



Atomic emission detector with gas chromatographic separation and cryogenic pre-concentration (CryoTrap-GC-AED) for atmospheric trace gas measurements

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Abstract. A gas detection system has been developed, characterized and deployed for pressurized gas phase sample analyses and near real-time online measurements. It consists of a cryogenic pre-concentrator (CryoTrap), a gas chromatograph (GC), and a new high-resolution atomic emission detector (AED III). Here the CryoTrap–GC–AED instrumental setup is presented and the performance for iodine (1635 ± 135 counts I-atom⁻¹ pptv⁻¹), sulfur (409 ± 57 counts S-atom⁻¹ pptv⁻¹), carbon (636 ± 69 counts C-atom⁻¹ pptv⁻¹), bromine (9.1 ± 1.8 counts Br-atom⁻¹ pptv⁻¹) and nitrogen (28 ± 2 counts N-atom⁻¹ pptv⁻¹) emission lines is reported and discussed. The limits of detection (LODs) are in the low pptv range (0.5 – 9.7 pptv) and the signal is linear to at least 4 orders of magnitude, which makes it a suitable method for diverse volatile organic compound (VOC) measurements in the atmosphere, even in remote, unpolluted regions. The new system was utilized in a field study in a boreal forest at Hyytiälä, Finland in late summer 2016 which made monoterpene measurements possible among the other VOCs. Furthermore, pressurized global whole-air sample measurement collected onboard the Lufthansa Airbus A340-600 IAGOS-CARIBIC aircraft in the upper troposphere and lower stratosphere region was carried out with the new setup, providing the observational data of many VOCs, including the long-lived organosulfur compound carbonyl sulfide.

1 Introduction

Atomic spectrometric analysis provides highly sensitive, equimolar, linear response at least over 5 orders of magnitude ($> 10^5$) and accurate elemental composition data of samples. If detection is preceded by analyte separation using gas chromatography (GC), compound specific data is obtainable. The first atomic emission detector (AED) with a plasma as an excitation source coupled with a GC was introduced in 1965 (McCormack et al., 1965). This group was the first to recognize the analytic potential of combining a GC separation with microwave induced plasma excitation and an electronic emission spectra detector. Early AEDs, using microwave induced plasmas, were operated at reduced pressures (Risby et al., 1983), until an atmospheric pressure version was developed in 1977 (Beenaker, 1977). The first commercial AED based on a microwave induced plasma and photodiode array detector coupled to a GC was released by Hewlett Packard in 1990 (Quimby and Sullivan, 1990). These systems provided speciated element specific chromatograms and were used in a variety of applications including the analysis of oils for sulfur containing compounds (Link et al., 2002). In contrast with the alternative analytical approach of gas



30 chromatography coupled to mass spectrometry (GC–MS), the AED has the advantage of being equimolar and linear in detector
response. That said it is important to keep in mind that a detector is often the last step in the whole analytical instrumental
setup, therefore the response factor (RF) reflects the entire analyte pathway from the sample inlet up to the detector signal
recording. This means that analyte losses by adsorption and absorption effects taking place in the transfer lines, pre-
concentration stages, GC column and in the detector flow paths etc. contribute to the final RF. In this study, great care was
35 taken to minimize the loss factors. Hence, the RF for a particular compound group can be derived for one element (e.g. carbon)
from the similar single compound specific RFs and then applied to all carbon containing compounds within the group in the
chromatogram. This greatly simplifies the calibration and allows quantification of compounds not present in a calibration
standard. For this reason, the AED has been used for the quantification of species that have been identified by GC-MS (Apel
et al., 1998; Greenberg et al., 1999). Recently, further technical developments in the AED have led to improvements in
40 sensitivity that allow trace gases in air to be quantified directly, making such systems of interest to atmospheric scientists. The
simultaneous high-resolution wavelength recording capability of the detector (161 – 211 nm) in the CryoTrap–GC–AED
system allows for reliable screening for elements present in the speciated compounds, thus simplifying the identification of
unknown compounds. Furthermore, the newly developed system in principle allows for unknown compound quantification in
a chromatogram within the uncertainty range when a broad range of compound specific RFs are used for the determination of
45 the single element RF.

In this study, the instrumental setup and performance of the CryoTrap–GC–AED system are examined. The performance for
iodine, sulfur, carbon, bromine and nitrogen emission lines is examined. The calibration linearities, limits of detection and
compound specific response factors are reported for 64 compounds.

2 Experimental

50 The CryoTrap–GC–AED system consists of three stages: a liquid nitrogen based pre-concentration system (Entech model
7200, USA); a gas chromatographic separation (Agilent GC 7890B, USA); and a helium plasma based third generation atomic
emission detector (Joint Analytical Systems AED III High Resolution, Germany). The schematic of the instrumental setup is
shown in Fig. 1. Ultra-high purity helium (UHP, purity 99.9999%, Westfalen, Germany) flowing through a heated purifying
catalyst (Valco Instruments VICI, USA) is used throughout the system as the carrier and purging gas.

55 2.1 Cryogenic pre-concentration (CryoTrap)

The sample is introduced to the pre-concentration unit (CryoTrap) via an eight port multi position valve, consisting of the
helium supply gas, four sample introduction inlets, an internal standard, a calibration standard and a blind port (Fig. 1, upper
panel). The four sample introduction lines are each 2.0 m long (Restek Corp. Silcosteel, USA) with outer diameter 1/16" (1.59
mm), inner diameter 0.040" (1.02 mm). The sample is drawn onto the two enrichment traps via an evacuated volumetric
60 reservoir, where the sample introduction volume is accurately determined by measuring the pressure at given temperature. All



the CryoTrap internal flow path surfaces are coated with a thin high density ceramic Silonite-D layer to provide extremely inert surfaces, which nearly eliminates the adsorption of the analytes to the surfaces.

The CryoTrap internal flow path is flushed with the sample gas before each pre-concentration step. After that the sample air is drawn through the first stage of pre-concentration, called the dehydration module (Fig. 1, middle panel), where H₂O is selectively removed on an empty Silonite-D coated stainless-steel trap (outer diameter 1/8" (3.18 mm), 31 cm long) held at -50°C. After collecting the required volume of sample, the trap is flushed with 75.0 mL of helium to remove any remaining air. Then module 1 is heated to 10°C and forward purged with 50.0 mL (10 mL min⁻¹) helium flow onto the main Tenax packed and Silonite-D coated volatile organic compound (VOC) enrichment trap (module 2, the second step of pre-concentration; outer diameter 1/8" (1.02 mm), 31 cm long) held at -60°C. The forward helium purge helps to successfully transfer heavy, polar and semi-volatile organic compounds (SVOCs) onto the main module 2 VOC trap. The last pre-concentration step is pre-cooling of the cryo-focusing trap (module 3, ~4 cm long part of Silonite-D coated 1/32" (0.79 mm) transfer line) to -180°C and thereafter kept at -160°C during module 2 back flushing at 230°C for 2.5 min, which will refocus the volatiles to a much smaller dead volume for splitless injection onto a GC column. The module 3 1/32" (0.79 mm) transfer line is rapidly heated to 60°C for 2 min inside a sheathing 1/8" (3.18 mm) perfluoroalkoxy alkane (PFA) tube with hot air supplied through the rotary plate kept at 100°C. This ensures rapid and splitless injection of analyte molecules to the GC column through a transfer line heated to 110°C. The pre-concentration unit is equipped with two bulkhead heaters in between the heated rotary plate and the traps for better water condensation management.

2.2 Gas chromatograph (GC)

Gas chromatography is a common choice for analytical separation of VOCs for measurement with various detectors (Bourtsoukidis et al., 2017; Apel et al., 1998). We use an Agilent 7890B GC for the compound separation. The GC was fitted with a Supelco SPB-624 capillary column (length × inner diameter: 60 m × 250 μm; film thickness 1.40 μm), which is an intermediate polar, proprietary phase bonded fused silica GC column. The SPB-624 type columns are widely used for volatile non-halogenated, halogenated, and aromatic compounds analysis.

