- $1 \qquad \text{Estimation of pollen counts from light scattering intensity when sampling multiple pollen taxa} \qquad --$
- 2 Establishment of Automated Multi-taxa Pollen Counting Estimation System (AME System)—
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12 Abstract.

Laser optics have long been used in pollen counting systems. To clarify the limitations and potential new applications of laser optics for automatic pollen counting and discrimination, we determined the light scattering patterns of various pollen types, tracked temporal changes in these distributions, and introduced a new theory for automatic pollen discrimination. Our experimental results indicate that different pollen types often have different light scattering characteristics, as previous research has suggested. Our results also show that light scattering distributions did not undergo significant temporal changes. Further, we show that the concentration of two different types of pollen could be estimated separately from the total number of pollen grains by fitting the light scattering data to a probability density curve. These findings should help realize a fast and simple automatic pollen monitoring system.

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1 Introduction

Pollen counting is a time-consuming and labor-intensive task that requires professional skills. However, recent technological developments have made automatic pollen sampling and identification possible (Buters et al. 2018), for example, with recognition systems using microscopic images of pollen grains (Boucher et al. 2002; Ranzato et al. 2007; Oteros et al. 2015), pollen color patterns from pollen images (Landsmeer et al. 2009), fluorescence emission signals, (Swanson and Huffman 2018; Mitsumoto et al. 2009; Mitsumoto et al. 2010; Richardson et al. 2019), light scattering (Crouzy et al. 2016; Šaulienė et al. 2019, holographic images (Sauvageat et al. 2019), size and morphological characteristics (O'Connor et al. 2013), real-time PCR (Longhi et al. 2009), texture and infrared patterns of microscopic images of pollen (Marcos et al. 2015; Gottardini et al. 2007; Chen et al. 2006), or a combination of several of these. Many studies applied machine learning algorithms to the problem (Punyasena et al. 2012; Tcheng et al. 2016; Crouzy et al. 2016; Gonçalves et al. 2016; Gallardo-Caballero et al. 2019; Šaulienė et al. 2019). These automated pollen identification methods have been applied not only to aerobiological research but also to palynological studies for the identification of fossilized pollen (France et al. 2000; Kaya et al. 2014; Li et al. 2004; Zhang et al. 2004; Rodríguez-Daminán et al. 2006).

Analysis using light scattering patterns has a particular focus, with several methods being developed for establishing an automatic aerosol or bioaerosol counting system (Huffman et al. 2016). For example, polarization signals can be used to discriminate *Cryptomeria japonica* from polystyrene spherical particles (Iwai 2013). Studies applying machine learning algorithms have shown that light scattering patterns can be used for automatic classification and counting of multiple pollen taxa simultaneously (Crouzy et al., 2016; Sauliene et al., 2019). Other studies have applied statistical techniques to compare the light scattering data and number of multiple taxa pollen grains (Kawashima et al. 2007, 2017; Matsuda and Kawashima 2018). Surbek et al. (2011) also studied the discrimination method for Hazel, Birch, Willow, Ragweed, and Pine pollen showing that they have distinct characteristics in the backward and sideward light scattering patterns.

In the present study, light scattering patterns from various pollen taxa are investigated with a KH-3000 to verify whether they have different light scattering patterns. A novel method is also proposed to discriminate between two taxa with similar scattering patterns.

51 2 Materials and methods

A protection cylinder (radius = 5 cm, height = 30 cm) was attached to the sampling tube of a KH-3000-01 laser-optics-based automatic pollen counter (Yamatronics, Japan). The KH-3000-01 is a widely used automatic pollen counting system (e.g. Wang et al. 2014; Takahashi et al. 2001; Miki et al. 2017, 2019; Kawashima et al. 2007, 2017; Matsuda and Kawashima 2018). A laser irradiates particles that pass through the sampling system and the forward and side scattering signals from each particle are recorded. In this study pollen grains from known taxa were injected through an injection tube in the wall of the protection cylinder and sampled in the KH-3000-01 (Fig. 1). The side and forward scattering intensities were evaluated by converting the light intensity into a voltage. The relationship between the light intensity and the physical properties which are size and roughness of the particle surface of sampled particle (Matsuda and Kawashima 2018).

2.1 Temporal changes in light scattering patterns

Alnus pollen grains were directly sampled from catkins on a tree growing at the Swiss Federal Office of Meteorology and Climatology on a sunny morning on February 28 2019. Light scattering measurements were taken using the fresh pollen grains soon after they were collected. The remaining pollen grains were stored in tubes and scattering patterns were reevaluated after storing them for 1 h, 2 h, 6 h, and 10 days. Multiple comparisons using the Bonferroni method were performed on the side and forward scattering data to assess whether the light scattering distributions showed changes after storage. Bonferroni method is a multiple comparison method used for non-parametric data sets. In order to carry out the multiple comparison, 316 scattering data of each taxa were picked up because the Bonferroni method requires the same amount of data of each taxa and 316 scattering data was the smallest amount of data amongst each time step (10 day).

