificant datapoints in a calculation. It is also likely that such a distribution would not be log-normal, and then it wouldn't be meaningful to calculate the geometric mean and 95% confidence intervals.

Referee: "Line 192: What is meant by 'in situ' collection? Similarly, in line 241 'in situ dust' is vague."

Line 263 and Line 338 have been updated in the Discussion section: "The challenge in selecting a storage protocol for atmospheric samples (e.g. precipitation, cloud water, ambient atmosphere) is that the INP population composition is unknown, diverse, and the impact of any given technique on the different species may vary.
Considering that well-characterized IN-active dust and biological standards (Arizona Test Dust and Snomax®, respectively) are sensitive to storage conditions, it is possible that dust or biological INPs contributed to the observed INP losses.

Response to: Interactive comment on "Best practices for precipitation sample storage for offline studies of ice nucleation" by Charlotte M. Beall et al

The Authors wish to thank Referee 4 for their comments and discussion. We summarize our responses and changes made to the manuscript below. Note that line numbers refer to the latest, marked-up version of the manuscript:

Referee: "(1) Figure 1 and line 136: I am wondering, why the error bars shown are symmetrical. In a log-plot, I would assume unsymmetrical bars for symmetrical errors."

I am assuming that the referee is referring to the error bars on the most visible points (e.g. the points at -4 and -4.75 °C). The error bars represent 95% binomial sampling intervals as in (Agresti and Coull, 1998), and are not symmetrical. I realize the figure is a bit congested and it isn't possible to see the individual data points. I considered decreasing the x-axis, but given the updates to Fig. 1, I think it is important to keep the full composite INP spectrum visible.

Referee: "(2) L.138: Section 3.2: I am missing storage experiments with 'pure' water, since we know from our own experiments that even deionized/distilled water can become ice nucleating after several days."
This is very interesting, but we have not experienced this ourselves, in DI water that we have stored for weeks in polypropylene bottles, and are not aware of any literature reporting such an effect in deionized, distilled or otherwise 'pure' water. For those reasons we did not carry out such a test.

Referee: "(3) L.192-193, L262-263 (conclusion), L.292-293: Apparently, there are significant deviations in the stored:fresh ratios, both above and below 1. How can then simple correction factors be applied? In addition, I am highly skeptical about these correction factors: given that the actual correction factors are usually small (mostly between 0.9 and 1.8), they are likely much smaller than most other errors in such type of ice nucleation studies, and so their usefulness is questionable in my view. In addition, it is highly questionable whether these correction factors can be applied to studies at other locations,

using different sampling and investigation methods, and studying different(marine) samples. I would very much prefer to see instead the uncorrected raw data then in such studies. In summary, I do not concur with conclusion no. 2. The authors also seem to be skeptical as they state in lines 292-293: "However, it remains to be seen how INP sensitivity to storage varies by environment or INP composition.""

Considering these issues and suggestions made by the other reviewers, we agree that the results are better represented as indications of the uncertainties associated with storage. Updates have been made to the tables, table legends and text to reflect this change:

Abstract: Finally, the estimated uncertainties associated with the 4 storage protocols are provided for untreated, heat-treated and filtered samples for INPs between -9 and -17 °C.

Conclusion, Line 375: "2. Estimates of uncertainty attributed to storage impacts and 95% confidence intervals for INP measurements obtained from stored samples are provided (see Tables 5-7)."

Legend Table 5: **Table 5. Estimate of uncertainty associated with storage impacts for INPs with activation temperatures between -9 and -17 °C measured in stored, untreated precipitation samples.** Confidence intervals were derived from the log-normal distribution of changes observed in INP concentrations due to storage (see Fig. 2 and details in Sect. 3.2). Temperature intervals where datapoints were too few to derive confidence intervals are indicated with "NA". Changes in INP concentration corresponding to enhancements or losses greater than 1 order of magnitude (losses <= -90% or enhancements >= +900%) in bold.

Legend Table 6: **Table 6. Estimate of uncertainty associated with storage impacts for INPs with activation temperatures between -9 and -17 °C measured in stored, heat-treated precipitation samples.** Confidence intervals were derived from the log-normal distribution of changes observed in INP concentrations due to storage (see Fig. 3 and details in Sect. 3.2). Changes in INP concentration corresponding to enhancements or losses greater than 1 order of magnitude (losses <= -90% or enhancements >= +900%) in bold.

Legend Table 7**: Table 7. Estimate of uncertainty associated with storage impacts for INPs with activation temperatures between -11 and -19 °C measured in stored, filtered precipitation samples.** Confidence intervals were derived from the log-normal distribution of changes observed in INP concentrations due to storage (see Fig. 2 and details in Sect. 3.2). Temperature intervals where datapoints were too few to derive confidence intervals are indicated with "NA". Changes in INP concentration corresponding to enhancements or losses greater than 1 order of magnitude (losses <= -90% or enhancements >= +900%) in bold.

Referee: "Figure S1-S4: I do not understand what is plotted in Figures S1 through S4 in the supplement, and I am in doubt that it is correct. The captions say "INP losses or enhancements (%) *: : :*" What are losses in %? Shouldn't they be given as negative numbers? How can losses and enhancements be fitted simultaneously as a basis for correlation analysis, as the figure captions imply?
Even if not losses in percent are meant but if loss factors are presented, then losses would imply values smaller than 1. However, in none of the figures S1-S4 is there any point below the 10^0 line. How can that be, as figures 2-4 of the main paper clearly show that losses do occur?

Moreover, I am wondering whether plotting the losses or enhancements in percent does make sense at all. I think factors would be more suitable, because some of the changes are several orders of magnitudes. In particular for losses (not such much for enhancements), plotting them in percent may be misleading: for example losses by a factor of 10ˆ-2 or 10ˆ-4 (i.e., a difference of two orders of magnitude) would lead to a loss of nearly -100% in both cases (-99% or -99.99%). Note that losses cannot be lower than -100%!"

Thank you for bringing this to our attention. Figures S1-S4 were originally intended to show whether absolute change in INP concentration relates to the storage time interval, so that we could determine whether the magnitude of the change correlated in time, independent of the sign of the change. However, the referee brings up a good point that there is a problem of scale as losses approach (-) 100%, but enhancements have no upper bound.
Figures S1-S4 have been updated as suggested using the INP change factors, and the text has been updated as follows (to reflect the updated correlation factors):

Abstract, Line 23: Correlations between total storage time (1-166 days) and changes in INP concentrations were weak across sampling protocols, with the exception of INPs with freezing temperatures >= -9 ºC in samples stored at room temperature.

Sec. 3.2, Line 209: For INPs with freezing temperatures >= -9 °C in samples stored at room temperature, time is moderately correlated with changes in INP concentrations ($R^2 = 0.58$).

Conclusions, Line 379: 4. With the exception of warm-freezing INPs (freezing temperatures >= -9 °C) in samples stored at room temperature, we found little to no correlation between changes in INP concentrations and storage intervals on timescales between 1-166 days, indicating that most enhancements or losses are likely happening during freezing or on timescales < 24 hours.

Referee: "(5) L.88: "At the MESOM Laboratory parking lot: : :" To which of the two collection points given (lines 81-83) does this location belong?"

Corrected so that the lines referred to are consistent in how they refer to the 2nd location.
Sec. 2.1, Line 113: At the Isaacs Hall location, an ISO 6706 plastic graduated cylinder and plastic funnel, 27 cm in diameter, was used for precipitation collection.

Referee: "(6) L.262-263: ": : : it is worth noting that freezing is lethal for most cells" This statement is too general. Note that INTRACELLULAR freezing is lethal for most cells, while EXTRACELLULAR freezing is often not critical and, thus, survived by freeze-tolerating species."

These lines have been removed due to this issue, and others, also brought to our attention by the other referees.

Referee: "(7) L.458 (caption to Fig.3): "measured in heated precipitation samples" When were the samples heated? Directly after collection, or just before measurement?"
Treatments were applied just prior to measurement. The following changes have been made to the text:

Sec. 2.2: Heat treatments and filters were applied to samples just prior to processing (i.e. treatments were not applied to samples prior to storage).

Figure 3 Legend: **Figure 3: Ratio of INP concentrations measured in heated precipitation samples (stored:fresh), calculated in successive 2 ºC increments between -19 and -7 ºC.** Same samples as shown in Figure 2, but heated to 95 ºC for 20 minutes just prior to measurement to eliminate heat-labile INPs (see Methods Sect. 2.2 for details).

Referee: "(8) L.468 (caption to Fig.4): "measured in filtered (0.45 μm) precipitation samples"
When were the samples filtered? Directly after collection, or just before measurement?"
This legend has also been updated:

Figure 4 Legend: **Figure 4: Ratio of INP concentrations measured in filtered (0.45 µm) precipitation samples (stored:fresh), calculated in successive 2 ºC increments between -19 and -7 ºC.** Same samples as in Fig. 2 but filtered with a 0.45 µm syringe filter prior to measurement (see Methods Sect. 2.2 for details).

Referee: "(9) Tables 5-7: Please provide a few sentences of explanation on the 95% confidence interval limits. What exactly do these values imply and, more importantly, how can they be applied? For example, considering line 2 in Table 5: the suggested correction factor is 1.72. The confidence limits of this correction factor are 0.25 and 11.27, implying that the correction factor could also be significantly below 1. I was wondering then, given this large confidence interval, whether it is useful at all to make such a correction (see also my comment 3 above)"

Tables 5-7 and text have been updated to reflect changes suggested in the referee's comment #3. Additionally, the following has been added to the text to explain how these values may be interpreted:

Discussion, Line 289: While mean INP changes are within a factor of ~2 or less of fresh sample INP concentrations for all protocols except "Room temperature" (Table 5), none of the 4 storage protocols prevented significant losses or enhancements of INP concentrations in all samples (Fig. 2), indicating that INP concentration measurements on fresh precipitation are superior to measurements on stored samples. 95% confidence intervals in Table 5 span losses > 1 order of magnitude in all protocols across multiple temperature intervals. These uncertainties equal or exceed INP measurement uncertainties (1-2 orders of magnitude) at temperatures > -20 °C due to discrepancies between instruments (DeMott et al., 2017). If correspondence within 1 order of magnitude (or 2-3 °C) is desired, uncertainties associated with storage should also be considered in studies using samples from coastal or marine environments. Thus, uncertainty distributions provided in Tables 5-7 can be used to evaluate observed INP concentrations and responses to treatments in the context of potential changes due to storage. However, the degree to which INP sensitivity to storage varies by INP source (e.g. with soil-derived INP populations) remains to be tested.

