

1 **Estimation of pollen counts from light scattering intensity when sampling multiple pollen taxa —**
2 **Establishment of Automated Multi-taxa Pollen Counting Estimation System (AME System)—**

3 Kenji Miki^{1,2*}, Shigeto Kawashima¹

4

5 ¹ Graduate School of Agriculture, Kyoto University, Oiwake-cho, Kitashirakawa, Sakyo-ku, Kyoto 606-8502,
6 Japan

7 ² Tokyo Institute of Technology Earth-Life Science Institute, 2-12-1-IE-1, Ookayama, Meguro-ku, Tokyo, 152-
8 8550, Japan

9

10 **Correspondence to:* Kenji Miki (kmiki@elsi.jp)

11

12 **Abstract.**

13 Laser optics have long been used in pollen counting systems. To clarify the limitations and potential new
14 applications of laser optics for automatic pollen counting and discrimination, we determined the light scattering
15 patterns of various pollen types, tracked temporal changes in these distributions, and introduced a new theory for
16 automatic pollen discrimination. Our experimental results indicate that different pollen types often have different
17 light scattering characteristics, as previous research has suggested. Our results also show that light scattering
18 distributions did not undergo significant temporal changes. Further, we show that the concentration of two
19 different types of pollen could be estimated separately from the total number of pollen grains by fitting the light
20 scattering data to a probability density curve. These findings should help realize a fast and simple automatic pollen
21 monitoring system.

22

23

24 **1 Introduction**

25 Pollen counting is a time-consuming and labor-intensive task that requires professional skills. However, recent
26 technological developments have made automatic pollen sampling and identification possible (Buters et al. 2018),
27 for example, with recognition systems using microscopic images of pollen grains (Boucher et al. 2002; Ranzato
28 et al. 2007; Oteros et al. 2015), pollen color patterns from pollen images (Landsmeer et al. 2009), fluorescence
29 emission signals, (Swanson and Huffman 2018; Mitsumoto et al. 2009; Mitsumoto et al. 2010; Richardson et al.
30 2019), light scattering (Crouzy et al. 2016; Šaulienė et al. 2019), holographic images (Sauvageat et al. 2019), size
31 and morphological characteristics (O'Connor et al. 2013), real-time PCR (Longhi et al. 2009), texture and infrared
32 patterns of microscopic images of pollen (Marcos et al. 2015; Gottardini et al. 2007; Chen et al. 2006), or a
33 combination of several of these. Many studies applied machine learning algorithms to the problem (Punyasena et
34 al. 2012; Tcheng et al. 2016; Crouzy et al. 2016; Gonçalves et al. 2016; Gallardo-Caballero et al. 2019; Šaulienė
35 et al. 2019). These automated pollen identification methods have been applied not only to aerobiological research
36 but also to palynological studies for the identification of fossilized pollen (France et al. 2000; Kaya et al. 2014; Li
37 et al. 2004; Zhang et al. 2004; Rodríguez-Daminán et al. 2006).

38 Analysis using light scattering patterns has a particular focus, with several methods being developed for
39 establishing an automatic aerosol or bioaerosol counting system (Huffman et al. 2016). For example, polarization
40 signals can be used to discriminate *Cryptomeria japonica* from polystyrene spherical particles (Iwai 2013).
41 Studies applying machine learning algorithms have shown that light scattering patterns can be used for automatic
42 classification and counting of multiple pollen taxa simultaneously (Crouzy et al., 2016; Sauliene et al., 2019).
43 Other studies have applied statistical techniques to compare the light scattering data and number of multiple taxa
44 pollen grains (Kawashima et al. 2007, 2017; Matsuda and Kawashima 2018). Surbek et al. (2011) also studied the
45 discrimination method for Hazel, Birch, Willow, Ragweed, and Pine pollen showing that they have distinct
46 characteristics in the backward and sideward light scattering patterns.

47 In the present study, light scattering patterns from various pollen taxa are investigated with a KH-3000 to verify
48 whether they have different light scattering patterns. A novel method is also proposed to discriminate between
49 two taxa with similar scattering patterns.