Helium is used as the column carrier gas, of which the flow rate is controlled over the GC electronic pneumatic control (EPC) valve number 3. First, helium flows from the GC EPC valve into a 1/16" (1.59 mm) stainless steel line connected to the CryoTrap rotary valve number 2 (Fig. 1). After the pre-concentration procedure the helium flow with the remobilized analytes is guided back to the GC oven through a heated (110°C) Silonite-D coated 1/32" (0.79 mm) transferline. Then, in the GC oven a Swagelok T-split union guides the flow to the analytical column for splitless injection. The other end of the GC capillary column was led directly to the AED cavity through a heated transferline (250°C).



90 **2.3 Atomic emission detector, third generation (AED III)**

AED measures the energy emitted at characteristic wavelengths by sample atoms present in the helium plasma cavity to quantify their number in a chromatographic peak. Combining this data with GC analyte separation, the amount of the substance can be quantitatively determined.

The helium carrier gas (3.5 mL min^{-1}) eluting from the GC is led to the AED cavity through the capillary column that is housed
95 in the heated transfer line (250°C). The helium plasma discharge cavity is also kept at constant 250°C temperature. The mixture of the carrier (helium) and reagent gases (hydrogen and oxygen) flow through a fused silica discharge tube where the gases are ionized into a plasma state by microwave energy. In the high-energy plasma the eluted sample compounds from the GC are broken down into free radicals, ions and atoms. As they return from their excited state to ground state configuration, light radiation is emitted in their element specific characteristic wavelengths. The emitted ultraviolet radiation passes through a
100 fused silica lens and a narrow slit, and then is reflected onto a reflective holographic grating by a fixed mirror. The grating disperses the light into discrete vertical bandwidths along a plane-concave polychromatic grating. Thereafter, the grating reflects and focuses the light in the wavelength range 161 – 211 nm in a horizontal plane onto two back-thinned charge-coupled devices (CCDs). The CCDs convert light intensities across the given spectrum into electrical currents which are recorded by the “AED III Instrument Control Software”. The software calibrates the received light intensity signal from the CCD diodes
105 into wavelengths intensity data according to a calibration table. This process (element installation) takes place automatically before every sample measurement. After each measurement, all the wavelength dependent emission data are stored with 0.01 nm resolution. An example of a 5 element simultaneously recorded chromatogram of the 84 component Apel-Riemer-2015 gas calibration standard is shown in Fig. 2.

2.4 Characterization experiments

110 All the characterization experiments started with a zero-air (hydrocarbon free synthetic air with 20.5% oxygen and nitrogen rest (Westfalen, Germany), which flows through an extra catalyst kept at 500°C to remove the trace amounts of hydrocarbons) measurement for determining the instrumental background. Furthermore, at the beginning of the experimental design the highest calibration standard levels’ carryover and retention potentials of all the compounds of interest were tested with the zero-air measurement directly after as the following sample. The trap back-flushing and bakeout times were adapted
115 accordingly.

Three independent gas calibration standards were used for the characterization experiments: the 84 component (each nominally at 50 ppbv mixing ratio (MR) in UHP nitrogen rest) Apel-Riemer-2015 gas calibration standard (Table S1) and the 30 component (each nominally at 4 ppbv MR in UHP nitrogen rest) non-methane hydrocarbon (NMHC) ozone precursor National Physical Laboratory 2017 (NPL-2017) primary calibration standard gas cylinders (Table S2).

120 The Apel-Riemer-2015 standard used for the characterization measurements was diluted with the same zero-air as described above with two mass flow controllers (MFC). The MFC for the calibration gas had a range up to 20 mL min^{-1} and the MFC



for zero-air a range up to 5 L min⁻¹. For these experiments a constant 1 L min⁻¹ zero-air flow was used and five steps (1, 5, 10, 15, 20 mL min⁻¹) of calibration gas were used, to produce nominal MRs of 50, 250, 500, 750 and 1000 pptv. This MR range is relevant for many ambient VOC measurements. With each sample pre-concentration a 1400 ml aliquot of the diluted sample flow was trapped and the rest of the flow was led to lab exhaust. Calibration level measurements were repeated four times. No dilution for the NPL primary calibration standard was used and thus trapped directly in the amounts of 40, 60, 80 and 250 mL (producing MR in the range of 114, 171, 229 and 714 pptv for 1.4 L unknown sample relevance). The NPL calibration step measurements were also repeated four times.

With separate tests the maximum linearity range of the CryoTrap-GC-AED was tested with the undiluted Apel-Riemer-2015 calibration gas measurement (~ 50 ppbv per compound). For higher MR samples also a smaller sample volume can be trapped, making it possible to measure even higher MR samples. The system was tested to be linear to at least 4 orders of magnitude.

3 Instrument performance

The AED system was always recalibrated immediately prior to and during the measurement of the samples with unknown MRs, using the independent gas calibration standards: the Apel-Riemer-2015 and the NPL-2017 gas cylinders. In addition, during the regular automated IAGOS-CARIBIC pressurized sample measurement procedure, the National Oceanic and Atmospheric Administration (NOAA) ambient air calibration standard and the NMHC NPL standards were periodically measured after every five samples. From the calibration standard measurements, the linearities (R-squared), limits of detection (LOD), response factors per atom and measurement uncertainty were determined for all quantifiable compounds in the standard and listed in Table 1 and 2.

The LODs were calculated from the average height of the lowest calibration levels and the average height of the noise signals corresponding to the same chromatogram for each separable and measurable compound of the calibration standards. Three times signal-to-noise ratio (by peak height) was maintained to define the LOD, representing the ~99.7% confidence interval. The LOD calculation is represented by Eq. (1):

$$LOD = 3 \times \frac{\overline{noise} \times \overline{MR}}{\overline{peak\ height}}, \quad (1)$$

where \overline{noise} is the average height of the noise signal, \overline{MR} is the average mixing ratio of the lowest level calibration level and $\overline{peak\ height}$ is the average peak height of that calibration level of the corresponding compound.

The average per carbon atom RFs were calculated separately for each species using the average area under the peak curve (\overline{AUC}), the corresponding average MRs (\overline{MR}) and the number of carbon atoms (NOC) present in the compound of interest as show by Eq. (2):

$$\overline{RF} = \frac{\overline{AUC}}{\overline{MR} \times \text{NOC}}. \quad (2)$$



The RFs were calculated separately for all compounds and their individual calibration MR levels and thereafter all the individual RFs over the whole calibration range were averaged to a single RF per compound.

3.1 Carbon emission line performance

The performance of the carbon emission line (193 nm) derived from measurements of three calibration standard gases is listed in Table 1. For Apel-Riemer-2015, the derived LODs are in the range 0.8 – 9.7 pptv. Each compound's RF is the average of 18 independent data points in the whole 50 – 1000 pptv calibration range. The RFs for these compounds lie between 343 ± 147 counts C-atom⁻¹ pptv⁻¹ (benzyl chloride) to 800 ± 291 counts C-atom⁻¹ pptv⁻¹ (isobutyl nitrate) and on average 587 ± 91 counts C-atom⁻¹ pptv⁻¹ (47 compound mean). For NPL-2017, the LODs are in a smaller range of 0.5 – 1.1 pptv (more similar compounds) and the RFs per carbon atom range from 581 ± 23 counts C-atom⁻¹ pptv⁻¹ (1,2,4-trimethylbenzene) to 913 ± 77 counts C-atom⁻¹ pptv⁻¹ (toluene). The measured 22 compound NPL-2017 gas standard average RF was 684 ± 46 counts C-atom⁻¹ pptv⁻¹ (about 17% higher compared to Apel-Riemer-2015 measurements and with about half the standard deviation). The both experiment average per carbon atom RF was determined 636 ± 69 counts C-atom⁻¹ pptv⁻¹.