Best practices for precipitation sample storage for offline studies of ice nucleation in marine and coastal environments

~~Best practices for precipitation sample storage for offline studies of ice nucleation~~

Charlotte M. Beall[1], Dolan Lucero[2], Thomas C. Hill[3], Paul J. DeMott[3], M. Dale Stokes[1], Kimberly A. Prather[1,4]

[1]Scripps Institution of Oceanography, University of California San Diego, La Jolla, CA, 92037, USA
[2]Department of Earth and Environmental Science, New Mexico Institute of Mining and Technology, Socorro, NM, 87801, USA
[3]Department of Atmospheric Sciences, Colorado State University, Fort Collins, CO, 80523, USA
[4]Department of Chemistry and Biochemistry, University of California San Diego, La Jolla, CA, 92093, USA

Correspondence to: Kimberly A. Prather (kprather@ucsd.edu)

Abstract. Ice nucleating particles (INPs) are efficiently removed from clouds through precipitation, a convenience of nature for the study of these very rare particles that influence multiple climate-relevant cloud properties including ice crystal concentrations, size distributions, and phase-partitioning processes. INPs suspended in precipitation can be used to estimate in-cloud INP concentrations and to infer their original composition. Offline droplet assays are commonly used to measure INP concentrations in precipitation samples. Heat and filtration "treatments" are also used to probe INP composition and size ranges. Many previous studies report storing samples prior to INP analyses, but little is known about the effects of storage on INP concentration or their sensitivity to treatments. Here, through a study of 15 precipitation samples collected at a coastal location in La Jolla, CA, USA, we found ~~significant~~ INP concentration changes up to > 1 order of magnitude caused by storage to concentrations of INPs with warm to moderate freezing temperatures (-7 to -19 ºC). We compared four conditions: 1.) storage at room temperature (~~+~~ 21-23 ºC), 2.) storage at +4 ºC 3.) storage at -20 ºC, and 4.) flash freezing samples with liquid nitrogen prior to storage at -20 ºC. Results demonstrate that storage can lead to both enhancements and losses of greater than one order of magnitude, with non-heat-labile INPs being generally less sensitive to storage regime, but significant losses of INPs smaller than 0.45 µm in all tested storage protocols. Correlations between total storage time (1-166 days) and changes in INP concentrations were weak across sampling protocols, with the exception of INPs with freezing temperatures >= -9 ºC in samples stored at room temperature ~~or +4 ºC~~. We provide the following recommendations for preservation of precipitation samples from coastal or marine environments intended for INP analysis: that samples be stored at -20 ºC to minimize storage artifacts, that changes due to storage are likely ~~and~~ an additional uncertainty in INP concentrations, and that filtration treatments be applied only to fresh samples. At the freezing temperature -11 °C, a~~A~~verage INP concentration losses of 51~~72~~%, 7~~44~~2%, 16~~25~~% and 41~~32~~% were observed for untreated samples stored using the room temperature, +4 ºC, -20 ºC, and flash frozen protocols, respectively. Finally, the estimated uncertainties associated with the 4 storage protocols are provided for untreated, heat-treated and filtered samples for INPs between -9 and -17 °C.

~~Finally, correction factors are provided so that INP measurements obtained from stored samples may be used to estimate concentrations in fresh samples.~~

1. Introduction

In-cloud ice crystals and their formation processes are critical features of Earth's radiative and hydrological balance, affecting multiple climate-relevant cloud properties including cloud lifetime, reflectivity, and precipitation efficiency (DeMott et al., 2010; Lohmann, 2002; Lohmann and Feichter, 2005; Tan et al., 2016; Creamean et al., 2013). Ice nucleating particles (INPs) impact ice crystal concentrations and size distributions in clouds by triggering the freezing of droplets at temperatures above the homogeneous freezing point of water (-38 ºC).

INPs have been sampled in clouds and precipitation for decades (e.g. Rogers et al., 1998; Vali, 1971; Vali, 1966) to measure abundances, probe their compositions and investigate the extent to which they impact the properties of clouds. There are several caveats to consider when inferring in-cloud INP concentrations or properties from precipitation samples (Petters and Wright, 2015~~a~~), including "sweep-out" of additional INPs as the hydrometeor traverses the atmosphere below the cloud (Vali, 1974) and heterogeneous chemistry due to adsorption or absorption of gases (Hegg and Hobbs, 1982; Kulmala et al., 1997; Lim et al., 2010). However, assessing the composition of INPs in precipitation samples is more straightforward than cloud particles. Thus, the number of publications reporting measurements of INP concentrations in precipitation has increased ~~significantly~~ over the past decade. ~~Significant~~ Numerable insights have been obtained in previous precipitated-based INP studies, including the efficient depletion of INPs relative to other aerosols of similar size in precipitating clouds (Stopelli et al., 2015), constraints on minimum enhancement factors for secondary ice formation processes (Petters and Wright, 2015~~b~~), and the identification, characteristics and distribution of various INP populations (e.g. Christner et al., 2008a; Hader et al., 2014; Stopelli et al., 2017). INP concentrations in precipitation have been used to estimate in-cloud concentrations, based on assumptions that the majority of particles (86%) in precipitation originate from the cloud rather than the atmospheric column through which the hydrometeor descended (Wright et al., 2014). Along the same line of reasoning, INPs in precipitation have also been used to infer sources and composition of in-cloud INP populations (e.g. Martin et al., 2019 and Michaud et al., 2014, respectively).

A number of online (real-time) and offline (processed post-collection) techniques exist for measurement of INPs for each ice nucleation mechanism, including condensation, deposition, immersion and contact freezing. However, as some simulations have shown that immersion mode freezing is the dominant mode of primary freezing in the atmosphere between 1000 and 200 hPa (Hoose et al, 2010), most techniques target immersion freezing. Despite the lack of time resolution, offline techniques enable measurement of INPs at modest supercooling (e.g. up to -5 °C) and temperature regimes where concentrations typically fall below detection limits of online instruments (DeMott et al., 2017). Offline instruments capable of immersion mode INP measurement include a number of droplet assays, in which sample suspensions are distributed among an array of droplets that are then cooled and frozen (e.g. Budke and Koop, 2015, Harrison et al., 2018, Hill et al., 2014, Whale et al., 2015) as well as other systems in which water is condensed onto particles collected on substrates prior to cooling and freezing (e.g. Mason et al., 2015). As they are designed for analysis of liquid suspensions, droplet freezing assay techniques are commonly used for m~~M~~easurement~~s~~ of INPs suspended in precipitation (e.g. Creamean et al., 2019,

Rangel-Alvarado et al., 2015, Michaud et al., 2015, Stopelli et al., 2014, Wright et al., 2014).~~are commonly made offline using a droplet freezing assay technique,~~

~~and~~

Many studies report results from samples stored prior to processing. Storage protocols vary widely, including total storage time, time between collection and storage, and temperature fluctuations between collection, shipment and storage (if these details are provided at all, see summary Table S1). Storage temperatures range from -80 ºC (Vali et al., 1971) to +4 ºC (e.g. Petters and Wright, 2015, Failor et al., 2017, Joyce et al., 2019), yet generally samples are stored between +4 ºC and -20 ºC. Reported storage intervals range between hours (Schnell et al., 1977; Christner et al., 2008) to 48 years (Vasebi et al., 2019).

~~many studies report results from samples stored prior to processing. Storage protocols vary widely, including total storage time, time between collection and storage, and temperature fluctuations between collection, shipment and storage (if these details are provided at all), yet generally samples are stored between + 4 ºC and -20 ºC (see Table S1).~~

~~The~~ The understanding of ~~effects of storage~~ storage effects ~~on~~on INPs suspended in precipitation ~~are highly uncertain~~is limited (Petters and Wright, 2015~~b~~), and the understanding of storage effects on INPs collected on filters is similarly lacking (Wex et al., 2019). Stopelli et al. (2014a) studied INP concentrations in a snow sample stored at +4 ºC and observed a decrease in the concentration of INPs active at -10 ºC over 30 days by a factor of ~2. Schnell (1977) reported significant losses in fog and seawater samples after storage at room temperature for short periods (6-11 hours). Several studies have reported on the lability of commercially available dust and biological IN entities in storage above 0 ºC or under freezing conditions, including Arizona Test Dust and SnoMax® (Perkins et al., 2020; Polen et al., 2016; Wex et al., 2015), and similar labilities could affect the INPs of similar composition in precipitation samples (Creamean et al., 2013; Martin et al., 2019). Considering the abundance of precipitation based INP studies, the lack of bounds on potential impacts of storage on INP concentration measurements represents a critical uncertainty in conclusions derived from data on stored samples. Furthermore, to determine INP activation mechanisms and composition, previous studies have applied "treatments" to precipitation samples, including heat, filtration, enzymes and peroxide, (e.g. Hill et al., 2014) but it is unknown to what extent storage affects the results of such experiments.

Here we investigate the effects of four storage protocols on INPs using 15 precipitation samples collected between 9/22/2016 and 11/22/2019 at two coastal sites at Scripps Institution of Oceanography, La Jolla, CA, USA: 1.) storage at room temperature (+ 21-23 ºC) , 2.) storage at +4 ºC ("refrigerated"), 3.) storage at -20 ºC ("frozen"), and 4.) flash freezing samples with liquid nitrogen prior to storage at -20 ºC ("flash frozen"). The abundance of previous studies that report storage between +4 ºC and -20 ºC motivated the choice of techniques 2 and 3 (see Table S1). Room temperature storage was chosen to provide context as a "worst-case scenario", and the flash freezing technique was chosen to investigate whether any changes of INP concentrations could be mitigated by instantaneous freezing prior to storage. The 15 precipitation samples in this study were divided into several replicates so that the concentration of INPs could be measured in untreated, heated, and filtered samples when fresh, and again after storage using the 4 techniques described above. Sample replicates were additionally processed at 2 different points in time to investigate the effects of total storage time on INP concentration measurements. Enhancements

and losses of INPs according to storage protocol and treatment are reported, as well as recommendations for storage protocols that best preserve INPs in untreated, heated, and filtered precipitation samples from ~~coastal environments~~marine or coastal environments.