2.2 Light scattering patterns of different pollen taxa

Dried pollen grains from *Alnus*, *Ambrosia*, *Artemisia*, *Betula*, *Castanea*, *Cedrus*, *Corylus*, *Fagus*, *Fraxinus*, *Helianthus*, *Olea*, *Phleum*, *Quercus*, *Taxus*, and *Zea* were sampled in a similar way. These taxa are representative of the pollen types commonly observed in Europe. After collecting the light scattering distributions of each pollen type, multiple comparisons using the Bonferroni method were performed to evaluate whether these distributions

- differ significantly from each other. In order to carry out the multiple comparison, 210 scattering data of each taxa were picked up based on the smallest amount of data amongst the taxon (*Helianthus*).
- 78 2.3 Automatic discrimination theory
- To carry out simple and fast automatic pollen discrimination, the number of pollen grains of each type from the total number of pollen grains was calculated as follows.
- For two different types of pollen (A and B) in the side scattering intensity range a b and in the forward scattering intensity range c d, the following equation holds:

$$\begin{split} &\int_{a}^{b} P_{A_{side}(x)} \, \mathrm{d}x = p_{A_{side}} \\ &\int_{a}^{b} P_{B_{side}(x)} \, \mathrm{d}x = p_{B_{side}} \\ &\int_{c}^{d} P_{A_{front}(x)} \, \mathrm{d}x = p_{A_{front}} \\ &\int_{c}^{d} P_{B_{front}(x)} \, \mathrm{d}x = p_{B_{front}} \end{split} \tag{1}$$

- where *P* is the representative probability density function of the scattering intensity. *p* is the representative probability of the scattering intensity of each pollen grain lying in the integration intervals.
- Next, the scattering intensity distribution that gives the number of pollen grains at each scattering intensity was fitted to a distribution function. In this experiment, the normal distribution was fitted to the number of pollen grains in every 100 mV steps. The gaussian function is written as:

$$f_{(x)} = \frac{\alpha}{\sqrt{2\pi}} \exp\left\{-\frac{(x-\mu)^2}{2\sigma^2}\right\} + c$$
 (2)

- where α and c are coefficients, μ is the mean, σ is the standard deviation.
- Fitting the data to the normal distribution function enables one to calculate the probability of a pollen grain showing a certain light scattering intensity. The probability density of the normal distribution function (*P*) is written as:

$$P_{(x)} = \frac{1}{\sqrt{2\pi\sigma^2}} \exp\left\{-\frac{(x-\mu)^2}{2\sigma^2}\right\}$$
(3)

- Fitting was performed by nonlinear optimization. The normal distribution was chosen so that we can handle the light scattering plots using a known function.
- 94 Equation (1) gives

$$C_1 p_{A_{side}} N_A + C_2 p_{B_{side}} N_B = n_{side \ a-b}$$

$$C_3 p_{A_{front}} N_A + C_4 p_{B_{front}} N_B = n_{front \ c-d}$$

$$N_A + N_B = N_{total}$$
(5)

Here, *N* is the number of sampled pollen grains of each pollen type, which are the values to be calculated. *N*_{total} is the total number of sampled pollen grains and *n* is the total number of sampled pollen grains in the integration interval, which are known numbers. *C* is the correction factor defined by the following equation:

$$C = \frac{\int_{-\infty}^{+\infty} P_{(x)} dx}{\int_{0}^{4500} P_{(x)} dx}$$

$$= \frac{1}{\int_{0}^{4500} P_{(x)} dx}$$
(6)

- C is needed for renormalization of the probability distribution because the device KH-3000-01 is able to detect
- 99 the scattering intensity only in the range of 0–4500mV.
- By solving two equations in Eq. (5), N_A and N_B will be theoretically estimated.
- In this paper, *Alnus* and *Artemisia* were chosen as examples to evaluate the usability of the theory above. Because
- fitting worked well in the range of 600–800mV for the side scattering and 300–500mV for the forward scattering,
- 103 a = 600, b = 800, c = 300 and d = 500 were substituted in Eq. (5). The evaluation tests were carried out five
- times using the light scattering data for both *Alnus* and *Artemisia* (Fig. 2).
- The magnitude of the estimation error is calculated as follows.

$$error (\%) = \frac{|actual - estimation|}{actual} \times 100$$
 (7)

108 3 Results

109 3.1 Temporal changes in light scattering pattern

- 110 The scattering distribution of Alnus pollen (Fig. 3) showed no significant temporal changes in scattering
- distributions in 10 day (Table 1).