The CryoTrap-GC-AED system sensitivity and precision are also dependent on the discharge tube, as the two experiments were conducted with different discharge tubes (as the old one broke). Nevertheless, unknown samples were always directly determined against calibration standards with the same discharge tube. If a discharge tube breakage took place, a complete new calibration was carried out. The same 10 compounds (isobutene, 1,3-butadiene, pentane, isoprene, hexane, benzene, toluene, ethylbenzene, 1,3,5-trimethylbenzene and 1,2,4-trimethylbenzene) which were measured from both calibration standards are compared in Fig. 3. Three compounds: pentane, ethylbenzene and especially toluene do not adhere to the 1 σ confidence interval on the one to one RF comparison between these two experiments. The reason remains unknown as there are other similar compounds to these in the comparison which stick to the correlation.

Like the RFs, also the compound specific LODs depend on the CryoTrap pre-concentration efficiency, compound transmission efficiency through the gas transfer lines, GC separation and carry through efficiency and AED helium plasma atomization and excitation efficiency. Throughout the whole instrument development process, best efforts were undertaken to use suitable transfer lines and gas union (Swagelok, USA) materials like Silcosteel (fused silica coated stainless steel), stainless steel, PFA and Teflon for minimizing analyte flow path losses, i.e. through wall losses (absorption and adsorption effects), condensation etc. Furthermore, high purity stainless steel pressure regulators were used for calibration standard gas bottles with analyte free degassing O-rings.

The compound specific per carbon atom RFs from the NPL-2017 NMHC calibration standard are summarized in Fig. 4. The AED detector should have an equimolar response but nevertheless a spread for the carbon 193 nm emission line intensity between around 550 to 900 counts pptv⁻¹ C-atom⁻¹ was observed depending on the compound. Thus, the CryoTrap-GC-AED detected response must be considered as a whole for the entire instrument where the trapping and transmission efficiency is compound specific and directly influence the final detected signal. The same applies also to the other elements measured. The average of the compound specific median RFs is 663 ± 66 counts pptv⁻¹ C-atom⁻¹. Therefore, within the uncertainty range the



185 compound independent calibration could be applied, lowering the uncertainty is possible if similar compound group's RF is used for the target species. Nevertheless, for higher accuracy a direct calibration method is recommended. Within this study a direct calibration method was used.

3.2 Iodine, sulfur, bromine and nitrogen emission line performances

The Apel-Riemer-2015 gas phase calibration standard was used to evaluate the performance of other available and relevant atomic emission lines for atmospheric chemistry (Table 2).

190 Methyl iodide (iodine wavelength measured at 178 nm) has LOD of 0.7 pptv, and RF of 1635 ± 135 counts I-atom⁻¹ pptv⁻¹. The LOD of methyl iodide on iodine emission line was 9.6 times more sensitive compared to carbon emission line. Carbonyl sulfide (OCS) and carbon disulfide (CS₂) had LODs of 1.9 and 1.8 pptv, respectively. OCS had RF of 342 ± 34 counts S-atom⁻¹ pptv⁻¹, and CS₂ had RF of 476 ± 79 counts S-atom⁻¹ pptv⁻¹. The sulfur emission line at 181 nm was determined to be on average 3.1 times more sensitive compared to the carbon 193 nm emission line, calculated from OCS and CS₂ LOD values.

195 The average sulfur response factor determined from the two latter compounds is 409 ± 57 counts S-atom⁻¹ pptv⁻¹. Bromodichloromethane (9.9 ± 1.5 counts Br-atom⁻¹ pptv⁻¹), 1,2-dibromoethane (9.0 ± 1.8 counts Br-atom⁻¹ pptv⁻¹) and bromoform (8.3 ± 2.0 counts Br-atom⁻¹ pptv⁻¹) at bromine emission line at 163 nm had significantly higher LODs of 115.7, 61.9 and 64.2 pptv, respectively compared to iodine, sulfur and carbon emission lines. The average per bromine atom response factor was determined 9.1 ± 1.8 counts Br-atom⁻¹ pptv⁻¹. Nitrogen at 174 nm proved to be the least sensitive element for AED

200 III detector. Nitrogen containing compounds: acetonitrile, acrylonitrile, isopropyl nitrate, propyl nitrate and isobutyl nitrate varied significantly with their LODs (see Table 2), despite all containing a single nitrogen atom. The different transmission efficiency of each of the five nitrogen compounds through the system's flow paths could cause such differences. The average per nitrogen atom response factor determined from the five nitrogen compounds was 28 ± 2 counts N-atom⁻¹ pptv⁻¹. The different element count scales are non-related to each other.

205 3.3 Advantages and disadvantages of the CryoTrap-GC-AED

Considering all the species measured with the CryoTrap-GC-AED, the system proved to be very sensitive detecting iodine, sulfur and carbon elements with RFs 1635 ± 135 counts pptv⁻¹ I-atom⁻¹, 409 ± 57 counts pptv⁻¹ S-atom⁻¹ and 636 ± 69 counts C-atom⁻¹ pptv⁻¹, respectively. The different elemental emission wavelength intensity count scales are not directly comparable due to different elemental background noise levels. From these three elements the lowest background noise level was measured

210 for iodine, followed by sulfur and then by carbon. The element background noise level determines the LODs for these elements in the same increasing order, also seen in the Table 1 and 2.

The great advantage of the AED system is that compounds which are insensitive to the flame ionization detector (FID) and other detectors, e.g. carbonyl sulfide, can be measured by exploiting other emission lines such as sulfur. Generally, the CryoTrap-GC-AED system is in the same sensitivity range as an FID detector (Baker et al., 2010). The electron capture detector (ECD) remains more sensitive and sometimes no pre-concentration of the air sample is necessary (e.g. Schuck et al.,

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2009) however, an ECD is limited only to the electron capturing compounds. The GC-MS systems generally have high sensitivity and good selectivity but require separate calibration gases for all species to be quantified since response factors vary considerably. Modern online mass spectrometry (MS) detectors, especially the proton transfer reaction time-of-flight mass spectrometry (PTR-TOF-MS) provide high sensitivity (sub pptv) and highly time resolved (< 1 s) measurement techniques
220 without the need for pre-concentration, e.g. the new Tofwerk / Aerodyne Research Vocus PTR-TOF (Krechmer et al. 2018). However, without pre-separation with gas chromatography the chemical identity of signals can be ambiguous.

The specific emission wavelength range of the high-resolution AED III enables measurement capability to accurately detect at least 11 elements: antimony, arsenic, carbon, germanium, iodine, nitrogen, oxygen (requires 5% methane in 95% nitrogen reactant gas), phosphorus, selenium, silicon and sulfur, of which a maximum of 8 can be simultaneously recorded from the
225 raw data (JAS product description on their webpage, 2020). This makes the first screening of an unknown gas phase sample for compounds with various elements much easier and faster compared to more complex mass spectra.

Some disadvantages of the current AED III detector are the short lifetime of the fused silica tube where the plasma is being maintained and the early software / electronic issues inhibiting the proper acquisition of the data. The helium plasma is sustained in a small quartz tube (47 mm long, 1.0 mm inner diameter, 1.25 mm outer diameter) with a polyimide coating (27
230 mm long) in the middle part of the outer surface. The tube is positioned in a water-cooled cavity, maintained at 80°C. The lifetime of the discharge tube was around 5 to 30 days, sometimes up to 3-4 months, depending on the sample throughput and quality of the replacement of the fragile discharge tube. The discharge tube could break at any moment without a warning during a measurement sequence, leading to the loss of a sample.

Furthermore, the large consumption of liquid nitrogen (around 2-3 L sample⁻¹) for the sample cryogenic pre-concentration
235 makes it demanding on operator time, logistical field support, and somewhat expensive to operate. The use of liquid nitrogen makes it difficult to operate this instrument at remote areas. Nevertheless, the CryoTrap-GC-AED instrument was taken to a field measurement campaign at Finnish boreal forest in 2016 where among the other species it contributed to the quantification of monoterpene compounds, when accounting for the directly measured NO₃ radical reactivity in the boreal forest (Liebman et al., 2018).