2. Methods

2.1 Precipitation Sample Collection

Precipitation samples were collected at two coastal locations at Scripps Institution of Oceanography (32.87 N 177.25 W): the rooftop of the Ellen Browning Scripps Memorial Pier laboratory (32.8662 °N, 117.2544 °W) (10 meters above sea level) and the rooftop of a storage container next to Isaacs Hall (32.8698 °N, 117.2522 °W, 58 meters above sea level, 500 m inland). Collection technique varied based on location. At the SIO pier, the Teledyne ISCO model 6712 commercial water sampler (Teledyne ISCO, Inc., US) was used. A plastic funnel, 27 cm in diameter, and Tygon tubing, connected the sampler inlet to the water sampler's distributor arm. The samples were distributed via the distributor arm into one of twenty-four 1-liter polypropylene bottles on an hourly time interval. Bottles corresponding to consecutive 1-hour time intervals were combined when the hourly precipitation volume was insufficient for sample separation and analysis (< 50 mL per bottle). At the ~~MESOM Laboratory parking lot,~~ Isaacs Hall location, an ISO 6706 plastic graduated cylinder and plastic funnel, 27 cm in diameter, was used for precipitation collection. At both sites, ring stands supported the collection funnels approximately 60 cm above the rooftop. All funnels, tubing, cylinders, and bottles were cleaned with 10% hydrogen peroxide for 10 minutes and rinsed with milli-Q purified water three times immediately before each sampling event. Satellite composites from the National Weather Service Weather Prediction Center's North American Surface Analysis Products were used for synoptic weather analysis to generally characterize each rain event (see Table 1). Atmospheric river (AR) events were identified using the AR Reanalysis Database described in (Guan and Waliser, 2015) and (Guan et al., 2018).

2.2 Storage Protocols

The following sample storage protocols were used: frozen at -20 ºC, refrigerated at 4 ºC, room temperature (21 - 23 ºC), and flash freezing, or flashing with liquid nitrogen (-196 ºC) before frozen at -20 ºC. All techniques except storage at room temperature are commonly used for offline INP analysis (see Table S1). Excluding the samples that were flash frozen, all samples were stored in 50 mL sterile plastic Falcon® tubes (Corning Life Sciences, Corning, NY, USA). Flash frozen samples were stored in polypropylene 5 mL cryovials. ~~Excluding the samples that were flash frozen, all samples were stored in 50 mL sterile plastic Falcon® tubes (Corning Life Sciences, Corning, NY, USA). Flash frozen samples were stored in polypropylene 5 mL cryovials.~~ Prior to storage, 25 - 50 mL bulk sample aliquots were distributed from collection bottles into Falcon® tubes, shaking bottles ~10 s between each distribution. Not all samples were stored using all four~~4~~ of the storage protocols due to limited volume for some samples. See Tables 2-4 for a summary of the number of samples studied for each storage protocol. Precipitation samples were stored for varying intervals between 1 and 166 days to investigate effects of storage time on INP concentrations. INP measurements were made in two or three time steps: within two hours of collection, and once or twice after storing using one of four storage protocols described ~~in the section~~ above, depending on volume. Stored and fresh samples

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were analysed in three treatment conditions: 1) raw untreated precipitation, 2) heated over a 95 ºC water bath for 20 minutes and 3) filtered through a 0.45 µm surfactant-free cellulose acetate syringe-filter (Thermo Scientific™ Nalgene™, Waltham, MA, USA). Heat treatments and filters were applied to samples just prior to processing (i.e. treatments were not applied to samples prior to storage).

2.3 INP Analysis

The automated ice spectrometer (AIS) is an offline immersion-mode freezing assay which is described elsewhere (Beall et al., 2017). Briefly, 50 uL aliquots of sample are pipetted into two sterile 96-well polypropylene PCR plates. The plates are inserted into an aluminium block, machined to hold PCR plates, that sits in the coolant bath of a Fisher Scientific Isotemp® Circulator. A thermistor placed atop the left side of the aluminium block, below the PCR plate, recorded temperature. An acrylic plate separated the PCR plates from the ambient lab air. In the headspace between the acrylic plate and the PCR plates, nitrogen gas flowed at a flow rate of 14 Lpm to reduce temperature stratification in the samples (Beall et al., 2017). The nitrogen gas was cooled before emission by passing through the chiller via copper tubing. A 0.5 Megapixel monochrome camera (Point Grey Blackfly 0.5MP Mono GigE POE) performed the image capture. Custom LabView software controlled the camera settings, the rate the chiller cooled, and displayed the temperature of the thermistor.

A control milli-Q water sample is used, typically in the first 30 wells of each sample run, to detect contamination and for subsequent INP concentration calculations. Thirty wells were used per sample to achieve a limit of detection of 0.678 IN mL^{-1}. For each run, the chiller was cooled to -35°C. As the chiller cools the sample plates (1 ˚C/min), the custom LabView virtual instrument records the location and temperature of the freezing event as they occur. Freezing events are detected by the change in pixel intensity of the sample as it changes from liquid to solid.

2.4 Particle Size Distributions

Size distributions of insoluble particles suspended in the fresh and stored precipitation samples were measured using the Multi-sizing Advanced Nanoparticle Tracking Analysis (MANTA) ViewSizer 3000 (Manta Instruments Inc.). The Manta ViewSizer 3000 applies multi-spectral particle tracking analysis (m-PTA) to obtain size distributions of particles of sizes between 10 and 2000 nm with three solid-state lasers with wavelengths of 450 nm, 520 nm and 650 nm. m-PTA has been shown to outperform traditional dynamic light scattering (DLS) techniques when measuring polydisperse particles in suspension (McElfresh et al., 2018). For analysis, 300 videos of the illuminated particles in suspension are recorded, each 10 seconds in length. The software tracks each particle individually, obtaining particle size and number concentration from their Brownian motion and the imaged sample volume.

3 Results

3.1 INP concentrations in fresh precipitation samples

Figure 1 shows INP concentrations of 15 coastal ~~precipitation~~ rain samples, collected in a variety of meteorological conditions including scattered, low coastal rainclouds, frontal rain, and atmospheric river events (see Table 1). Observations generally fall within bounds of previously reported INP concentrations from precipitation and cloud water samples (grey shaded region,

adapted from Petters and Wright, 2015). Observed freezing temperatures ranged from -4.0 to -18.4 ºC, with concentrations up to the limit of testing at 10^5 INP L^{-1} precipitation. AIS measurement uncertainties are represented with 95% binomial sampling intervals (Agresti and Coull, 1998).

Following the assumptions in (Wright and Petters, 2015) to estimate in-cloud INP concentrations from precipitation samples (i.e. condensed water content of 0.4 g m^{-3} air), observations of INP concentrations in fresh precipitation samples are additionally compared to studies of field measurements conducted in marine and coastal environments. Figure 1 shows that atmospheric INP concentration estimates compare with INP concentrations observed in a range of marine and coastal environments, including the Caribbean, East Pacific, and Bering Sea, as well as laboratory-generated nascent sea spray aerosol (DeMott et al., 2016). However, two of the warmest-freezing INP observations in Fig. 1 (at -4.0 and -4.75 ºC) exceed temperatures commonly observed in marine-influenced atmospheres, precipitation and cloudwater samples.

In 5 of the 15 heat-treated samples, INP concentrations were increased by 1.9 – 13X between -9 and -11 °C (see Discussion). Excluding these 5 samples, the fraction of heat-resilient INPs varied between samples and generally increased with decreasing temperature. Geometric means and standard deviations of heat-treated:untreated INP ratios were 0.40 ×/÷ 1.9, 0.51 ×/÷ 2.0, and 0.62 ×/÷ 2.1 at -11 , -13, and -15 °C respectively.

Fractions of INPs < 0. 45 µm also varied between samples, with geometric means and standard deviations of 0.48 ×/÷ 1.73, 0.30 ×/÷ 3.4 and 0.37 ×/÷ 1.9 at -11 , -13, and -15 °C respectively. Mean values of heat-resilient INP fractions and INPs < 0.45 µm were calculated using the geometric mean, which is more appropriate than the arithmetic mean for describing a distribution of ratios (Fleming and Wallace, 1986).~~Observations generally fall within bounds of previously reported INP concentrations from precipitation and cloud water samples (grey shaded region, adapted from Petters and Wright, 2015b). Observed freezing temperatures ranged from -4.0 to -18.4 ºC, with concentrations up to the limit of testing at 10^5 INP L^{-1} precipitation. AIS measurement uncertainties are represented with 95% binomial sampling intervals (Agresti and Coull, 1998).~~

3.2 Effects of sample storage on INP concentration measurements

INP concentrations of stored replicate samples are compared with original fresh precipitation samples in Figures 2-4, ~~binned~~ calculated in successive ~~by~~ 2 ºC increments between -7 and -19 ºC. This temperature range was chosen for the analysis because most fresh precipitation samples exhibited ~~significant~~ freezing activity between -7 and -19 ºC. Numbers of datapoints in Figs 2-4 differ across the temperature intervals due to limits of detection (i.e. ratios were not calculated at temperatures where zero or all wells were frozen in the fresh and/or stored sample).

All stored:fresh ratios were calculated from cumulative INP distributions ~~binned by~~in 2 ºC intervals, meaning that the INP concentration in each interval~~bin~~ is inclusive of the concentration in all of the preceding (warmer) temperature intervals~~ bin~~. The choice of the cumulative distribution was motivated by the fact that it is standard in INP studies to report INP concentrations in terms of the cumulative distribution, and it is important to consider impacts of storage on cumulative INP distributions and any conclusions derived from them. Thus, in this study, ~~significant~~ deviations observed in a stored sample are not necessarily independent, i.e. the sensitivity of INPs to storage in one temperature ~~bin~~ interval could impact the observed changes in ~~each~~ all of the following (colder) temperature ~~bins~~interval. For example, in fresh untreated precipitation samples

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(see Fig. 1), 32% of the INP concentration calculated at -11 °C activated in one of the preceding (warmer) 2 °C temperature intervals. At -17 °C, this fraction is increased to 46%.

~~For example, in the fresh untreated precipitation samples (see Fig. 1), the contribution of INPs from the preceding 2 ºC bin ranges from 32 to 46% between -9 and -17 ºC.~~

To investigate correlations between sample storage time and INP enhancements or losses, duplicate samples were archived (when sufficient volume was available) so that each sample could be processed at two distinct points post-collection (see example Fig. S1). For INPs with freezing temperatures >= -9 °C in samples stored at room temperature ~~or +4 °C~~, time is moderately correlated with changes in INP concentrations (R^2 = 0.58~~62 and 0.53, respectively~~). Figure S5 shows how losses of warm-freezing INPs in samples stored at +4 °C and room temperature impact the cumulative INP spectra for a select sample. Beyond these exceptions, little to no correlation between storage time and INP enhancements or losses was found for untreated, heated and filtered samples (see Figs S1-S4). This indicates that most of the changes in INPs observed may occur on shorter timescales than those studied here, i.e. < 24 hours.