112 3.2 Light scattering distributions of different pollen taxa

- Pollen grains with smaller sizes tend to show smaller voltage values (Fig. 4). The results of the multiple
- comparisons (Table 2) indicated that there is always a significant different between side and forward scattering
- between two different pollen types except between:
- 116 Side scattering: Alnus-Ambrosia, Alnus-Corylus, Alnus-Olea, Ambrosia-Fraxinus, Betula-Phleum, Betula-
- 117 Quercus, Corylus-Olea, Fagus-Zea, Artemisia-Fraxinus, Helianthus-Zea, Phleum-Quercus
- 118 Forward scattering: Alnus-Corylus, Alnus-Quercus, Ambrosia-Artemisia, Ambrosia-Fraxinus, Artemisia-
- 119 Fraxinus, Betula-Phleum, Betula-Quercus, Castanea-Olea, Cedrus-Helianthus, Corylus-Quercus, Fagus-
- 120 Helianthus, Fagus-Zea, Phleum-Quercus

121 3.3 Automatic counting

- Counting the number of pollen grains of each type can be carried out by solving the two equations from Eq. (5),
- side $(n_{side\ a-b})$ and forward $(n_{front\ c-d})$, side $(n_{side\ a-b})$ and total (N_{total}) , forward $(n_{front\ c-d})$ and total
- (N_{total}) . The parameters of the probability density curve of the side and the forward (Fig. 5) light scattering
- distributions of *Alnus* and *Artemisia* were estimated as follows:

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$$P_{Alnus_{side}}$$
: $(\alpha, \mu, \sigma, c) = (433.58, 555.13, 223.85, 14.74)$

127
$$P_{Artemisia_{side}}: (\alpha, \mu, \sigma, c) = (588.98, 419.45, 192.67, 10.31)$$

128
$$P_{Alnus_{front}}$$
: $(\alpha, \mu, \sigma, c) = (600.25, 348.67, 159.96, 16.25)$

129
$$P_{Artemisia_{front}}: (\alpha, \mu, \sigma, c) = (1028.57, 202.64, 107.32, 13.00)$$

- The results (Fig. 6) show that the estimated number of pollen grains had average errors of 46.80%, 33.9%, 39,12%
- 131 for *Alnus* and 30.81%, 18.77%, 20.57% for *Artemisia* (Table 3).

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4 Discussion

- Temporal changes in the shapes of pollen grains are expected to affect the changes in light scattering patterns.
- However, our experimental data indicate that light scattering patterns show little to no changes over time (up to
- at least 10 days). Thus, there should be no problem using pollen grains that are either fresh or have been stored
- for several days for studies with the KH-3000. Further investigation is required to understand whether this is true

- 138 for species other than Alnus and for longer periods of time. Understanding the morphological stability of each 139 pollen type would be helpful to understand the temporal stability of light scattering patterns.
- 140 Light scattering data from various pollen taxa indicate that it is not possible to discriminate between the side
- 141 scattering patterns of Alnus vs Ambrosia, Alnus vs Corylus, Alnus vs Olea, Ambrosia vs Fraxinus, Betula vs
- 142 Phleum, Betula vs Quercus, Corylus vs Olea, Fagus vs Zea, Artemisia vs Fraxinus, Helianthus vs Zea, Phleum
- 143 vs Quecus and the forward scattering patterns between Alnus vs Corylus, Alnus vs Quercus, Ambrosia vs
- 144 Artemisia, Ambrosia vs Fraxinus, Artemisia vs Fraxinus, Betula vs Phleum, Betula vs Quercus, Castanea vs Olea,
- 145 Cedrus vs Helianthus, Corylus vs Quercus, Fagus vs Helianthus, Fagus vs Zea, , and Phleum vs Quercus, all of
- 146 which show similar scattering intensities. Although it is not clear if the classification theory introduced above is
- 147 applicable to these groups, the theory should be applicable to other pairs as long as they have different scattering
- 148 intensity distributions.
- 149 The estimation of the pollen counts of *Alnus* and *Artemisia* had average errors of approximately 40% and 23%,
- 150 respectively. Test 4 had the largest error, with approximately 134% for Alnus and approximately 44% for
- Artemisia, which increased the average error. It is difficult to identify an obvious reason for these large values, 151
- 152 but it is possible that the pollen samples were contaminated by dusts or pollen grains picked up for this experiment
- 153 were biased in size or shape. Additionally, other estimations derived from the fitted curve of the forward and the
- 154 side scattering distributions showed that even when the pollen counts are estimated only from scattering intensity
- 155 data without using total number of pollen grains, which is a known number, the pollen counts are able to be
- 156 calculated accurately. The KH-3000-01 has been widely used to estimate airborne concentrations of Cryptomeria
- 157 japonica. In this study, we found average errors of 20-40% for Alnus and Artemisia, values which are also likely
- 158 applicable to other taxa such as Cryptomeria japonica. Other taxa should, however, be investigated in future.
- 159 Pollen counts can be estimated by solving Eq. (5), which contains three equations, meaning that it is possible to
- 160 make estimates for three different pollen taxa simultaneously. If more integration intervals were picked up from
- 161 the probability density curve of the scattering intensity and added to the equation, in theory it would be possible 162 to count more pollen taxa. It is possible, however, that the accuracy of the estimated values might decline due to
- 163 the accuracy of the fitted curve. Therefore, narrowing down a target to two or three pollen types considering the
- 164 season should be helpful to make accurate automatic counts of several pollen taxa simultaneously.
- 165 In this study, the normal distribution function was chosen for fitting because of its universal property. However,
- 166 further consideration is required to determine the best function for fitting actual light scattering characteristics.