240 **4 CryoTrap-GC-AED case studies**

The newly developed instrument has been deployed in near real-time field measurements and in lab based pressurized canister measurements. In the following we demonstrate two case studies, one based in the forest in Hyytiälä, Finland and the other in the home laboratory in Mainz, Germany.

4.1 Finnish boreal forest field measurements at Hyytiälä site

245 Boreal forest (taiga) makes up around 33% of Earth's forest cover, making it the largest terrestrial biome in the world. Even in the epoch of the Anthropocene about $\frac{3}{4}$ of the boreal forest remains natural (Brandt et al., 2013). The field campaign took place in a boreal forest measurement site SMEAR II (Station for Measuring Forest Ecosystem-Atmosphere Relations II) at



250 Hyytiälä, Finland in September 2016 (Hari and Kulmala, 2005). The site is situated approximately 50 km away from the first more densely populated location, thus anthropogenic influence is relatively low, particularly when the wind masses originate from the north with low human activity and pollution.

The instrumentation was installed in an air-conditioned measurement container maintained at 25°C temperature. The CryoTrap-GC-AED system measured ambient air which was pulled from the center of a shared 8.5 m tall high-flow inlet (15 cm diameter, flowrate ~ 10 m³ min⁻¹). From the 8.5 m inlet the air was drawn to the container through a 15 m long ½" (1.27 cm diameter) PFA tubing at a flowrate of ~ 20 L min⁻¹ (transmission time 3.3 s). The inlet line was insulated and heated (10°C
255 above ambient) to avoid sunlight interactions and condensation. Membrane filters (polytetrafluoroethylene with pore size 5.0 µm, 47 mm diameter by Sartorius AG, Germany) were used to avoid particles entering the tubing at the connection from the high-flow inlet and replaced every 5 days. From the 15 m long sampling line inside the container the cryogenic sample trapping phase took place with a flowrate of 200 mL min⁻¹ and a total of 1 400 mL air was trapped for each sample. After the injection of a pre-concentrated sample from the last focusing trap to the GC, the traps were baked out and the pre-concentration of a
260 next sample started in parallel with the previous GC run. This enabled mean sample throughput time intervals of 1 h 22 min. The average liquid nitrogen consumption rate was 2.5 L h⁻¹. The CryoTrap and AED parameters presented in Section 2 and GC program listed in Table 3a were used.

The diel cycles of isoprene and 5 monoterpene species (α -pinene, Δ -3-carene, β -pinene, camphene, d-limonene and isoprene) are presented in Fig. 5. All monoterpene species show daytime minima and night-time maxima. This is unexpected as the
265 emissions of monoterpenes are primarily temperature dependent and ambient temperatures were higher by day (Tarvainen et al., 2005). Here the recurring night-time MR maxima can be explained with the shallow, nocturnal boundary layer, typically accompanied by a temperature inversion in the lower troposphere, effectively trapping the ground-level emissions (Liebmann et al., 2018). The measured monoterpene species and isoprene on average accounted for about 70% of the directly measured NO₃ reactivity at night-time and about 40% during daytime, published in a separate research article (Liebmann et al., 2018).

270 The diel cycles of carbonyl sulfide and carbon disulfide are shown in Fig. 6. OCS showed daytime maxima between 9:00 and 15:00 UTC (12:00 and 18:00 local time), which is unexpected as daytime uptake by vegetation is generally regarded as the dominant sink for OCS (Sandoval-Soto et al., 2005). Again, the previously discussed shallow nocturnal boundary layer plays a role, as much less in-mixing of higher concentration OCS from the free troposphere takes place during night-time. Furthermore, it confirms the recently published finding at the same measurement location that OCS uptake is light independent
275 and controlled by stomatal opening, and therefore stomatal conductance and OCS uptake can continue during night-time under these conditions (Kooijmans et al., 2017). The night-time uptake of OCS by vegetation should be further characterized and parameterized for 3D global chemistry-transport model applications. CS₂ did not show any significant diel cycle and MRs significantly varied between 0.5 and 5 pptv.



4.2 IAGOS-CARIBIC whole-air sample measurement with CryoTrap-GC-AED

280 The IAGOS-CARIBIC (In-service Aircraft for a Global Observing System – Civil Aircraft for the Regular Investigation of the
Atmosphere Based on an Instrument Container) (Brenninkmeijer et al., 2007) is a regular observation project with scientific
instruments on board commercial aircraft (Lufthansa Airbus A340-600 in cooperation with Lufthansa AG) which carries out
atmospheric monitoring on a global scale, especially in the upper troposphere / lower stratosphere (UT/LS, ~ 10 – 12 km)
285 region. Since December 2015, a new CryoTrap–GC–AED system has been applied to measure whole-air pressurized gas
samples that were taken as part of this project. Many VOCs such as NMHCs (Baker et al., 2010), sulfur containing species,
oxygenated and halogenated trace gases can be measured in the C3 – C14 range. These species are important to tropospheric
ozone production, stratospheric ozone depletion and radiative forcing.

The whole-air samples (WAS) were pressurized at about 4 bar absolute pressure with two triggered retrospective air collectors
(TRACs) each consisting of 14 specially manufactured glass vessels (2.74 L), and a high-resolution sampler (HIRES) which
290 consists of 88 stainless steel canisters (1.0 L). Every time after the flights took place and the samplers were delivered to the
lab, the measurement procedure began with the initial pressure measurement of all the 116 individual samples.

CryoTrap-GC-AED was applied to measure the IAGOS-CARIBIC TRAC and HIRES whole-air samples. At the beginning of
each measurement sequence, a 2.0 m long sample line was connected to a TRAC or HIRES sampler port, and a leak check is
conducted by evacuating the connected sample line to ~3.0 psi (0.21 bar) absolute pressure with the CryoTrap instrument
295 pump. After the pressure stabilization in the line, a leak check was carried out for 3 min. If the pressure reading was less than
 ± 0.5 psi (0.03 bar) different, which is within the precision range of the pressure gauge, the line is considered leak tight. The
CryoTrap traps Module 1 and Module 2 (Fig. 1) are heated at 150°C and 220°C, respectively for 20 min for conditioning. The
GC oven program is summarized in the Table 3 below. Again, the three independent gas phase calibration standards (Apel-
Riemer-2015, NPL-2017, NOAA-2017) were used for calibrating the system (Table S1 and S2). A 1400 mL aliquot of an
300 IAGOS-CARIBIC air sample was pre-concentrated. Exactly the same conditions were used for the calibration standards
(except NPL standard where 50 mL were pre-concentrated) in order to have the same conditions and to thereby minimize the
instrumental uncertainty. The IAGOS-CARIBIC WAS measurement sequence with the CryoTrap-GC-AED starts with a
calibration. The measurement of zero-air (produced the same way as described in Section 2) is followed by the certified
ambient air standard (NOAA-2017) measurement. The latter is followed by diluted Apel-Riemer-2015 gas phase standard
305 measurement (as described in Section 2). After that the continuous measurement of the WAS samples started. The NOAA-
2017 certified ambient air reference calibration standard and the NPL 30 component ozone precursor NMHC reference
calibration standard were measured after every five WAS sample measurement to compensate for instrumental sensitivity drift
throughout the analysis.

An example dataset of OCS MRs measured in the UT/LS region from two flights (Munich to Los Angeles and Munich to
310 Shanghai in February 2019) are depicted in Fig. 7. The OCS data combined with the other VOC species from the same samples
was used to determine the global atmospheric and lower stratospheric lifetime, troposphere to stratosphere flux of OCS and



the stratospheric sink using the linear relationship between the long-lived species MRs (and their variability) to their known atmospheric lifetimes. This will be discussed in detail in a separate research article (article under review).