Figure 2 shows the ratio of stored sample to fresh sample INP concentrations for untreated precipitation samples stored under four conditions: **(a)** room temperature (21 – 23 ºC), **(b)** refrigerated (+ 4 ºC), **(c)** frozen (-20 ºC) and **(d)** flash frozen with liquid nitrogen before storing at -20 ºC. Markers above the 1:1 line indicate enhancements in INP concentration from the fresh sample, while markers below indicate losses. For each temperature ~~bin~~ interval containing data from at least two sets of replicate samples, the average difference in stored:fresh concentration ratios between replicates are represented with grey bars to indicate measurement variability. Replicate samples were processed for each storage protocol so that impacts of sample handling can be distinguished from storage impacts. For example, if settling occurs in bulk rain samples that are then divided into smaller volumes prior to storage, INP concentrations may differ between replicates of the bulk sample. Thus, it is assumed that INP concentration changes that are greater than differences between replicates (grey bars in Figs 2-4) can be attributed to storage impacts. We also assume that stored:fresh INP concentration ratios of 1:1 indicate insensitivity to storage, although it is possible that enhancements and losses of equal magnitude could also result in a 1:1 concentration ratio.

Finally, Fisher's Exact Test was applied to frozen and unfrozen well ~~counts~~ fractions between each stored sample and its corresponding fresh sample at each of the 2 ºC temperature intervals~~bins~~. Stored sample frozen well ~~counts~~ fractions that were significantly different ($p < 0.01$) from fresh sample frozen well ~~fractions~~ fractions ~~as a fraction of total sample wells at each of the 5 temperatures~~ are indicated with filled markers. The term "significant" henceforth is intended to describe INP losses or enhancements that correspond to frozen well fractions that are determined to be significantly different from corresponding fresh sample frozen well fractions, according to Fisher's Exact Test (i.e. filled markers in Figs. 2-4).

Results in Fig. 2 show that significant enhancements or losses of INPs occurred ~~for all stored samples~~in all storage protocols between -9 and -17 ºC, and that on average, stored samples exhibit INP losses (as indicated by the mean change in each temperature interval~~bin~~). In frozen and flash frozen samples, all enhancements and losses fall within ± 1 order of magnitude, whereas several significant INP losses beyond 1 order of magnitude are shown in room and refrigerated samples. INP concentration changes >= 1 order of magnitude are greater than changes in the ratios of the total insoluble particle population

10 – 2000nm during storage (see Fig. S6). This indicates that the INPs in these samples are more sensitive to storage than the total insoluble particle population. Fig. S5 illustrates the impacts of the 4 storage protocols on the full IN spectra of a select untreated precipitation sample at two time intervals, 27 days and 64 days after collection.

Figure 3 shows the effects of storage on INP observations in heat-treated precipitation samples. Non-heat-labile INPs represented the majority (6259% on average at -15 °C, see Sec. 3.1) of the total INPs observed in the fresh samples (i.e. 3841% of the INPs in fresh samples were heat-labile). Fewer significant losses of non-heat-labile INPs are observed for heat-treated samples stored at room temperature and at 4 °C compared with untreated samples. Again, slightly fewer (2-3) of the total frozen and flash frozen samples exhibit significant losses and enhancements. All observations other than the one significantly enhanced sample in (b) fall within ranges of stored:fresh ratios observed in the total insoluble particle population (see Fig. S7, within an order of magnitude). This demonstrates that non-heat-labile INPs are generally less sensitive to storage than the total INP population (Fig. 2).

Effects of storage protocol on INP concentrations of filtered precipitation samples are shown in Figure 4 (0.45 µm syringe filter, see Sect. 2.23 for details). INPs >< 0.45µm represented the majority (69% 52 and 63 % on average at -11 and -15 °C, respectively, see Sec. 3.1) of total INPs measured at the limit of detection in the fresh precipitation samples. A higher number of filter-treated samples exhibit significant losses across all 4 storage types when compared with the untreated samples. Furthermore, significant losses > 1 order of magnitude are observed across all storage types indicating that INPs < 0.45 µm are generally more sensitive to storage than the total INP population present in precipitation samples.

As the stored:fresh ratios follow a log-normal distribution (one-sample Kolmorgorov-Smirnov test), correction factorsthe uncertainties associated with storage and 95% confidence intervals were calculated in using the geometric mean and standard deviation of ratios of unique samples only between -9 and -17 °C (i.e. omitting any replicates, see Tables 5-7). and 95% confidence intervals for each storage protocol and sample type (untreated, heat-treated, filtered) were calculated using the geometric mean and standard deviation of ratios of unique samples only between -7 and -17 °C (i.e. omitting any replicates, see Tables 5-7). These correction factors can be applied to measurements from stored samples to estimate the concentrations of INPs in the fresh sample.

4. Discussion

The challenge in selecting a storage protocol for atmospheric samples (e.g. precipitation, cloud water, ambient atmosphere) is that the INP population composition is unknown, diverse, and the impact of any given technique on the different species may vary.The challenge in selecting a storage protocol for INPs collected in situ is that the population composition is unknown, diverse, and the impact of any given technique on the different species may vary. Many types of aerosols can serve as INPs, including dusts, metals and metal oxides, organic and glassy aerosols, bioaerosols, organic and mineral soil dust, and combustion products (Kanji et al., 2017). The aim of this study was to identify a storage protocol that best preserves the concentrations and characteristics of the general INP population observed in precipitation samples collected in a coastal environment. To this end, the impacts of 4 storage protocols on 15 untreated, heated, and filtered precipitation samples

collected between September 22, 2015 and November 22, 2019 in La Jolla, CA were investigated by comparing measured INP concentrations between fresh and stored replicates. The fractions of INPs > 0.45 µm observed in this study ~~INPs observed in this study were predominantly small, < 0.45 µm~~varied between 52 and 63% at -11 and -15 °C, respectively.~~, and insensitive to heat treatment~~ Excluding the five heat-treated samples in which INP concentrations were enhanced (e.g. 1.9 - 13X between -9 and -11 °C), the average fraction of non-heat-labile INPs varied between 40 and 62% at -11 and -15 °C, respectively. INP enhancements in heat-treated samples are unexpected, as heat-treatments are typically applied assuming that heat destroys proteinaceous (e.g. biological) INPs. The c~~.~~auses of INP enhancements in heat-treated samples are unknown and have only been reported in coastal precipitation samples (Martin et al., 2017) and nascent sea spray aerosol (McCluskey et al., 2018). Possible sources include the redistribution of dissolved IN-active molecules onto particles (McCluskey et al. 2018), and the release of IN-active content from cells (McCluskey et al. 2018, Wilson et al. 2015). These findings demonstrate that in samples influenced by marine sources, a superposition of both positive and negative ΔINP in samples could result in the observed changes in INP concentrations post heat-treatment.

Additionally, the INP freezing temperatures and concentrations observed in this study compare with INPs observed in studies of marine and coastal environments (Fig. 1). As spectra in this regime ~~(~~-5 to -20 °C and 10^{-5} to ~10^{-1} per L air, respectively) cluster distinctly by source type (see Fig. 1-10 in Kanji et al., 2017), Fig. 1 indicates that the dominant sources to ~~a~~air masses sampled in this study were marine~~.~~ Considering that data in this study compare well with marine and coastal INPs from a variety of marine-influenced air masses (DeMott et al., 2016, Yang et al., 2019), the findings herein are likely relevant to samples from other marine and coastal environments. ~~However, it is worth noting that the degree to which INP sensitivity to storage varies by site or INP source (e.g. with desert or soil-dominant INP populations) remains to be seen.~~

While mean INP changes are within a factor of ~2 or less of fresh sample INP concentrations for all protocols except "Room temperature" (Table 5), none of the 4 storage protocols prevented significant losses or enhancements of INP concentrations in all samples (Fig. 2), indicating that INP concentration measurements on fresh precipitation are superior to measurements on stored samples. 95% confidence intervals in Table 5 span losses > 1 order of magnitude in all protocols across multiple temperature intervals. These uncertainties equal or exceed INP measurement uncertainties (1-2 orders of magnitude) at temperatures > -20 °C due to discrepancies between instruments (DeMott et al., 2017). If correspondence within 1 order of magnitude (or 2-3 °C) is desired, uncertainties associated with storage should also be considered in studies using samples from coastal or marine environments. Thus, uncertainty distributions provided in Tables 5-7 can be used to evaluate observed INP concentrations and responses to treatments in the context of potential changes due to storage. However, the degree to which INP sensitivity to storage varies by INP source (e.g. with soil-derived INP populations) remains to be tested.

~~None of the 4 storage protocols prevented significant losses or enhancements of INP concentrations in all samples, demonstrating that INP concentration measurements on fresh precipitation are superior to measurements on stored samples. However, s~~Samples stored under freezing and flash freezing conditions exhibited fewer changes overall compared to refrigerated samples. For example, at the INP activation temperature of -13 °C, in the rain sample that exhibited the ~~greatest losses with~~highest sensitivity to storage, over ~~one-fifth~~20% of the original concentration was preserved in the frozen sample,

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whereas only ~~1/20th~~5% of the original concentration was preserved in the refrigerated sample. These losses are more extreme than those of (Stopelli et al., 2014b), which demonstrated that INP concentrations of a snow sample refrigerated over 30 days decreased only two-fold from 0.027 to 0.013 L^{-1} at -10 ºC.

Despite the range of enhancements and losses of heat-sensitive INPs observed in fresh samples, ~~N~~non-heat-labile INPs were generally less sensitive to storage than the total INP population, and with the exception of samples stored at room temperature, all techniques yielded similar results with fewer enhancements or losses. Interestingly, INPs < 0.45 µm exhibited more sensitivity to all storage conditions tested than the total INP population, with significant losses (Fishers Exact Test, $p < 0.01$) observed in several samples leaving between ~~one fourth~~25% and ~~1/30th~~3% of the value observed in the original fresh sample. Losses of INPs < 0.45 µm in samples stored at room temperature and +4 °C were comparable to the losses of total INPs in untreated samples and are likely a result of chemical aging in solution. However, losses of INPs < 0.45 micron in samples stored at -20 °C (both frozen and flash frozen) exceeded losses observed in the corresponding untreated samples. This is surprising given that ~~the majority~~a large fraction of INPs in this study were resilient to heat treatments of +95 °C. Lacking the identities of INPs observed in this study, a clear mechanism for their losses remains elusive. However, we offer the following points for consideration. It is well known that as a solution freezes, some solute is incorporated into the crystal and some is rejected, leading to enrichment of the solution phase and aggregation of dissolved or colloidal organic matter (Butler, 2002). Thus, as precipitation samples are freezing, small organic INPs may be lost simply due to aggregation in channels of enriched solute. In coastal precipitation samples for example, INPs may be so "lost" as the increased salinity in solution-phase channels destabilizes small suspended particles, allowing them to coagulate and settle (Jackson and Burd, 1998). Another possibility is that as the solution phase is enriched during freezing, smaller INPs may be adsorbing onto the surface of larger particles. The size distributions of total insoluble particles in the frozen samples show that most samples exhibit losses between 0-500 nm after storage and enhancements in sizes > 500 nm (see Fig. S6). This effect is not observed for samples stored at room temperature or at +4 °C.