168 **5 Conclusion**

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- 169 By applying the statistical analysis method, the Bonferroni method to the scattering patterns of Alnus at each time
- 170 step, our experiment showed that there seems to be no significant temporal changes in the light scattering patterns.
- 171 We also confirmed that different pollen types do not always have different light scattering patterns. However,
- 172 when two different pollen types have different light scattering patterns, it was possible to calculate the number of
- 173 pollen grains of each taxa using these light scattering patterns by solving the probability density function of the
- 174 pattern.
- 176
 - Code/Data availability: The authors confirm that the data supporting the findings of this study are available
 - 177 within the article.
 - 179 Author contributions: Kenji Miki established the system, performed the data analysis, and wrote the manuscript.
 - 180 Shigeto Kawashima arranged the experimental setup and proofread the manuscript.
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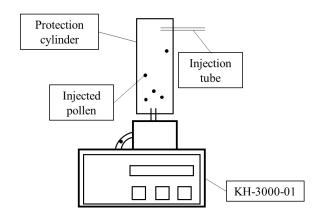
190 References

- Boucher, A., Hidalgo, P.J., Thonnat, M., Belmonte, J., Galan, C., Bonton, P., and Tomczak, R.: Development of a semi-automatic system for pollen recognition. Aerobiologia, 18, 195–201, 2002.
- Buters, J.T.M., Antunes, C., Galveias, A., Bergmann, K.C., Thibaudon, M., Galán, C., Schmidt-Weber, C., and Oteros, J.: Pollen and spore monitoring in the world. Clin. Transl. Allergy, 8, doi.org/10.1186/s13601-018-0197-8, 2018.
- 196 Chen, C., Hendrinks, E.A., Duin, R.P.W., Reiber, J.H.C., Hiemstra, P.S., de Weger, L.A., and Stoel, B.C.: 197 Feasibility study on automated recognition of allergenic pollen: grass, birch and mugwort. Aerobiologia, 198 22, 275–284. doi:10.1007/s10453-006-9040-0, 2006.
- Crouzy, B., Stella, M., Konzelmann, T., Calpini, B., and Clot, B.: All-optical automatic pollen identification: Towards an operational system. Atmos. Environ., 140, 202–212, 2016.
- France, I. Duller, A.W.G., Duller, G.A.T., and Lamb, H.F.: A new approach to automated pollen analysis. Quat. Sci. Rev., 19, 537–546, 2000.
- Gallardo-Caballero, R., García-Orellana, C. J., García-Manso, A., González-Velasco, H., Tormo-Molina, R., and Macías-Macías, M.: Precise pollen grain detection in bright field microscopy using deep learning technique. Sensors, 19, 3583. doi:10.3390/s19163583, 2019.
- Gonçalves, A.B., Souza, J.S., de Silva, G.G., Cereda, M.P., Pott, A., Naka, M.H., and Pistori, H.: Feature extraction and machine learning for the classification of Brazilian savannah pollen grains. PLoS ONE, 11, e0157044. doi:10.1371/journal.polne.0157044, 2016.
- Gottardini, E., Rossi, S., Cristofolini, F., and Benedetti, L.: Use of Fourier transform infrared (FT-IR) spectroscopy as a tool for pollen identification. Aerobiologia, 23, 211–219, 2007.
- Huffman, D. R., Swanson, B. E., and Huffman, J. A.: A wavelength-dispersive instrument for characterizing fluorescence and scattering spectra of individual aerosol particle on a substrate. Atmos. Meas. Tech., 9, 3987–3998, 2016.
- Iwai, T.: Polarization analysis of light scattered by pollen grains of Cryptomeria japonica. Jpn. J. Appl. Phys., 52, 062404. doi.org/10.7567/JJAP.52.062404, 2013.
- Kaya, Y., Mesut Pinar, S., Emre Erez, M., Fidan, M., and Riding, J.B.: Identification of *Onopordum* pollen using the extreme learning machine, a type of artificaial neural network. Palynology, 38, 129–137. doi.org/10.1080/09500340.2013.868173, 2014.
- Kawashima, S., Clot, B., Fujita, T., Takahashi, Y., and Nakamura, K.: An algorithm and a device for counting
 airborne pollen automatically using laser optics. Atmos. Environ., 41, 7987–7993, 2007.
- Kawashima, S., Thibaudon, M., Matsuda, S., Fujita, T., Lemonis, N., Clot, B., and Oliver, G.: Automated pollen monitoring system using laser optics for observing seasonal changes in the concentration of total airborne pollen. Aerobiologia, 33, 351–362, 2017.
- Landsmeer, S.H., Hendrinks, E.A., De Weger L.A., Reiber, J.H.C., and Stoel, B.C.: Detection of pollen grains in multifocal optical microscopy images of air samples. Microsc. Res. Tech., 72, 424–430, 2009.
- Li, P., Treloar, W.J., Flenley, J.R., and Empson, L.: Towards automation of palynology 2: the use of texture measures and neural network analysis for automated identification of optical images of pollen grains. J. Quat. Sci., 19, 755–762. doi: 10.1002/jqs.874, 2004.
- Longhi, S., Cristofori, A., Gatto, P., Cristofolini, F., and Grando, M.S., Gottardini, E.: Biomolecular identification of allergenic pollen: a new perspective for aerobiological monitoring? Ann. Allergy Asthma Immunol., 103, 508–514, 2009.