5 Conclusions

315 The newly developed CryoTrap-GC-AED analytical instrument enables the measurement of gas phase samples in minute concentrations (low pptv level) suitable for ambient air VOC species measurements in the troposphere and lower stratosphere. With this instrument it is possible to measure samples from canisters (e.g. IAGOS-CARIBIC whole-air samples) and also directly with circa 1-hour sampling frequency at a measurement station (e.g. Hyytiälä Finnish boreal forest measurement campaign in 2016) (Karu, 2019; Liebmann et al., 2018). The instrument proved to be sensitive and linear over more than 4
320 orders of magnitude ($> 10^5$), however, the application of the equimolarity feature of the detector is limited by pre-concentration and transmission losses. Nevertheless, known compounds present in a chromatogram and not present in the calibration standard can be still quantified with an accuracy of around 30-40%. For more accurate results direct calibration against certified gas phase calibration standards with exactly the same measurement conditions and volumes is recommended. The possibility to measure 11 elements and 8 of them simultaneously might allow discovery of new atmospheric species of interest, e.g.
325 containing selenium or silicon atoms, particularly in marine of volcanically influence environments. The instrument would become significantly more reliable if longer lasting helium discharge tubes were developed and the commercial software and electronics were improved. For further developments of the detector the possibility to measure a wider spectral range would allow detection of even more elements simultaneously.

330 *Competing interests.* The authors declare that they have no conflict of interest.

Author contribution: EK and JW developed the idea. EK established the new measurement technique. EK, ML and LE carried out the experiments. CAMB developed the CARIBIC air sampling systems. EK wrote the manuscript with support from ML and JW. All authors discussed the results, commented and helped to improve the manuscript.

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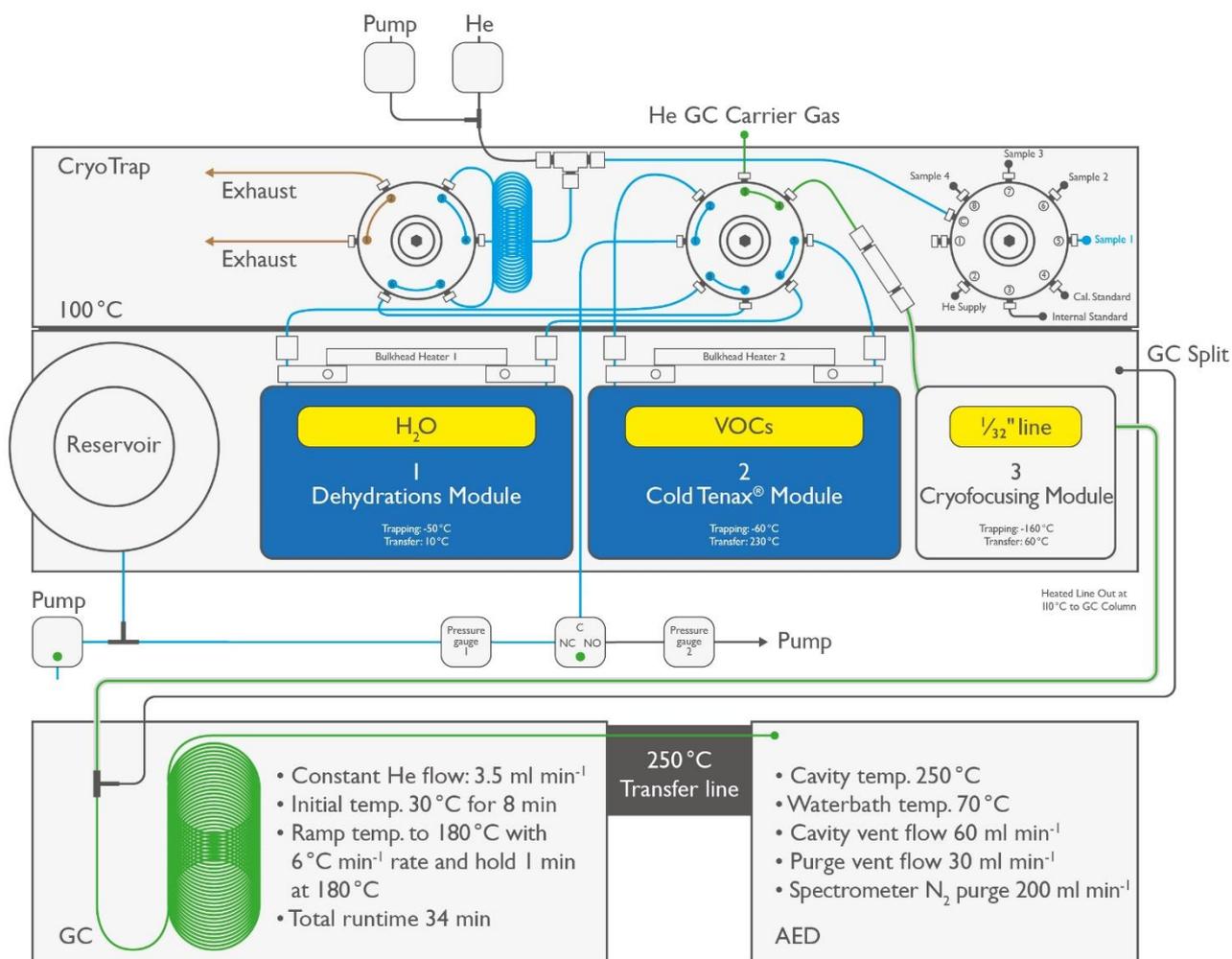
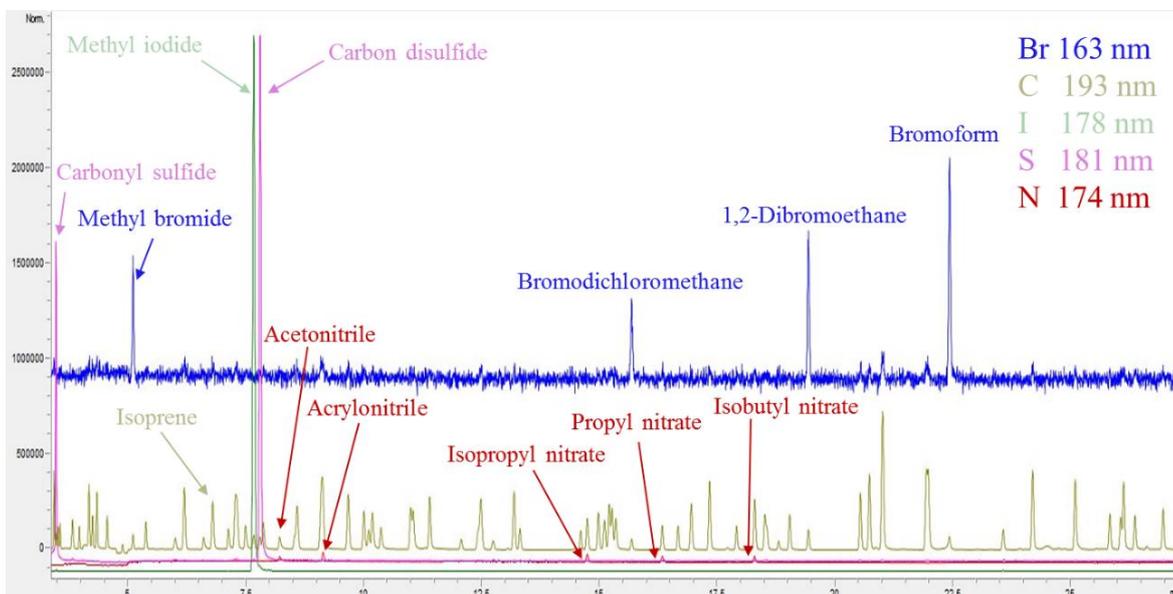
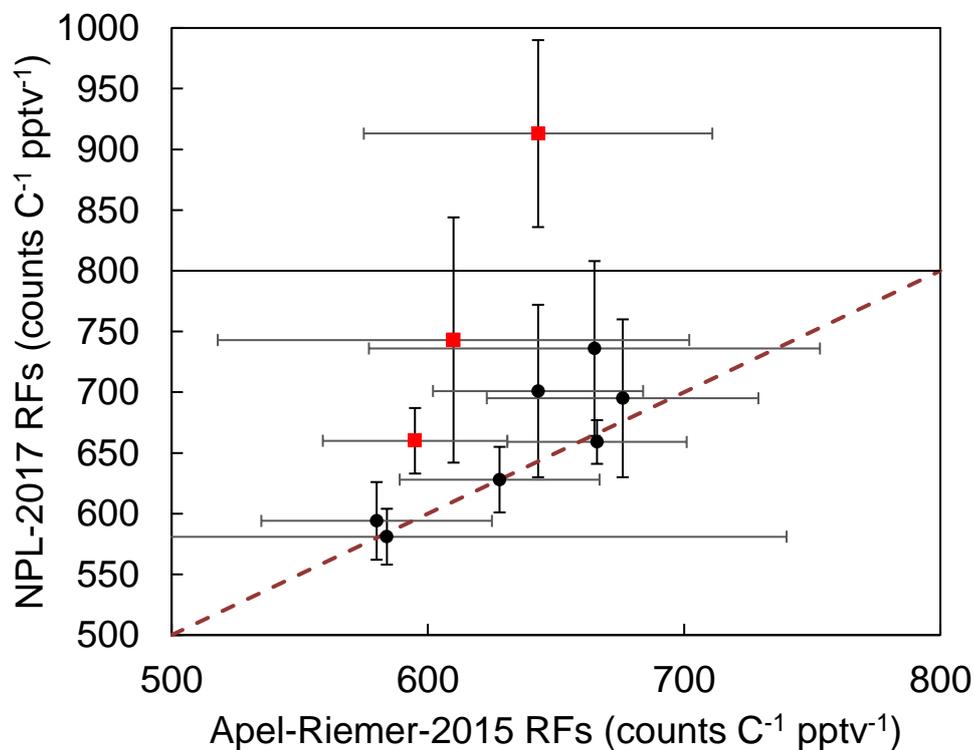