Changes in the total insoluble particle size distribution (± 1 order of magnitude between 10 and 2000 nm, see Figs S6 and S7) may also have contributed to the observed INP concentration enhancements. Potential mechanisms for INP enhancements include increases in the number concentration of small particles due to breakup of loosely clumped masses of smaller particles, the redistribution of dissolved IN-active molecules onto particles (McCluskey et al. 2018), and the release of IN-active content from cells (McCluskey et al. 2018, Wilson et al. 2015) during cell death and lysis post freezing (Mazur et al., 1984).

Previous studies on precipitation collected along the California coast have demonstrated the contribution of dust, marine and terrestrial bioparticles to INPs in precipitation (Levin et al., 2019; Martin et al., 2019). Considering that well-characterized IN-active dust and biological standards (Arizona Test Dust and Snomax®, respectively) are sensitive to storage conditions, it is possible that dust or biological INPs contributed to the observed INP changes. Perkins et al. (2020) found that the IN-ability of Arizona Test Dust is degraded in most conditions, including aging in deionized water for 1 day, and results from Polen et al. (2016) show that the most efficient (i.e. warmest freezing) components of biological ice nucleators are also the most labile and sensitive to storage.

~~These results~~The observed distributions of INP concentration changes in stored precipitation samples ~~have~~ have implications for the interpretation of heat and filtration treatment experiments. As heat denatures proteins, heat treatments are commonly used to infer contributions of proteinaceous or cellular contributions to INP populations, and filters are commonly applied to identify observed INP size ranges (e.g. McCluskey et al., 2018). For example, a typical analysis involves a comparison of the INP spectrum of an untreated sample to that of the heat-treated or filtered sample, and information about the sizes and biological composition of INPs are derived from this comparison. Our results demonstrate that these treatments may yield different results if treatments are applied to stored samples. Any losses of INPs due to filtering or heat application could be confounded by significant enhancements or losses caused by storage (up to > 1 order of magnitude), resulting in inaccurate conclusions about INP characteristics. In this study, ~~the majority~~ a large fraction (~~30~~69% to 48%, on average) of INPs observed in fresh precipitation samples were < 0.45 µm. Considering this and that INPs < 0.45 µm exhibit significant losses across all storage types, there is a ~~substantial~~ risk that filter-treatments on stored samples in this study would lead to the ~~underestimation of a false conclusion: that~~ INPs < 0.45 µm. ~~the majority of INPs were > 0.45 µm.~~ Losses of heat-labile INPs in storage could also impact treatment outcomes on stored samples. Assuming negligible effects of storage on the heat-treated sample but losses due to storage in the untreated sample (e.g. as was shown to be most likely for untreated samples stored at +4 °C), INP spectra of heat-treated samples could appear to indicate the entire INP population was heat-insensitive. This effect was observed in several samples across storage types (see Fig. S8).

~~Previous studies on precipitation collected along the California coast have demonstrated the contribution of dust, marine and terrestrial bioparticles to INPs in precipitation (Levin et al., 2019; Martin et al., 2019). Considering that well-characterized IN-active dust and biological standards (Arizona Test Dust and Snomax®, respectively) are sensitive to storage conditions, it is possible that *in situ* dust or biological INPs contributed to the observed INP losses. Perkins et al. (2020) found that the IN-ability of Arizona Test Dust is significantly degraded in most conditions, including aging in deionized water for 1 day, and results from Polen et al. (2016) show that the most efficient (i.e. warmest freezing) components of biological ice nucleators are also the most labile and sensitive to storage.~~ ~~Changes in the total insoluble particle size distribution (see Figs S6 and S7) (± 1 order of magnitude between 10 and 2000 nm) could have additionally contributed to enhancements and losses. For example, the overall losses observed in INPs < 0.45 µm correspond to overall losses of the smallest insoluble particles (10-500 nm) in Fig. S6.~~

~~Although some ice nucleating species such as *Pseudomonas syringae* are known to survive unprotected freezing events (Buttner and Amy, 1989), it is worth noting that freezing is lethal for most cells (Mazur, 1984). The fact that cellular INPs are damaged by freezing may have motivated the choice of storage above 0 °C in some studies (see Table S1). Non-cellular biological INPs, however, may be conserved in frozen storage. Wright et al., (2013) showed that Snomax® maintains much of its IN-activity even through multiple freeze-thaw cycles, indicating the persistence of large aggregates of the IN-active protein. The limited available research also suggests that freezing will not kill most viruses (Smith et al., 2004), nor will it alter the tertiary structure of gels, vesicles, or cell-free proteins.~~

~~Changes in the total insoluble particle size distribution (see Figs S6 and S7) (± 1 order of magnitude between 10 and 2000 nm) could have additionally contributed to enhancements and losses. For example, the overall losses observed in INPs < 0.45 µm correspond to overall losses of the smallest insoluble particles (10-500 nm) in Fig. S6.~~

~~Though non-heat-labile INPs generally exhibit less sensitivity to storage than untreated samples, losses of heat-labile INPs in storage could impact treatment outcomes on stored samples. Assuming negligible effects of storage on the heat-treated sample but significant losses due to storage in the untreated sample (e.g. as was shown to be most likely for untreated samples stored at +4 °C), INP spectra of heat-treated samples could appear to indicate the entire INP population was heat-insensitive. This effect was observed in several samples across storage types (see Fig. S8).~~

~~Significant enhancements in INP concentrations occurred less frequently than losses. Again, changes in the total particle size distribution could explain some of the observed INP concentration enhancements. Increases in the number concentration of small particles due to breakup of loosely clumped masses of smaller particles could contribute to the increase in INPs. Other possible explanations include the redistribution of dissolved IN-active molecules onto particles (McCluskey et al. 2018), and the release of IN-active content from cells (McCluskey et al. 2018, Wilson et al. 2015) during cell death and lysis post freezing (Mazur et al., 1974). Although some ice-nucleating species such as *Pseudomonas syringae* are known to survive unprotected freezing events (Buttner and Amy, 1989), it is worth noting that freezing is lethal for most cells (Mazur, 1984). The fact that cellular INPs are damaged by freezing may have motivated the choice of storage above 0 °C in some studies (see Table S1). Non-cellular biological INPs, however, may be conserved in frozen storage. Wright et al., (2013) showed that Snomax® maintains much of its IN-activity even through multiple freeze-thaw cycles, indicating the persistence of large aggregates of the IN-active protein. The limited available research also suggests that freezing will not kill most viruses (Smith et al., 2004), nor will it alter the tertiary structure of gels, vesicles, or cell-free proteins.~~

5. Conclusions

Based on all observations in this study, we provide the following recommendations for precipitation samples collected ~~in~~ in ~~coastal~~ coastal and marine environments ~~environments~~ for offline INP analyses:

1. Of the 4 storage protocols tested, none prevented changes in INP concentrations across all samples between -7 and -19 ºC. However, whenever processing fresh samples is not possible, our results demonstrate that storage at -20 ºC causes the least changes in INP concentrations.
2. ~~Correction factors~~Estimates of uncertainty attributed to storage impacts and 95% confidence intervals for INP measurements obtained from stored samples are provided (see Tables 5-7).
3. Flash freezing with liquid nitrogen before storing at -20 ºC did not ~~significantly~~ improve conservation of INPs.
4. With the exception of warm-freezing INPs (freezing temperatures >= -9 °C) in samples stored at room temperature ~~or +4 °C~~, we found little to no correlation between changes in INP concentrations and storage intervals on timescales between 1-166 days, indicating that most enhancements or losses are likely happening during freezing or on timescales < 24 hours.

5. INPs that are insensitive to heat treatments are also less sensitive to storage. However, potential enhancements or losses due to storage (e.g. an average loss of 5025% for INPs with freezing temperatures >= -15 °C in samples stored at -20 °C) should be treated as additional uncertainty in measurements of INP concentration when comparing heat-treated with untreated INP spectra.
6. Due to the significant losses of INPs < 0.45 µm in storage, regardless of protocol, we recommend applying filtration treatments to fresh samples exclusively.

As measurements of INPs suspended in precipitation samples are used to infer in-cloud INP composition and concentration estimates, they represent important contributions to studies of links between aerosols, cloud processes and precipitation outcomes. This study derives bounds and correction factors for the impacts of storage on INPs and treatment outcomes from changes in INPs observed in coastal precipitation samples. However, it remains to be seen how INP sensitivity to storage varies by environment or INP composition. Further studies are needed to bracket storage effects on INP populations with various distributions of terrestrial and marine sources, as well as on heat-labile (biological) INPs, and INPs with colder activation temperatures. These studies could additionally benefit from analysis on how storage impacts differential INP spectra, which could reveal how sensitivity to storage varies by specific freezing temperature ranges. Bounds on the impact of storage will enable more meaningful intercomparisons of datasets and illuminate best practices for preserving INPs for offline analysis.

Data Availability: The data set supporting this manuscript is hosted by the UCSD Library Digital Collections (https://doi.org/10.6075/J0M32T8B).

Supplement Link: The supplement related to this article is available online at:

Author Contributions: CMB wrote the manuscript, prepared figures, led the field campaign and laboratory analysis. DL contributed to the preparation of figures, precipitation sample collection and laboratory analysis. MDS, TCH, PJD and KAP provided feedback on the analyses and manuscript. KAP and PJD are principal investigators on awards CHE-1801971 and AGS-1451347.