- Marcos, J.V., Nava, R., Cristóbal, G., Redondo, R., Escalante-Ramírez, B., Bueno, G., Déniz, Ó., González-Porto,
 A., Pardo, C., Chung, F., and Rodríguez, T.: Automated pollen identification using microscopic imaging
 and texture analysis. Micron, 68, 36–46. doi.org/10.1016/j.micron.2014.09.002, 2015.
- Matsuda, S., and Kawashima, S.: Relationship between laser light scattering and physical properties of airborne pollen. J. Aerosol Sci., 124, 122–132, 2018.
- Miki, K., Kawashima, S., Fujita, T., Nakamura, K., and Clot, B.: Effect of micro-scale wind on the measurement of airborne pollen concentrations using volumetric methods on a building rooftop. Atmos. Environ., 158, 1–10. doi.org/10.1016/j.atmosenv.2017.03.015, 2017.
- Miki, K, Kawashima, S., Clot, B., and Nakamura, K.: Comparative efficiency of airborne pollen concentration evaluation in two pollen sampler designs related to impaction and changes in internal wind speed. Atmos. Environ., 203, 18–27, doi.org/10.1016/j.atmosenv.2019.01.039, 2019.
- Mitsumoto, K., Yabusaki, K., and Aoyagi, H.: Classification of pollen species using autofluorescence image analysis. J. Biosci. Bioeng., 107, 90–94, 2009.
- Mitsumoto, K., Yabusaki, K., Kobayashi, K., Aoyagi, H.: Development of a novel real-time pollen-sorting counter
 using species-specific pollen autofluorescence. Aerobiologia, 26, 99–111, doi:10.1007/s10453-009-9147-1, 2010.
- O'Connor, D.J., Healy, D.A., and Sodeau, J.R.: The on-line detection of biological particle emissions from selected agricultural materials using the WIBS-4 (Waveband Integrated Bioaerosol Sensor) technique. Atmos. Environ., 80, 415–425, doi.org/10.1016/j.atmosenv.2013.07.051, 2013.
- Oteros, J., Pusch, G., Weichenmeier, I., Heimann, U., Möller, R., Röseler, S., Traidl-Hoffmann, C., Schmidt-Weber, C., and Buters, J.T.M.: Automatic and online pollen monitoring. Int. Arch. Allergy Immunol., 167, 158–166, doi: 10.1159/000436968, 2015.
- Punyasena, S.W., Tcheng, D.K., Wesseln, C., and Mueller, P.G.: Classifying black and white spruce pollen using layered machine learning. New Phytol., 196, 937–944, doi: 10.1111/j.1469-8137.2012.04291.x, 2012.
- Ranzato, M., Taylor, P.E., House, J.M., Flagan, R.C., LeCun, Y., and Perona, P.: Automatic recognition of biological particles in microscopic images. Pattern Recognit. Lett., 28, 31–39, doi:10.1016/j.patrec.2006.06.010, 2007.
- Richardson, S.C., Mytilinaios, M., Foskinis, R., Kyrou, C., Papayannis, A., Pyrri, I., Giannoutsou, E., and Adamakis, I.D.S.: Bioaerosol detection over Athens, Greece using the laser induced fluorescence technique. Sci. Total Environ., 696, 133906, doi.org/10.1016/j.scitotenv.2019.133906, 2019.
- Rogríguez-Damián, M., Cernadas, E., Formella, A., Fernández-Delgado, M., and De Sá-Otero, P.: Automatic
 detection and classification of grains of pollen based on shape and texture. IEEE Trans. Syst. Man Cybern.
 Syst., 36, 531–542, doi:10.1109/TSMCC.2005.855426, 2006.
- Šaulienė, I., Šukienė, L., Daunys, G., Valiulis, G., Vaitkevičius, L. Matavulj, P., Brdar, S., Panic, M., Sikoparija,
 B., Clot, B., Crouzy, B., and Sofiev, M.: Automatic pollen recognition with the Rapid-E particle counter:
 the first-level procedure, experience and next steps. Atmos. Meas. Tech., 12, 3435–3452, 2019.
- Surbek, M., Esen, C., Schweiger, G., and Ostendorf, A.: Pollen characterization and identification by elastically scattered light. J. Biophotonics, 4, 49–56, doi:10.1002/jbio.200900088, 2011.
- Swanson, B.E., and Huffman, J.A.: Development and characterization of an inexpensive single-particle fluorescence spectrometer for bioaerosol monitoring. Opt. Express, 26, 3646–3660, 2018.
- Takahashi, Y., Kawashima, S., Fujita, T., Ito, C., Togashi, R., and Takeda, H.: Comparison between real-time pollen monitor KH-3000 and Burkard sampler. Arerugi, 50, 1136–1142, 2011.