Figure 1: Schematic overview of instrumental setup, CryoTrap–GC–AED.



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Figure 2: Example CryoTrap-GC-AED normalized multi-element overlay chromatogram of ~250 pptv 84 component Apel-Riemer-2015 gas calibration standard.

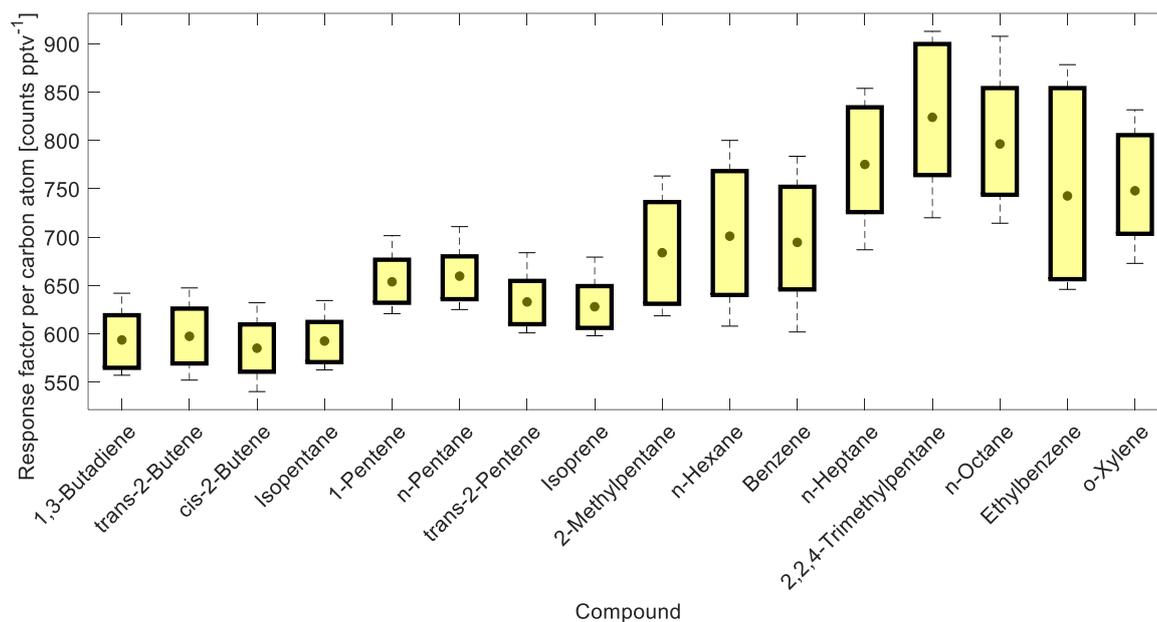


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Figure 3: Response factor comparison between the 10 common compounds measured from both Apel-Riemer-2015 and NPL-2017 gas calibration standards. The black circles show the average of all the calibration level data points for the according compounds measured in the Apel-Riemer-2015 and NPL-2017 gas calibration standards. The x and y-axis whiskers represent the standard



deviation of all the measured data points for the according compound in the Apel-Riemer-2015 and NPL-2017 standards, respectively. The brown dashed line shows the one to one RF correlation between the two measurement experiments. The red squares with black whiskers show the pentane, ethylbenzene and toluene 1σ outliers (from bottom of the graph to top, respectively).



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Figure 4: Boxplot graph from a selection of compound-specific per carbon atom RFs determined from the NPL-2017 NMHC primary calibration gas standard. The compound carbon emission line (193 nm) response factors as counts C-atom⁻¹ pptv⁻¹ are ordered by a growing number of carbon atoms on the x-axis. Central gray circles indicate the means, top and bottom edges of the box represent the 75th and 25th percentiles respectively, the whiskers show the single highest and lowest data point spread, N=176.