50

51 **2 Materials and methods**

52 A protection cylinder (radius = 5 cm, height = 30 cm) was attached to the sampling tube of a KH-3000-01 laser-
53 optics-based automatic pollen counter (Yamatronics, Japan). The KH-3000-01 is a widely used automatic pollen
54 counting system (e.g. Wang et al. 2014; Takahashi et al. 2001; Miki et al. 2017, 2019; Kawashima et al. 2007,
55 2017; Matsuda and Kawashima 2018). A laser irradiates particles that pass through the sampling system and the
56 forward and side scattering signals from each particle are recorded. In this study pollen grains from known taxa
57 were injected through an injection tube in the wall of the protection cylinder and sampled in the KH-3000-01 (Fig.
58 1). The side and forward scattering intensities were evaluated by converting the light intensity into a voltage. The
59 relationship between the light intensity and the physical properties which are size and roughness of the particle
60 surface of sampled particle (Matsuda and Kawashima 2018).

61 **2.1 Temporal changes in light scattering patterns**

62 *Alnus* pollen grains were directly sampled from catkins on a tree growing at the Swiss Federal Office of
63 Meteorology and Climatology on a sunny morning on February 28 2019. Light scattering measurements were
64 taken using the fresh pollen grains soon after they were collected. The remaining pollen grains were stored in
65 tubes and scattering patterns were reevaluated after storing them for 1 h, 2 h, 6 h, and 10 days. Multiple
66 comparisons using the Bonferroni method were performed on the side and forward scattering data to assess
67 whether the light scattering distributions showed changes after storage. Bonferroni method is a multiple
68 comparison method used for non-parametric data sets. In order to carry out the multiple comparison, 316 scattering
69 data of each taxa were picked up because the Bonferroni method requires the same amount of data of each taxa
70 and 316 scattering data was the smallest amount of data amongst each time step (10 day).

71 **2.2 Light scattering patterns of different pollen taxa**

72 Dried pollen grains from *Alnus*, *Ambrosia*, *Artemisia*, *Betula*, *Castanea*, *Cedrus*, *Corylus*, *Fagus*, *Fraxinus*,
73 *Helianthus*, *Olea*, *Phleum*, *Quercus*, *Taxus*, and *Zea* were sampled in a similar way. These taxa are representative
74 of the pollen types commonly observed in Europe. After collecting the light scattering distributions of each pollen
75 type, multiple comparisons using the Bonferroni method were performed to evaluate whether these distributions

76 differ significantly from each other. In order to carry out the multiple comparison, 210 scattering data of each taxa
 77 were picked up based on the smallest amount of data amongst the taxon (*Helianthus*).

78 2.3 Automatic discrimination theory

79 To carry out simple and fast automatic pollen discrimination, the number of pollen grains of each type from the
 80 total number of pollen grains was calculated as follows.

81 For two different types of pollen (A and B) in the side scattering intensity range $a - b$ and in the forward
 82 scattering intensity range $c - d$, the following equation holds:

$$\begin{aligned}
 \int_a^b P_{A_{side}(x)} dx &= p_{A_{side}} \\
 \int_a^b P_{B_{side}(x)} dx &= p_{B_{side}} \\
 \int_c^d P_{A_{front}(x)} dx &= p_{A_{front}} \\
 \int_c^d P_{B_{front}(x)} dx &= p_{B_{front}}
 \end{aligned} \tag{1}$$

83 where P is the representative probability density function of the scattering intensity. p is the representative
 84 probability of the scattering intensity of each pollen grain lying in the integration intervals.

85 Next, the scattering intensity distribution that gives the number of pollen grains at each scattering intensity was
 86 fitted to a distribution function. In this experiment, the normal distribution was fitted to the number of pollen
 87 grains in every 100 mV steps. The gaussian function is written as:

$$f(x) = \frac{\alpha}{\sqrt{2\pi}} \exp\left\{-\frac{(x - \mu)^2}{2\sigma^2}\right\} + c \tag{2}$$

88 where α and c are coefficients, μ is the mean, σ is the standard deviation.

89 Fitting the data to the normal distribution function enables one to calculate the probability of a pollen grain
 90 showing a certain light scattering intensity. The probability density of the normal distribution function (P) is
 91 written as:

$$P(x) = \frac{1}{\sqrt{2\pi\sigma^2}} \exp\left\{-\frac{(x - \mu)^2}{2\sigma^2}\right\} \tag{3}$$

92 Fitting was performed by nonlinear optimization. The normal distribution was chosen so that we can handle the
 93 light scattering plots using a known function.