274 Tcheng, D.K., Nayak, A.K., Fowlkes, C.C., and Punyasena, S.W.: Visual recognition software for binary 275 classification and its application to spruce pollen identification. PLoS ONE, 11, e0148879. 276 doi:10.1371/journal.pone.0148879, 2016. Wang, Q., Nakamura, S., Gong, S., Suzuki, M., Nakajima, D., Takai, Y., Lu, S., Sekiguchi, K., and Miwa, M.: 277 Release behaviour of Cryptomeria japonica pollen allergenic cry j1 and cry j2 in rainwater containing 278 279 air pollutants. Int. J. Sustain. Dev. Plann., 9, 42–53, doi:10.2495/SDP-V9-N1-42-53, 2014. 280 Zhang, Y., Fountain, D.W., Hodgson, R.M., Flenley, J.R., and Gunetileke, S.: Towards automation of palynology 281 3: pollen pattern recognition using Gabor transforms and digital moments. J. Quat. Sci., 19, 763-768, 282 doi: 10.1002/jqs.875, 2004. 283

284 Figure 1 Schematic drawing of device setup. Laser irradiates pollen particle inside the KH-3000-01. 285 Figure 2 Light scattering distribution data from Alnus and Artemisia used for estimation test. 286 Figure 3 Light scattering plots for Alnus pollen – fresh and after 1h, 2h, 6h, and 10 days storage. 287 Figure 4 Light scattering distribution of various pollen taxa. 288 Figure 5 Fitted curve for side scattering (top row) and probability density curve (second row) for Alnus (left) 289 and Artemisia (right) and fitted curve for forward scattering (third row) and probability density curve (bottom 290 row) for Alnus (left) and Artemisia (right). 291 Figure 6 Results of automatic counting of Alnus and Artemisia. Red and black dots represent actual and 292 estimated numbers of pollen grains, respectively. The pair of red and black dots with the same shape are in the 293 same test set. 294 295 Table 1 Multiple comparison between *Alnus* data stored for various periods. 296 Table 2 Multiple comparison between each pollen taxon 297 Table 3 Results of estimation of number of pollen grains of Alnus and Artemisia and errors of each estimation.