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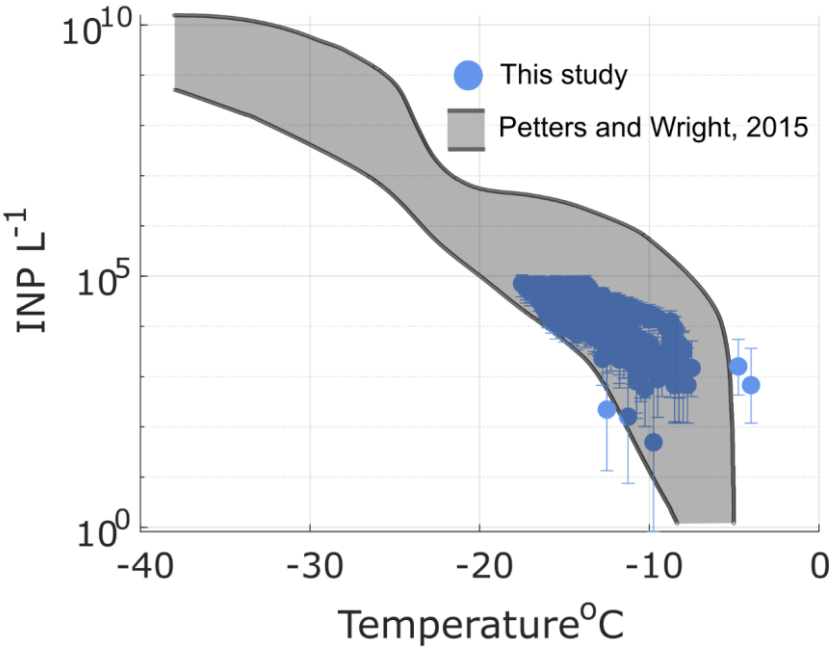
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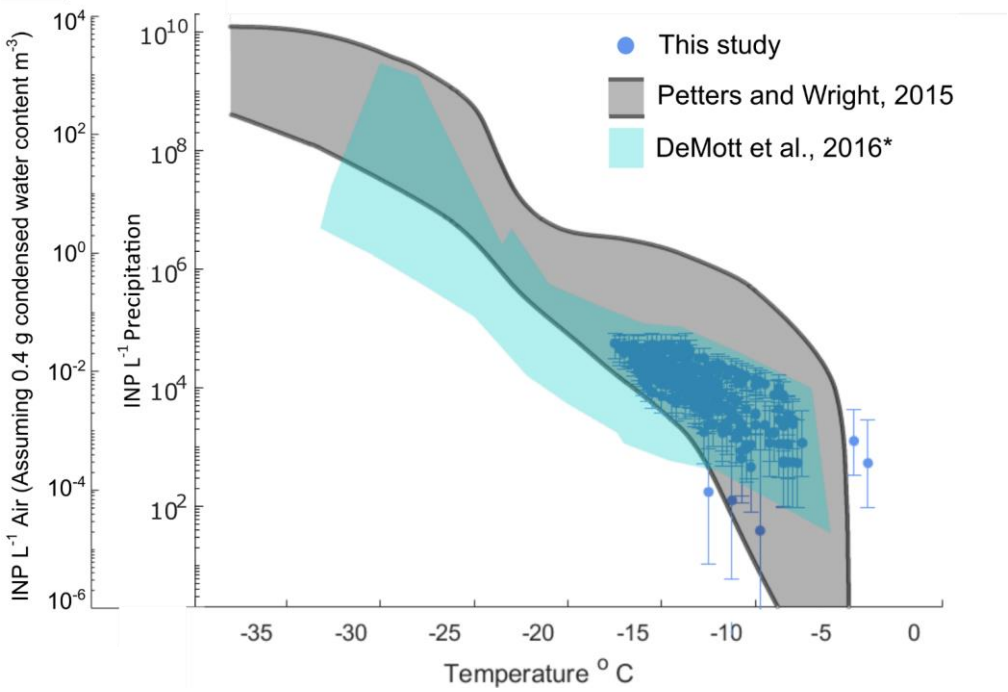
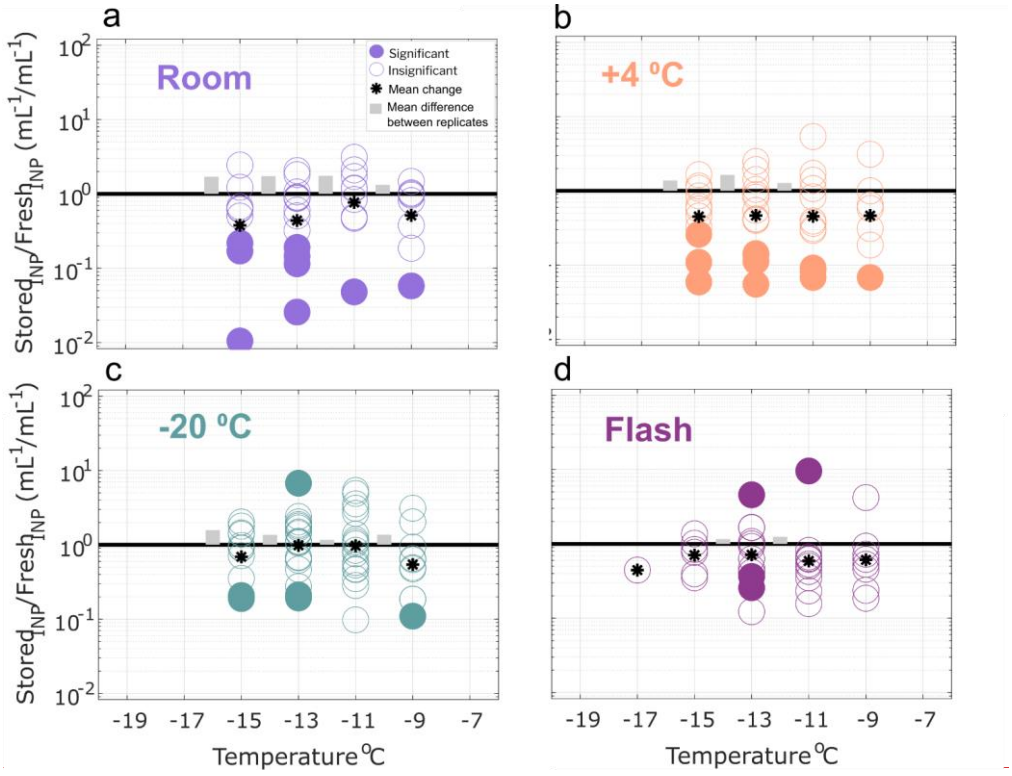


Figure 1: INP concentrations per liter of precipitation and estimated in-cloud INP concentrations per volume of air in 15 precipitation samples collected at two coastal sites at Scripps Institution of Oceanography (La Jolla, California, USA) between 9/22/2016 and 11/22/2019. Grey shaded region indicates the spectrum of INP concentrations reported in 9 previous studies of precipitation and cloud water samples collected from various seasons and locations worldwide, adapted from Fig. 1 in (Petters and Wright, 2015~~b~~). The blue shaded region denotes the composite spectrum of INP concentrations observed in a range of marine and coastal environments including the Caribbean, East Pacific and Bering Sea as well as laboratory-generated nascent sea spray (DeMott et al., 2016).

*DeMott et al., 2016 data has been updated with a completed dataset for the ICE-T study, as shown in Yang et al., 2020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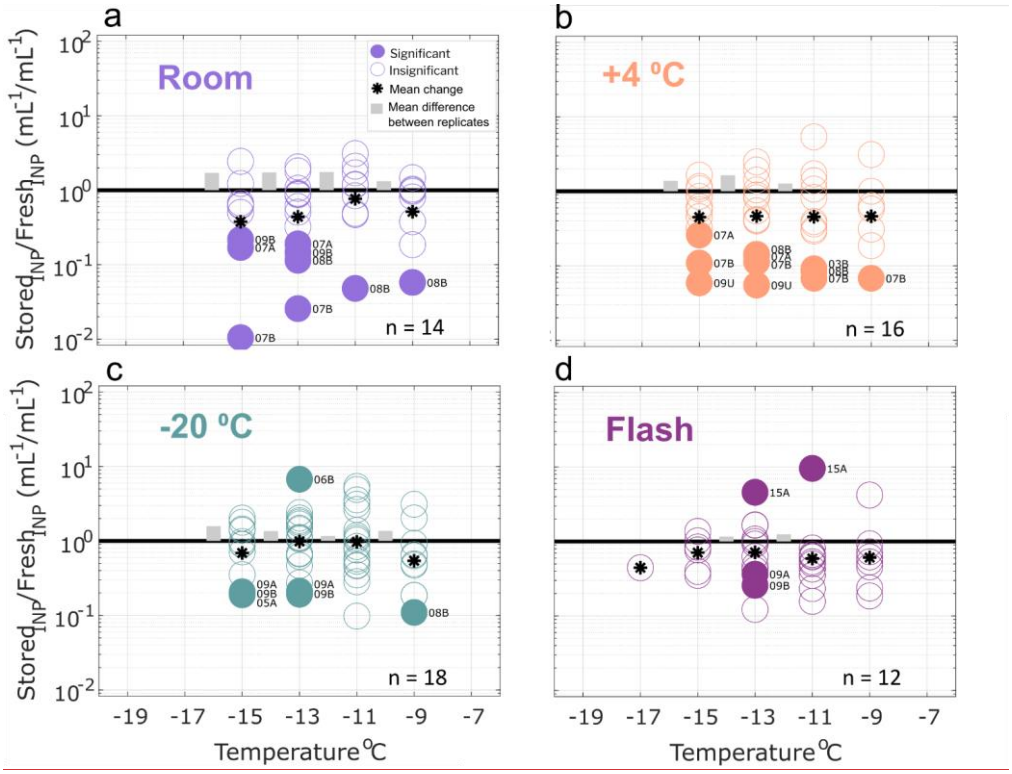

Figure 2: Ratio of INP concentrations measured in untreated precipitation samples (stored:fresh), calculated in~~binned by~~ successive 2 °C increments between -19 and -7 °C. Four storage protocols were applied: **(a)** room temperature (21-23 °C), **(b)** refrigerated (+4 °C), **(c)** frozen (-20 °C) and **(d)** flash freezing in liquid nitrogen before storing frozen (-20 °C). All samples were processed at one or two time intervals between 1 and 166 days post-collection (see Figs S1-S4). For samples processed at two intervals, both replicate samples are represented in the figure for a total of 14, 16, 18 and 12 samples in (a), (b), (c) and (d), respectively (see Table 2 for summary of sample and replicate numbers). ~~Eight unique samples are represented in the figure (9 in (c)), most of which were processed at two different time intervals between 1 and 166 days post-collection (see Table S1 and Figs S1-S4), and replicates are represented in the figure.~~ Markers above black 1:1 line indicate enhancement of INP concentrations in stored samples, and markers below indicate losses. In temperature ~~bins~~ intervals containing stored:fresh ratios from at least two sets of replicate samples, grey bars represent the average difference between replicates. Stored sample frozen well fractions that passed Fishers Exact Test ($p < 0.01$) for significant differences from original fresh sample frozen well fractions at each of the 5 temperatures are indicated with filled markers, and the mean change in each temperature ~~bin~~ interval is marked with a star. Significant data are also labelled to indicate the sample number (01-15, see Table 1), and replicate ("A" or "B", and "U" indicates there were no replicates for the sample). Results show that on average, INP concentrations decrease in stored samples, and that both room temperature storage and refrigeration result in significant INP losses. Frozen and flash frozen storage show comparable results, with fewer (3-4) of the observations exhibiting significant losses and enhancements in INP concentrations.