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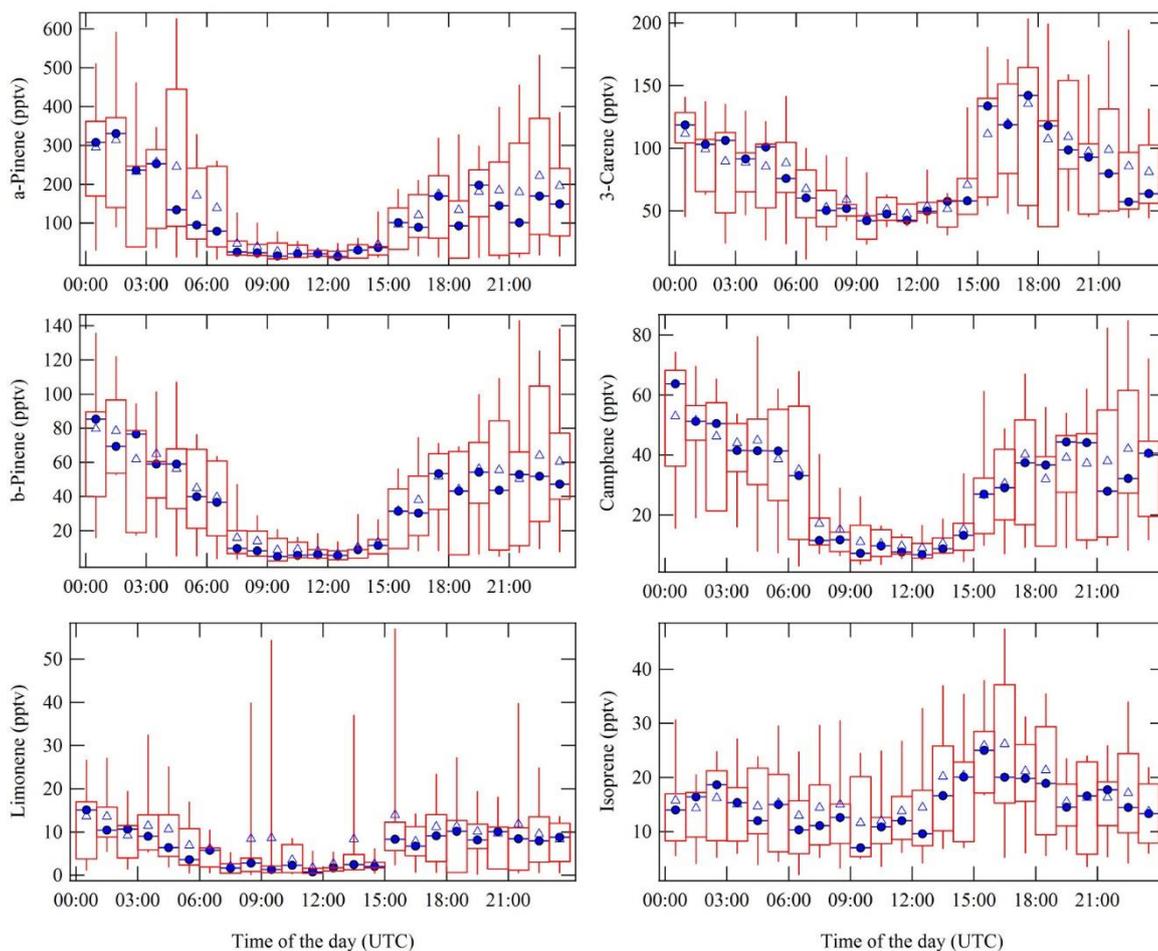
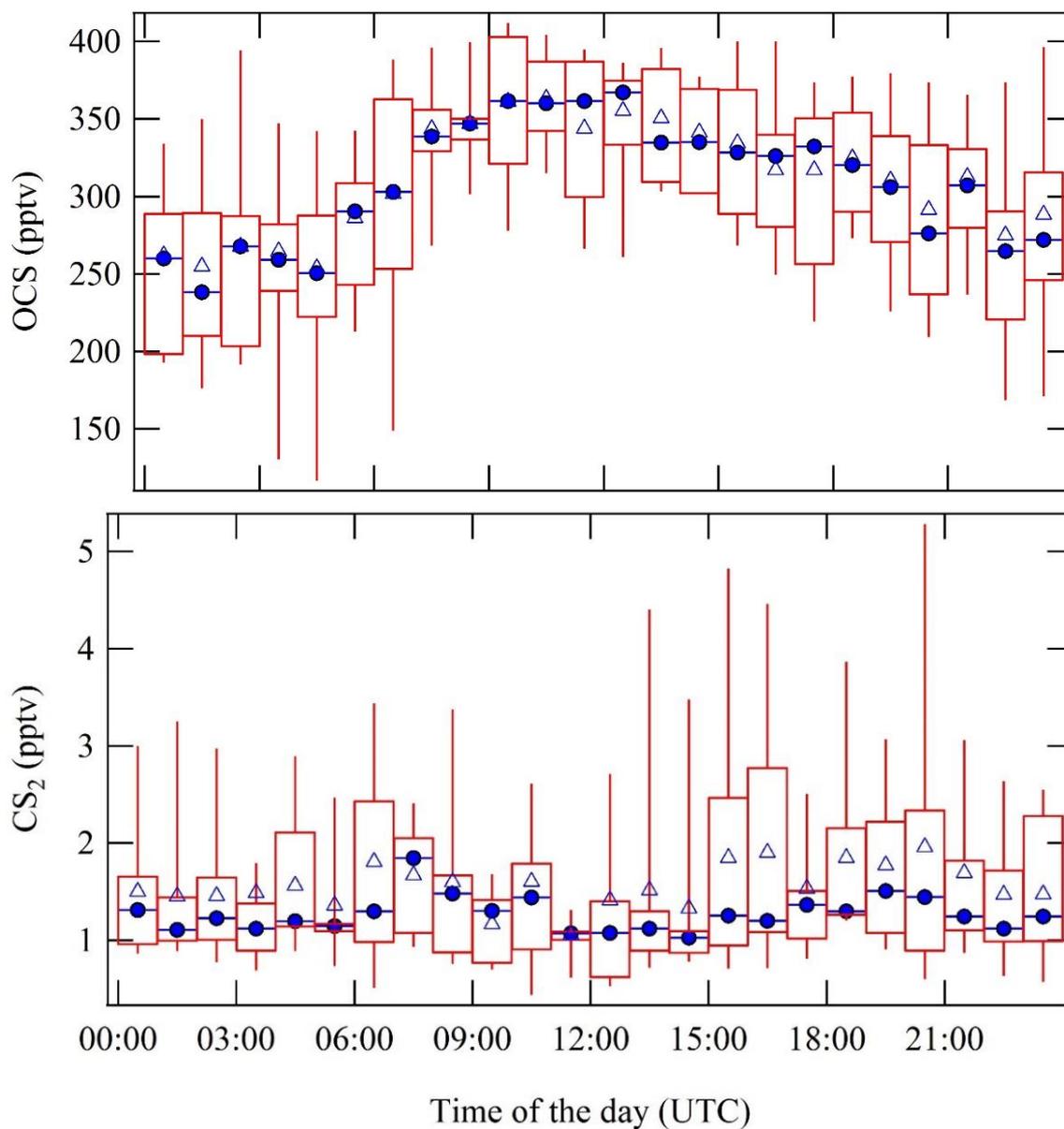
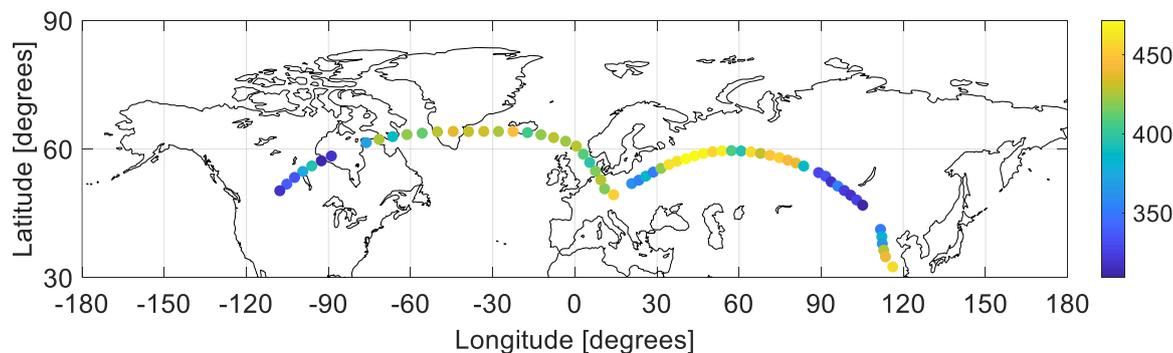


Figure 5: Complete measurement period average diel cycles of α -pinene, Δ -3-carene, β -pinene, camphene, d-limonene and isoprene. For excluding the nearby sawmill (5 km in southeast direction) monoterpene emission transport event, the data from September 9 until 11 6:30 UTC were not included in the diel cycles. Blue triangles resemble the campaign overall hourly average and blue circles represent the hourly median values. The red rectangles with the whiskers show the data spread, where 50% of the data falls into the rectangles. The upper whiskers signify the upper 75-percentile data spread and the lower whiskers indicate the lower 25-percentile of the data variation. Substantially higher MRs were observed at night-time compared to day.