302 Fig. 1 Miki and Kawashima

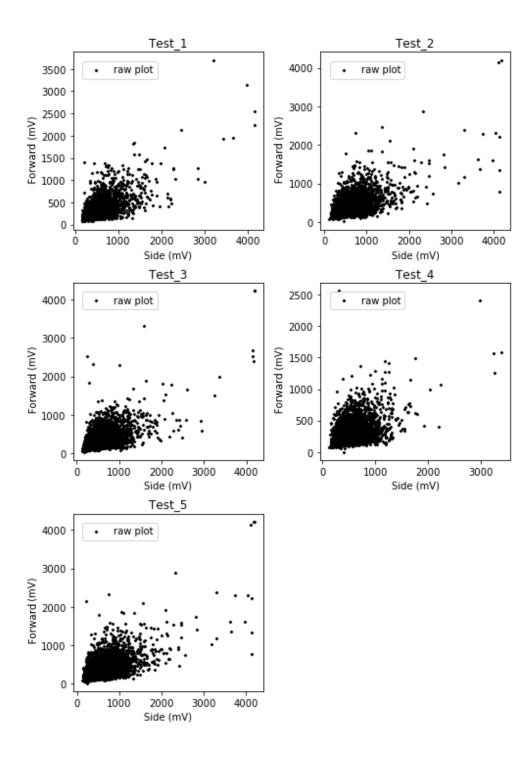


Fig.2 Miki and Kawashima

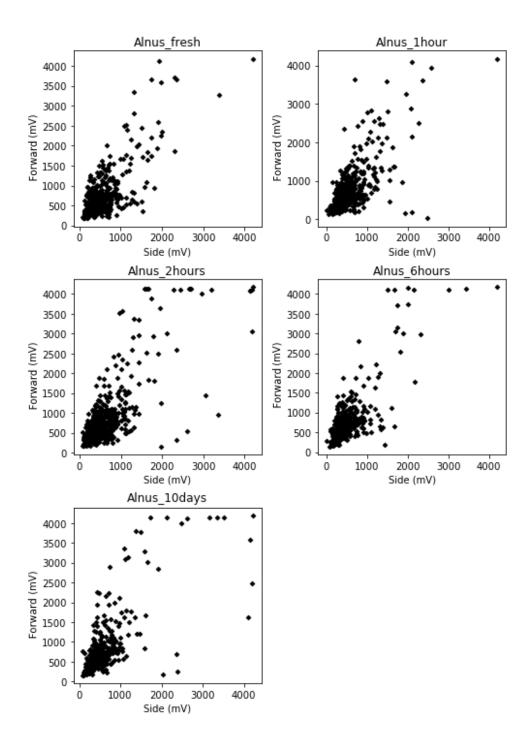


Fig.3 Miki and Kawashima

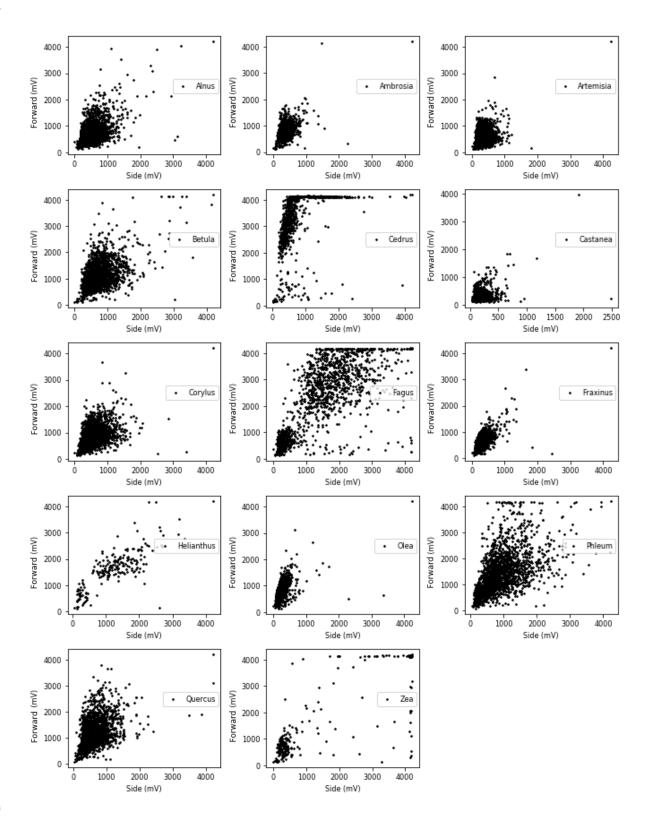


Fig.4 Miki and Kawashima

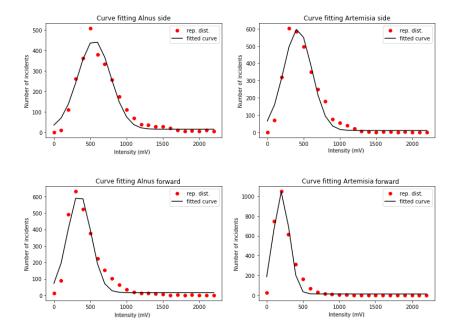
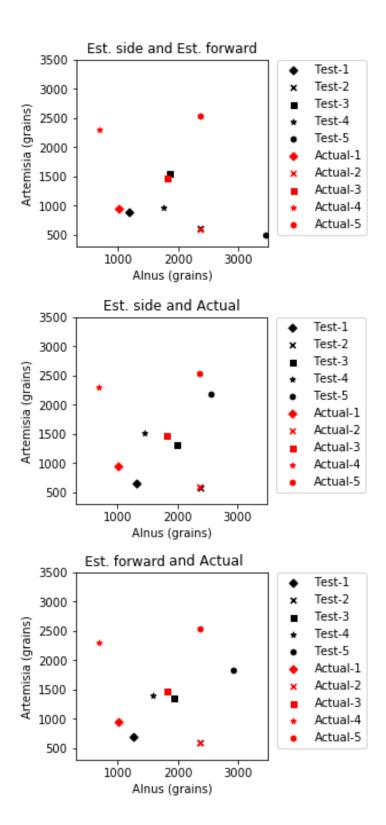