94 Equation (1) gives

$$\begin{aligned}
 C_1 p_{A_{side}} N_A + C_2 p_{B_{side}} N_B &= n_{side\ a-b} \\
 C_3 p_{A_{front}} N_A + C_4 p_{B_{front}} N_B &= n_{front\ c-d} \\
 N_A + N_B &= N_{total}
 \end{aligned} \tag{5}$$

95 Here, N is the number of sampled pollen grains of each pollen type, which are the values to be calculated. N_{total}
 96 is the total number of sampled pollen grains and n is the total number of sampled pollen grains in the integration
 97 interval, which are known numbers. C is the correction factor defined by the following equation:

$$\begin{aligned}
 C &= \frac{\int_{-\infty}^{+\infty} P(x) dx}{\int_0^{4500} P(x) dx} \\
 &= \frac{1}{\int_0^{4500} P(x) dx}
 \end{aligned} \tag{6}$$

98 C is needed for renormalization of the probability distribution because the device KH-3000-01 is able to detect
99 the scattering intensity only in the range of 0–4500mV.

100 By solving two equations in Eq. (5), N_A and N_B will be theoretically estimated.

101 In this paper, *Alnus* and *Artemisia* were chosen as examples to evaluate the usability of the theory above. Because
102 fitting worked well in the range of 600–800mV for the side scattering and 300–500mV for the forward scattering,
103 $a = 600$, $b = 800$, $c = 300$ and $d = 500$ were substituted in Eq. (5). The evaluation tests were carried out five
104 times using the light scattering data for both *Alnus* and *Artemisia* (Fig. 2).

105 The magnitude of the estimation error is calculated as follows.

$$\text{error (\%)} = \frac{|\text{actual} - \text{estimation}|}{\text{actual}} \times 100 \quad (7)$$

106

107

108 3 Results

109 3.1 Temporal changes in light scattering pattern

110 The scattering distribution of *Alnus* pollen (Fig. 3) showed no significant temporal changes in scattering
111 distributions in 10 day (Table 1).

112 3.2 Light scattering distributions of different pollen taxa

113 Pollen grains with smaller sizes tend to show smaller voltage values (Fig. 4). The results of the multiple
114 comparisons (Table 2) indicated that there is always a significant different between side and forward scattering
115 between two different pollen types except between:

116 Side scattering: *Alnus-Ambrosia*, *Alnus-Corylus*, *Alnus-Olea*, *Ambrosia-Fraxinus*, *Betula-Phleum*, *Betula-*
117 *Quercus*, *Corylus-Olea*, *Fagus-Zea*, *Artemisia-Fraxinus*, *Helianthus-Zea*, *Phleum-Quercus*

118 Forward scattering: *Alnus-Corylus*, *Alnus-Quercus*, *Ambrosia-Artemisia*, *Ambrosia-Fraxinus*, *Artemisia-*
119 *Fraxinus*, *Betula-Phleum*, *Betula-Quercus*, *Castanea-Olea*, *Cedrus-Helianthus*, *Corylus-Quercus*, *Fagus-*
120 *Helianthus*, *Fagus-Zea*, *Phleum-Quercus*

121 3.3 Automatic counting

122 Counting the number of pollen grains of each type can be carried out by solving the two equations from Eq. (5),
123 side ($n_{side\ a-b}$) and forward ($n_{front\ c-d}$), side ($n_{side\ a-b}$) and total (N_{total}), forward ($n_{front\ c-d}$) and total
124 (N_{total}). The parameters of the probability density curve of the side and the forward (Fig. 5) light scattering
125 distributions of *Alnus* and *Artemisia* were estimated as follows:

$$126 P_{Alnus_{side}}: (\alpha, \mu, \sigma, c) = (434, 555, 224, 14.7)$$

$$127 P_{Artemisia_{side}}: (\alpha, \mu, \sigma, c) = (589, 419, 193, 10.3)$$

$$128 P_{Alnus_{front}}: (\alpha, \mu, \sigma, c) = (600, 349, 160, 16.3)$$

$$129 P_{Artemisia_{front}}: (\alpha, \mu, \sigma, c) = (1029, 203, 107, 13.0)$$

130 The results (Fig. 6) show that the estimated number of pollen grains had average errors of 47%, 34%, 39% for
131 *Alnus* and 31%, 19%, 21% for *Artemisia* (Table 3).

132

133 4 Discussion

134 Temporal changes in the shapes of pollen grains are expected to affect the changes in light scattering patterns.
135 However, our experimental data indicate that light scattering patterns show little to no changes over time (up to
136 at least 10 days). Thus, there should be no problem using pollen grains that are either fresh or have been stored
137 for several days for studies with the KH-3000. Further investigation is required to understand whether this is true

138 for species other than *Alnus* and for longer periods of time. Understanding the morphological stability of each
139 pollen type would be helpful to understand the temporal stability of light scattering patterns.