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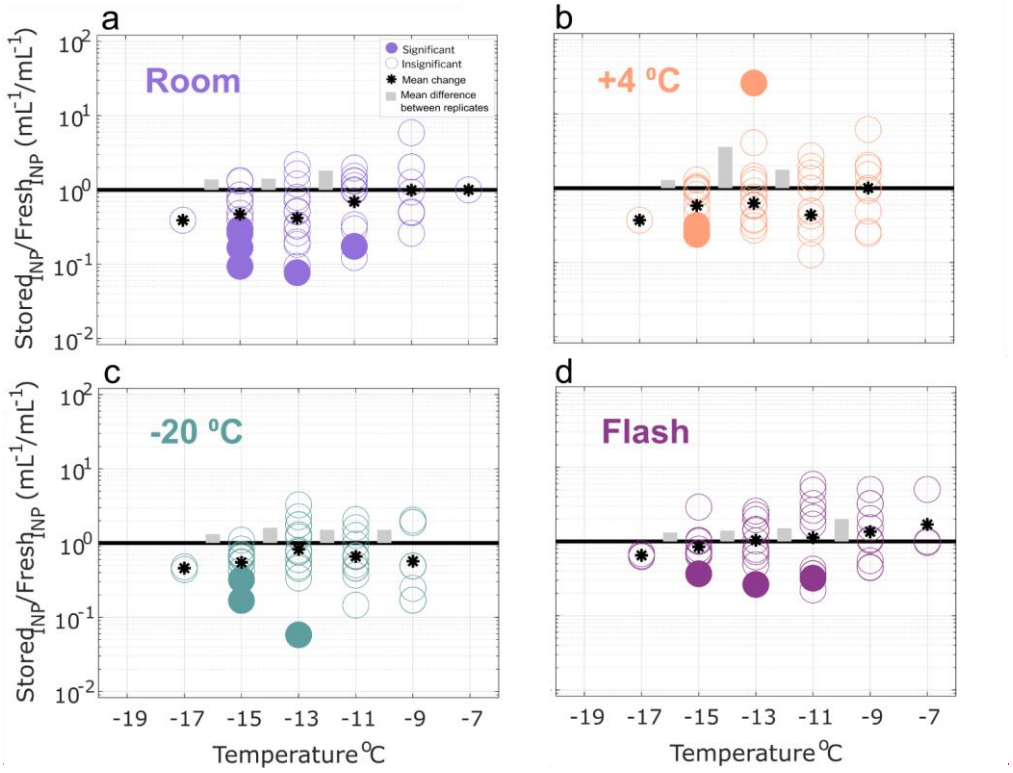
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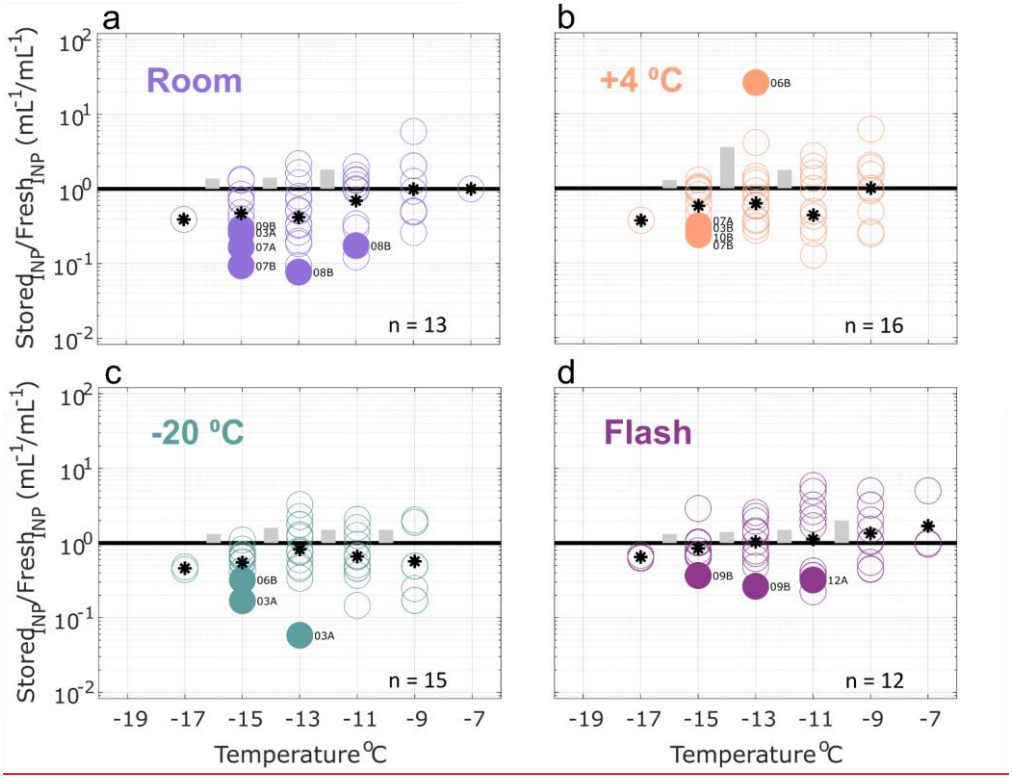

Figure 3: Ratio of INP concentrations measured in heated precipitation samples (stored:fresh), ~~binned by~~calculated in successive 2 °C increments between -19 and -7 °C. Same samples as shown in Figure 2, but heated to 95 °C for 20 minutes prior to measurement to eliminate heat-labile INPs ~~prior to measurement~~ (see Methods Sect. 2.2 for details). All samples were processed at one or two time intervals between 1 and 166 days post-collection. For samples processed at two intervals, both replicate samples are represented in the figure for a total of 13, 16, 15 and 12 samples in (a), (b), (c) and (d), respectively (see Table 3 for summary of sample and replicate numbers). ~~Eight unique samples are represented in the figures, most of which were processed at two different time intervals between 1 and 166 days post-collection (see Table S2), and replicates are represented in the figure.~~ In temperature ~~bins~~ intervals containing stored:fresh ratios from at least two sets of replicate samples, grey bars represent the average difference between replicates. Results show significant losses of INPs in heat-treated samples stored at room temperature. Refrigerated, frozen, and flash frozen samples show comparable results with a few (1-3) samples exhibiting significant losses and enhancements. Non-heat-labile INPs are generally less sensitive to storage protocol than the total INP population in precipitation samples (Fig. 2), with the exception of storage at room temperature.

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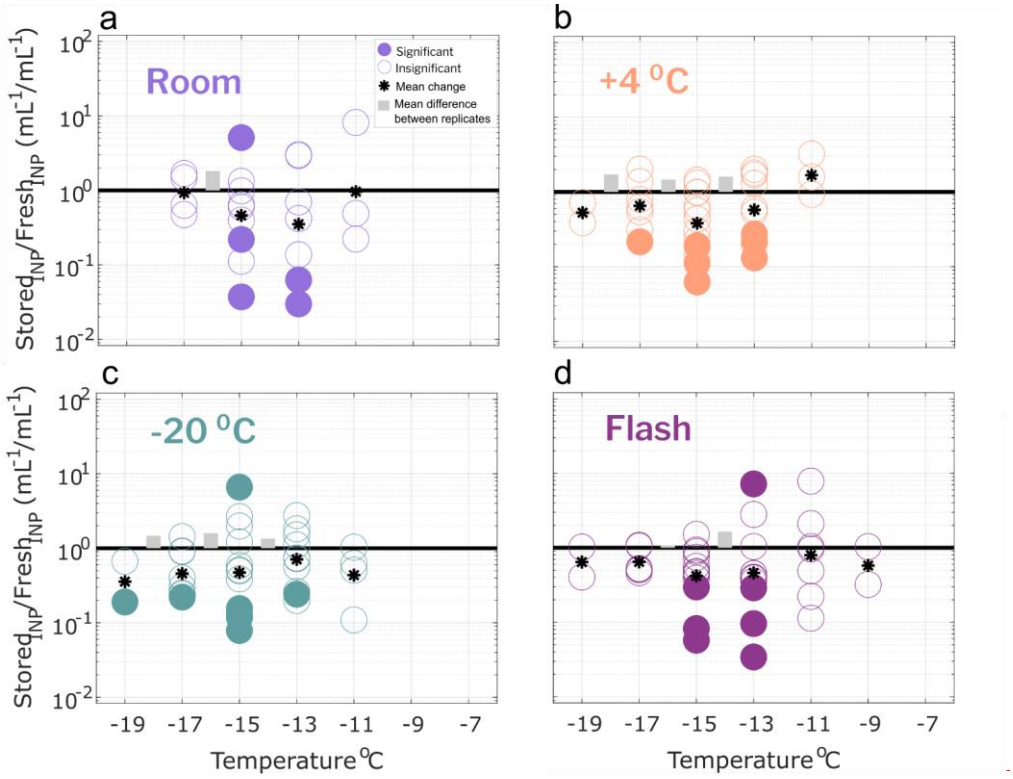
a
b
c
d
Room
+4 °C
-20 °C
Flash
Significant
Insignificant
Mean change
Mean difference between replicates
Stored$_{INP}$/Fresh$_{INP}$ (mL^{-1}/mL^{-1})
10^{2}
10^{1}
10^{0}
10^{-1}
10^{-2}
-19 -17 -15 -13 -11 -9 -7
Temperature °C