Fig.5 Miki and Kawashima



333 Fig.6 Miki and Kawashima

Table 1 Multiple comparisons between each time step (Alnus)

	Side				
		1hour	2hour	6hour	10day
_	fresh	1.00	0.38	1.00	1.00
	1hour	_	1.00	1.00	1.00
	2hour	_	_	0.71	1.00
	6hour				1.00

Forward

	1hour	2hour	6hour	10day
fresh	1.00	1.00	1.00	1.00
1hour	_	1.00	0.84	1.00
2hour	_	_	1.00	1.00
6hour	_	_	_	0.31

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Table 2 Multiple comparisons between each pollen taxon

Side													
	Ambrosia	Artemisia	Betula	Castanea	Cedrus	Corylus	Fagus	Fraxinus	Helianthus	Olea	Phleum	Quercus	Zea
Alnus	0.34	*	*	*	*	1.00	*	*	*	1.00	*	*	*
Ambrosia	_	*	*	*	*	*	*	0.08	*	*	*	*	*
Artemisia	_	_	*	*	*	*	*	0.06	*	*	*	*	*
Betula	_	_	_	*	*	*	*	*	*	*	0.06	1.00	*
Castanea	_	_	_	_	*	*	*	*	*	*	*	*	*
Cedrus	_	_	_	_	_	*	*	*	*	*	*	*	*
Corylus	_	_	_	_	_	_	*	*	*	0.49	*	*	*
Fagus	_	_	_	_	_	_	_	*	*	*	*	*	0.59
Fraxinus	_	_	_	_	_	_	_	_	*	*	*	*	*
Helianthus	_	_	_	_	_	_	_	_	_	*	*	*	1.00
Olea	_	_	_	_	_	_	_	_	_	_	*	*	*
Phleum	_	_	_	_	_	_	_	_	_	_	_	1.00	*
Quercus			_		_		_			_			*

^{*}*p* < 0.05

Forward													
	Ambrosia	Artemisia	Betula	Castanea	Cedrus	Corylus	Fagus	Fraxinus	Helianthus	Olea	Phleum	Quercus	Zea
Alnus	*	*	*	*	*	1.00	*	*	*	*	*	1.00	*
Ambrosia	_	0.95	*	*	*	*	*	1.00	*	*	*	*	*
Artemisia	_	_	*	*	*	*	*	1.00	*	*	*	*	*
Betula	_	_	_	*	*	*	*	*	*	*	1.00	1.00	*
Castanea	_	_	_	_	*	*	*	*	*	1.00	*	*	*
Cedrus	_	_	_	_		*	*	*	1.00	*	*	*	*
Corylus	_	_	_	_	_	_	*	*	*	*	*	1.00	*
Fagus	_	_	_	_		_	_	*	0.14	*	*	*	1.00
Fraxinus	_	_	_	_		_	_	_	*	*	*	*	*
Helianthus	_	_	_	_	_	_	_	_	_	*	*	*	*
Olea	_	_	_	_		_	_	_	_	_	*	*	*
Phleum	_	_	_	_	_	_	_	_	_	_	_	0.10	*
Quercus	_	_	_	_	_	_	_	_	_	_		_	*

^{*} p < 0.05

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		Test 1		7	Test 2	Test 3		
		Alnus (error)	Artemisia (error)	Alnus (error)	Artemisia (error)	Alnus (error)	Artemisia (error)	
	Side and Forward	1183 (17.36%)	881 (6.77%)	2367 (0.17%)	612 (3.20%)	1855 (1.76%)	1552 (5.43%)	
Estimation	Total and Side	1310 (29.96%)	642 (32.06%)	2386 (0.63%)	577 (2.70%)	1984 (8.83%)	1310 (11.01%)	
	Total and Forward	1259 (24.90%)	694 (26.56%)	2378 (0.30%)	585 (1.35%)	1932 (5.98%)	1362 (7.47%)	
	Actual	1008	945	2371	593	1823	1472	

		Te	st 4	Test 5		
		Alnus (error)	Artemisia (error)	Alnus (error)	Artemisia (error)	
	Side and Forward	1753 (157.42%)	968 (57.86%)	3469 (57.32%)	489 (80.76%)	
Estimation	Total and Side	1458 (114.10%)	1520 (33.83%)	2567 (16.42%)	2179 (14.28%)	
	Total and Forward	1577 (131.57%)	1402 (38.96%)	2929 (32.83%)	1817 (28.52%)	
	Actual	681	2297	2205	2542	

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