140 Light scattering data from various pollen taxa indicate that it is not possible to discriminate between the side
141 scattering patterns of *Alnus* vs *Ambrosia*, *Alnus* vs *Corylus*, *Alnus* vs *Olea*, *Ambrosia* vs *Fraxinus*, *Betula* vs
142 *Phleum*, *Betula* vs *Quercus*, *Corylus* vs *Olea*, *Fagus* vs *Zea*, *Artemisia* vs *Fraxinus*, *Helianthus* vs *Zea*, *Phleum*
143 vs *Quercus* and the forward scattering patterns between *Alnus* vs *Corylus*, *Alnus* vs *Quercus*, *Ambrosia* vs
144 *Artemisia*, *Ambrosia* vs *Fraxinus*, *Artemisia* vs *Fraxinus*, *Betula* vs *Phleum*, *Betula* vs *Quercus*, *Castanea* vs *Olea*,
145 *Cedrus* vs *Helianthus*, *Corylus* vs *Quercus*, *Fagus* vs *Helianthus*, *Fagus* vs *Zea*, , and *Phleum* vs *Quercus*, all of
146 which show similar scattering intensities. Although it is not clear if the classification theory introduced above is
147 applicable to these groups, the theory should be applicable to other pairs as long as they have different scattering
148 intensity distributions.

149 The estimation of the pollen counts of *Alnus* and *Artemisia* had average errors of approximately 40% and 23%,
150 respectively. Test 4 had the largest error, with approximately 134% for *Alnus* and approximately 44% for
151 *Artemisia*, which increased the average error. It is difficult to identify an obvious reason for these large values,
152 but it is possible that the pollen samples were contaminated by dusts or pollen grains picked up for this experiment
153 were biased in size or shape. Additionally, other estimations derived from the fitted curve of the forward and the
154 side scattering distributions showed that even when the pollen counts are estimated only from scattering intensity
155 data without using total number of pollen grains, which is a known number, the pollen counts are able to be
156 calculated accurately. The KH-3000-01 has been widely used to estimate airborne concentrations of *Cryptomeria*
157 *japonica*. In this study, we found average errors of 20-40% for *Alnus* and *Artemisia*, values which are also likely
158 applicable to other taxa such as *Cryptomeria japonica*. Other taxa should, however, be investigated in future.

159 Pollen counts can be estimated by solving Eq. (5), which contains three equations, meaning that it is possible to
160 make estimates for three different pollen taxa simultaneously. If more integration intervals were picked up from
161 the probability density curve of the scattering intensity and added to the equation, in theory it would be possible
162 to count more pollen taxa. It is possible, however, that the accuracy of the estimated values might decline due to
163 the accuracy of the fitted curve. Therefore, narrowing down a target to two or three pollen types considering the
164 season should be helpful to make accurate automatic counts of several pollen taxa simultaneously.

165 In this study, the normal distribution function was chosen for fitting because of its universal property. However,
166 further consideration is required to determine the best function for fitting actual light scattering characteristics.

167

168 **5 Conclusion**

169 By applying the statistical analysis method, the Bonferroni method to the scattering patterns of *Alnus* at each time
170 step, our experiment showed that there seems to be no significant temporal changes in the light scattering patterns.
171 We also confirmed that different pollen types do not always have different light scattering patterns. However,
172 when two different pollen types have different light scattering patterns, it was possible to calculate the number of
173 pollen grains of each taxa using these light scattering patterns by solving the probability density function of the
174 pattern.

175

176 Code/Data availability: The authors confirm that the data supporting the findings of this study are available
177 within the article.

178

179 Author contributions: Kenji Miki established the system, performed the data analysis, and wrote the manuscript.
180 Shigeto Kawashima arranged the experimental setup and proofread the manuscript.

181 Conflict of interest: The authors declare that they have no conflict of interest.

182

183 Acknowledgement

184 This research was supported by the Young Research Exchange Programme between Japan and Switzerland
185 under the Japan-Swiss Science and Technology Programme. The authors would like to express their gratitude to

186 Yamatronics. The authors also would like to show their appreciation to Mr. Samuel Allen for proofreading our
187 manuscript.

188

189 **References**

190 Boucher, A., Hidalgo, P.J., Thonnat, M., Belmonte, J., Galan, C., Bonton, P., and Tomczak, R.: Development of
191 a semi-automatic system for pollen recognition. *Aerobiologia*, 18, 195–201, 2002.