646

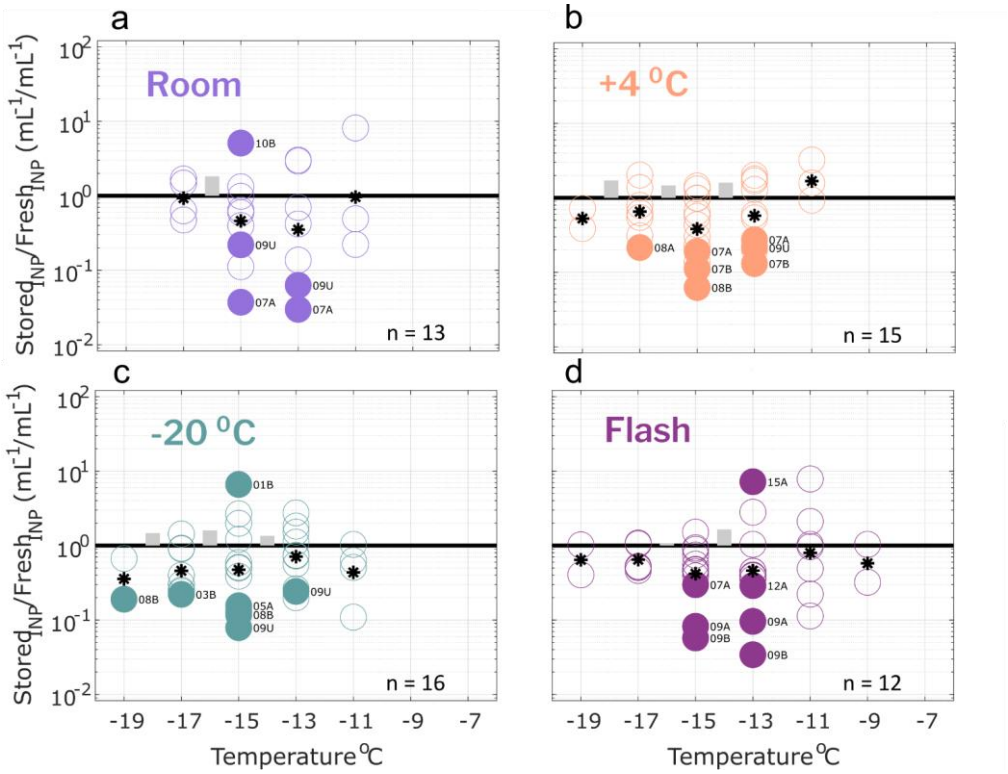

Figure 4: Ratio of INP concentrations measured in filtered (0.45 µm) precipitation samples (stored:fresh), ~~binned~~ calculated in successive ~~by~~ 2 °C increments between -19 and -7 °C. Same samples as in Fig. 2 but filtered with a 0.45 µm syringe filter prior to measurement (see Methods Sect. 2.2 for details). All samples were processed at one or two time intervals between 1 and 166 days post-collection. For samples processed at two intervals, both replicate samples are represented in the figure for a total of 13, 15, 16 and 12 samples in (a), (b), (c) and (d), respectively (see Table 4 for summary of sample and replicate numbers). ~~Eight unique samples are represented in the figures (9 in (c)), most of which were processed at two different time intervals between 1 and 166 days post-collection (see Table S3), and replicates are represented in the figure.~~ In temperature ~~bins~~ intervals containing stored:fresh ratios from at least two sets of replicate samples, grey bars represent the average difference between replicates. Results show significant losses of INPs in several filtered samples, regardless of storage protocol.

Table 1. Precipitation sampling periods

Sampling Period	UTC Date	UTC time start	UTC time end	Meteorological Conditions
1	9/22/2016	19:20	21:13	scattered, low coastal clouds, lack of dynamical system
2	9/22/2016	19:42	21:13	scattered, low coastal clouds, lack of dynamical system
3	12/31/2016	4:53	7:52	warm, low cloud rain
4	1/1/2017	7:53	10:52	post-frontal rain, meso-scale system
5	1/5/2017	21:02	22:01	pre-frontal rain, meso-scale system
6	1/9/2017	15:51	19:50	decaying atmospheric river
7	1/11/2017	19:00	23:30	frontal rain
8	1/14/2017	2:03	6:00	warm, low cloud rain
9	1/19/2017	12:30	17:30	pre-frontal rain, meso-scale system
10	1/20/2017	14:15	02:20 (next day)	weak atmospheric river
11	11/19/2019	22:34	22:45	pre-frontal rain, meso-scale system
12	11/22/2019	4:43	5:42	scattered, low coastal clouds, lack of dynamical system
13	11/22/2019	6:43	7:42	scattered, low coastal clouds, lack of dynamical system
14	11/23/2019	7:42	8:41	convective, local updraft rain
15	11/23/2019	8:42	9:41	convective, local updraft rain

Table 2. Summary of unique and replicate untreated precipitation samples used for INP concentration measurements featured in Fig. 2.

Storage technique	No. of unique samples	No. of stored samples measured at 2 timesteps
Room temperature (19 - 23 ºC)	8	6
Refrigeration (+4 ºC)	8	8
Freezing (-20 ºC)	9	9
Flash freezing (-20 ºC)	8	4

Table 3. Summary of unique and replicate heat-treated precipitation samples used for INP concentration measurements featured in Fig. 3.

Storage technique	No. of unique samples	No. of stored samples measured at 2 timesteps
Room temperature (19 - 23 ºC)	8	6
Refrigeration (+4 ºC)	8	8
Freezing (-20 ºC)	8	7
Flash freezing (-20 ºC)	8	4

Table 4. Summary of unique and replicate filtered (0.45 µm) precipitation samples used for INP concentration measurements featured in Fig. 4.

Storage technique	No. of unique samples	No. of stored samples measured at 2 timesteps
Room temperature (19 - 23 ºC)	8	5
Refrigeration (+4 ºC)	8	7
Freezing (-20 ºC)	9	7
Flash freezing (-20 ºC)	8	4

Table 5. ~~Correction factors~~Estimate of uncertainty associated with storage impacts **for INPs with activation temperatures between -~~9~~7 and -1~~7~~5 °C measured in stored, untreated precipitation samples.** ~~Correction~~, Confidence intervals were derived from the log-normal distribution of changes observed in INP concentrations due to storage (see Fig. 2 and details in Sect. 3.2). Temperature intervals where datapoints were too few to derive confidence intervals are indicated with "NA". Changes in INP concentration corresponding to enhancements or losses greater than 1 order of magnitude (losses <= -90% or enhancements >= +900%) in bold.~~factors and confidence intervals were derived from the log-normal distribution of changes observed in INP concentrations due to storage (see Fig. 2 and details in Sect. 3.2).~~

~~Storage protocol~~	~~Correction factor~~	~~95% CI Lower Limit~~	~~95% CI Upper Limit~~
~~Room temperature (21 - 23 ºC)~~	~~x3.59~~	~~0.19~~	~~69.29~~
~~Refrigeration (+4 ºC)~~	~~x1.72~~	~~0.25~~	~~11.27~~
~~Freezing (-20 ºC)~~	~~x1.34~~	~~0.22~~	~~8.34~~
~~Flash freezing (-20 ºC)~~	~~x1.48~~	~~0.22~~	~~9.88~~

Storage protocol	Mean Change -9 °C (%)	95% CI Low (%)	95% CI High (%)	Mean Change -11 °C (%)	95% CI Low (%)	95% CI High (%)	Mean Change -13 °C (%)	95% CI Low (%)	95% CI High (%)	Mean Change -15 °C (%)	95% CI Low (%)	95% Hig (%)
Room temperature (21 - 23 ºC)*	-26	-82	+200	-51	**-97**	+850	-77	**-98**	+220	-77	**-99**	**+10**
Refrigeration (+4 ºC)*	-42	-74	+32	-74	**-99**	+400	-46	**-95**	+520	-56	**-95**	+29
Freezing (-20 ºC)	-48	**-95**	+430	-16	**-90**	+580	+24	-80	+650	-50	**-90**	+15
Flash freezing (-20 ºC)	-21	**-90**	+520	-41	**-95**	+560	-33	**-91**	+390	NA	NA	NA

* For INPs with freezing temperatures >= -9 °C, changes in INP concentrations are moderately correlated with time in samples stored at room temperature or at +4 °C (see Sec. 3.2). Change factors for room temperature and refrigerated storage protocols are derived from samples stored in ranges of 27 – 76 and 8 – 46 days, respectively.

Table 6. Estimate of uncertainty associated with storage impacts for INPs with activation temperatures between -9 and -17 °C measured in stored, heat-treated precipitation samples. Confidence intervals were derived from the log-normal distribution of changes observed in INP concentrations due to storage (see Fig. 3 and details in Sect. 3.2). Changes in INP concentration corresponding to enhancements or losses greater than 1 order of magnitude (losses <= -90% or enhancements >= +900%) in bold.~~**Correction factors for INPs with activation temperatures between -7 and -15 °C measured in stored, heat-treated precipitation samples.** Correction factors and confidence intervals were derived from the log-normal distribution of changes observed in INP concentrations due to storage (see Fig. 3 and details in Sect. 3.2).~~

Storage protocol	Correction factor	95% CI Lower Limit	95% CI Upper Limit
Room temperature (21 - 23 ºC)	x1.41	0.23	8.6
Refrigeration (+4 ºC)	x1.24	0.22	7.04
Freezing (-20 ºC)	x1.05	0.25	4.41
Flash freezing (-20 ºC)	x0.93	0.19	4.4

Storage protocol	Mean Change -9 °C (%)	95% CI Low (%)	95% CI High (%)	Mean Change -11 °C (%)	95% CI Low (%)	95% CI High (%)	Mean Change -13 °C (%)	95% CI Low (%)	95% CI High (%)	Mean Change -15 °C (%)	95% CI Low (%)	95% CI High (%)
Room temperature (21 - 23 ºC)*	+32	-74	+550	-17	-86	+380	-65	**-95**	+155	-58	**-93**	+15
Refrigeration (+4 ºC)*	-5.6	**-91**	**+940**	-74	**-99**	**+1600**	-58	**-99**	**+6000**	-60	-87	+2
Freezing (-20 ºC)	-55	**-91**	+130	-53	-87	+69	-42	**-93**	+390	-34	-70	+4
Flash freezing (-20 ºC)	+36	-76	+660	+31	-88	**+1300**	-9.0	-81	+340	+1.0	-60	+15

Table 7. **Estimate of uncertainty associated with storage impacts for INPs with activation temperatures between -11 and -19 °C measured in stored, filtered precipitation samples.** Confidence intervals were derived from the log-normal distribution of changes observed in INP concentrations due to storage (see Fig. 2 and details in Sect. 3.2). Temperature intervals where datapoints were too few to derive confidence intervals are indicated with "NA". Changes in INP concentration corresponding to enhancements or losses greater than 1 order of magnitude (losses <= -90% or enhancements >= +900%) in bold. **Correction factors for INPs < 0.45 µm with activation temperatures between -9 and -17 °C measured in stored precipitation samples.** Correction factors and confidence intervals were derived from the log-normal distribution of changes observed in INP concentrations due to storage (see Fig. 4 and details in Sect. 3.2).

Storage protocol	Correction factor	95% CI Lower Limit	95% CI Upper Limit
Room temperature (21 - 23 ºC)	x2.23	0.15	32.36
Refrigeration (+4 ºC)	x2.37	0.29	19.24
Freezing (-20 ºC)	x1.54	0.19	12.48
Flash freezing (-20 ºC)	x1.82	0.32	10.31

Storage protocol	Mean Change -11 °C (%)	95% CI Low (%)	95% CI High (%)	Mean Change -13 °C (%)	95% CI Low (%)	95% CI High (%)	Mean Change -15 °C (%)	95% CI Low (%)	95% CI High (%)	Mean Change -17 °C (%)	95% CI Low (%)	95% CI High (%)
Room temperature (21 - 23 ºC)	NA	NA	NA	-80	**-99**	+360	-72	**-96**	+130	-7.0	-68	+170
Refrigeration (+4 ºC)	NA	NA	NA	-48	**-94**	+300	-65	**-97**	+250	-14	-80	+250
Freezing (-20 ºC)	NA	NA	NA	-31	-89	+330	-54	**-98**	+870	-32	-78	+110
Flash freezing (-20 ºC)	-26	-83	+230	-65	**-98**	+650	-68	**-96**	+140	NA	NA	NA