365



370 **Figure 6: Diel cycles of OCS MRs (upper) and CS₂ MRs (bottom) based on all AED measured data points throughout the measurement campaign. Blue triangles show the overall hourly mean and blue circles show the hourly median values. The red rectangles with the whiskers show the data spread, where 50% of the data falls into the rectangles. The upper whiskers represent the upper 75-percentile data spread and the lower whiskers denote the lower 25-percentile data variation.**



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Figure 7: Two IAGOS-CARIBIC example flights from Munich to Los Angeles and from Munich to Shanghai (in February 2019) with OCS MRs according to the color scale [pptv].

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Table 1: CryoTrap-GC-AED system limits of detection (LODs) on 3σ level, R^2 linearity and average response factors (RFs) per carbon atom values derived from (a) Apel-Riemer-2015 and (b) NPL-2017 gas calibration standard measurements.

(a) Apel-Riemer-2015

Compound	LOD (pptv)	R^2	No. of C atoms	Average RF per C atom \pm standard deviation (counts C-atom ⁻¹ pptv ⁻¹)
Carbonyl sulfide	6.1	0.99832	1	637 \pm 37
Dichlorodifluoromethane (R-12)	2.7	0.99189	1	589 \pm 39
Chlorodifluoromethane (HCFC-22)	2.4	0.99589	1	745 \pm 32
Chloromethane	2.9	0.99581	1	661 \pm 43
Methanol	3.7	0.92247	1	529 \pm 172
Methyl bromide	5.1	0.99429	1	553 \pm 64
Trichlorofluoromethane	5.6	0.99431	1	652 \pm 34
Methyl iodide	6.7	0.99668	1	552 \pm 68
Carbon disulfide	5.4	0.99015	1	696 \pm 109
Chloroform	5.7	0.99474	1	652 \pm 55
Tetrachloromethane	7.0	0.99447	1	637 \pm 69
Bromoform	5.8	0.99384	1	737 \pm 57
Vinyl chloride	2.0	0.99492	2	549 \pm 46
Acetaldehyde	1.8	0.99402	2	636 \pm 35
Chloroethane	2.6	0.99469	2	550 \pm 46
Acetonitrile	4.8	0.96653	2	523 \pm 133
1,1-Dichloroethane	3.9	0.97800	2	430 \pm 35



1,2-Dichloroethane	3.3	0.99494	2	563 ± 46
Trichloroethylene	9.7	0.95093	2	457 ± 280
1,1,2-Trichloroethane	3.3	0.99287	2	715 ± 99
1,2-Dibromoethane	3.4	0.99489	2	638 ± 98
1,1,1,2-Tetrachloroethane	3.2	0.99468	2	654 ± 100
Propene	0.8	0.99478	3	620 ± 33
2-Propanol	1.7	0.99457	3	698 ± 37
Isopropyl nitrate	2.2	0.99886	3	742 ± 137
Propyl nitrate	9.6	0.99813	3	439 ± 232
cis-1,3-Dichloropropene	2.8	0.99452	3	512 ± 84
Isobutene	0.9	0.99532	4	666 ± 35
1,3-Butadiene	1.0	0.99492	4	580 ± 45
Methacrolein	1.5	0.99458	4	728 ± 79
Butanol	2.5	0.99537	4	384 ± 42
Isobutyl nitrate	2.0	0.99878	4	800 ± 291
Pentane	1.1	0.98933	5	595 ± 36
Isoprene	1.2	0.99494	5	628 ± 39
Hexane	1.0	0.99496	6	643 ± 41
Benzene	0.9	0.99451	6	676 ± 53
4-Methyl-2-pentanone (MIBK)	1.9	0.99513	6	566 ± 150
2-Hexanone	2.7	0.99284	6	357 ± 129
Hexanal	6.1	0.98819	6	386 ± 182
Chlorobenzene	1.4	0.99540	6	573 ± 107
1,3-Dichlorobenzene	2.8	0.98789	6	399 ± 133
Benzyl chloride	4.7	0.97921	6	343 ± 147
1,2-Dichlorobenzene	2.6	0.99017	6	412 ± 125
Toluene	0.9	0.99502	7	643 ± 68
Ethylbenzene	0.9	0.99536	8	610 ± 92
1,3,5-Trimethylbenzene	0.9	0.99470	9	665 ± 88
1,2,4-Trimethylbenzene	1.1	0.99135	9	584 ± 156

(b) NPL-2017



Compound	LOD (pptv)	R ²	No. of C atoms	Average RF per C atom ± standard deviation (counts C-atom ⁻¹ pptv ⁻¹)
Propene & Propane	0.8	0.99444	3	627 ± 18
Isobutane	0.8	0.99164	4	659 ± 18
1,3-Butadiene	0.8	0.99432	4	594 ± 32
trans-2-Butene	0.8	0.99083	4	598 ± 33
cis-2-Butene	1.0	0.99129	4	585 ± 30
Isopentane	1.1	0.99598	5	593 ± 26
1-Pentene	0.9	0.99263	5	654 ± 26
n-Pentane	0.9	0.99263	5	660 ± 27
trans-2-Pentene	0.9	0.99026	5	633 ± 27
Isoprene	1.0	0.99181	5	628 ± 27
2-Methylpentane	0.9	0.97498	6	684 ± 54
n-Hexane	0.8	0.96687	6	701 ± 71
Benzene	0.7	0.96992	6	695 ± 65
n-Heptane	0.6	0.99386	7	775 ± 62
Toluene	0.5	0.99127	7	913 ± 77
2,2,4-Trimethylpentane	0.6	0.99205	8	824 ± 76
n-Octane	0.5	0.98977	8	796 ± 68
Ethylbenzene	0.5	0.92709	8	743 ± 101
o-Xylene	0.5	0.99552	8	748 ± 60
1,3,5-Trimethylbenzene	0.5	0.96353	9	736 ± 72
1,2,4-Trimethylbenzene	0.6	0.99727	9	581 ± 23
1,2,3-Trimethylbenzene	0.6	0.99681	9	619 ± 26

385 **Table 2: CryoTrap-GC-AED system limits of detection (LODs) on 3σ level, R² linearity and response factors (RFs) per iodine, sulfur, bromine and nitrogen atom values derived from Apel-Riemer-2015 calibration standard measurements.**

Compound	LOD (pptv)	R ²	Element	Average RF per atom ± standard deviation (counts atom ⁻¹ pptv ⁻¹)
Methyl iodide (iodine)	0.7	0.99668	I	1635 ± 135
Carbonyl sulfide (sulfur)	1.9	0.99964	S	342 ± 34



Carbon disulfide (sulfur)	1.8	0.99966	S	476 ± 79
Bromodichloromethane (bromine)	115.7	0.98930	Br	9.9 ± 1.5
1,2-Dibromoethane (bromine)	61.9	0.99226	Br	9.0 ± 1.8
Bromoform (bromine)	64.2	0.99082	Br	8.3 ± 2.0
Acetonitrile (nitrogen)	265.1	0.98281	N	19.8 ± 2.7
Acrylonitrile (nitrogen)	139.4	0.99657	N	28.2 ± 0.8
Isopropyl nitrate (nitrogen)	64.6	0.98954	N	35.8 ± 2.9
Propyl nitrate (nitrogen)	82.0	0.99180	N	25.8 ± 2.2
Isobutyl nitrate (nitrogen)	57.1	0.99060	N	28.2 ± 2.1

Table 3: The GC oven programs for (a) Finish boreal forest and (b) IAGOS-CARIBIC measurements.

(a) Finnish boreal forest

Rate (°C min ⁻¹)	Temperature (°C)	Hold time (min)
	35	5
6	180	5
Total runtime: 34.0 min		
Pressure (psi)	Average velocity (cm sec ⁻¹)	Holdup time (min)
44.25 (3.05 bar)	44.02	2.27

390

(b) IAGOS-CARIBIC

Rate (°C min ⁻¹)	Temperature (°C)	Hold time (min)
	30	8
6	180	1
Total runtime: 34.2 min		
Pressure (psi)	Average velocity (cm sec ⁻¹)	Holdup time (min)
45.02 (3.10 bar)	44.23	2.26



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