192 Buters, J.T.M., Antunes, C., Galveias, A., Bergmann, K.C., Thibaudon, M., Galán, C., Schmidt-Weber, C., and
193 Oteros, J.: Pollen and spore monitoring in the world. *Clin. Transl. Allergy*, 8, doi.org/10.1186/s13601-
194 018-0197-8, 2018.

195 Chen, C., Hendricks, E.A., Duin, R.P.W., Reiber, J.H.C., Hiemstra, P.S., de Weger, L.A., and Stoel, B.C.:
196 Feasibility study on automated recognition of allergenic pollen: grass, birch and mugwort. *Aerobiologia*,
197 22, 275–284. doi:10.1007/s10453-006-9040-0, 2006.

198 Crouzy, B., Stella, M., Konzelmann, T., Calpini, B., and Clot, B.: All-optical automatic pollen identification:
199 Towards an operational system. *Atmos. Environ.*, 140, 202–212, 2016.

200 France, I. Duller, A.W.G., Duller, G.A.T., and Lamb, H.F.: A new approach to automated pollen analysis. *Quat.*
201 *Sci. Rev.*, 19, 537–546, 2000.

202 Gallardo-Caballero, R., García-Orellana, C. J., García-Manso, A., González-Velasco, H., Tormo-Molina, R., and
203 Macías-Macías, M.: Precise pollen grain detection in bright field microscopy using deep learning
204 technique. *Sensors*, 19, 3583. doi:10.3390/s19163583, 2019.

205 Gonçalves, A.B., Souza, J.S., de Silva, G.G., Cereda, M.P., Pott, A., Naka, M.H., and Pistori, H.: Feature
206 extraction and machine learning for the classification of Brazilian savannah pollen grains. *PLoS ONE*,
207 11, e0157044. doi:10.1371/journal.pone.0157044, 2016.

208 Gottardini, E., Rossi, S., Cristofolini, F., and Benedetti, L.: Use of Fourier transform infrared (FT-IR)
209 spectroscopy as a tool for pollen identification. *Aerobiologia*, 23, 211–219, 2007.

210 Huffman, D. R., Swanson, B. E., and Huffman, J. A.: A wavelength-dispersive instrument for characterizing
211 fluorescence and scattering spectra of individual aerosol particle on a substrate. *Atmos. Meas. Tech.*, 9,
212 3987–3998, 2016.

213 Iwai, T.: Polarization analysis of light scattered by pollen grains of *Cryptomeria japonica*. *Jpn. J. Appl. Phys.*, 52,
214 062404. doi.org/10.7567/JJAP.52.062404, 2013.

215 Kaya, Y., Mesut Pinar, S., Emre Erez, M., Fidan, M., and Riding, J.B.: Identification of *Onopordum* pollen using
216 the extreme learning machine, a type of artificial neural network. *Palynology*, 38, 129–137.
217 doi.org/10.1080/09500340.2013.868173, 2014.

218 Kawashima, S., Clot, B., Fujita, T., Takahashi, Y., and Nakamura, K.: An algorithm and a device for counting
219 airborne pollen automatically using laser optics. *Atmos. Environ.*, 41, 7987–7993, 2007.

220 Kawashima, S., Thibaudon, M., Matsuda, S., Fujita, T., Lemonis, N., Clot, B., and Oliver, G.: Automated pollen
221 monitoring system using laser optics for observing seasonal changes in the concentration of total airborne
222 pollen. *Aerobiologia*, 33, 351–362, 2017.

223 Landsmeer, S.H., Hendricks, E.A., De Weger L.A., Reiber, J.H.C., and Stoel, B.C.: Detection of pollen grains in
224 multifocal optical microscopy images of air samples. *Microsc. Res. Tech.*, 72, 424–430, 2009.

225 Li, P., Treloar, W.J., Flenley, J.R., and Empson, L.: Towards automation of palynology 2: the use of texture
226 measures and neural network analysis for automated identification of optical images of pollen grains. *J.*
227 *Quat. Sci.*, 19, 755–762. doi: 10.1002/jqs.874, 2004.

228 Longhi, S., Cristofori, A., Gatto, P., Cristofolini, F., and Grando, M.S., Gottardini, E.: Biomolecular identification
229 of allergenic pollen: a new perspective for aerobiological monitoring? *Ann. Allergy Asthma Immunol.*,
230 103, 508–514, 2009.

- 231 Marcos, J.V., Nava, R., Cristóbal, G., Redondo, R., Escalante-Ramírez, B., Bueno, G., Déniz, Ó., González-Porto,
232 A., Pardo, C., Chung, F., and Rodríguez, T.: Automated pollen identification using microscopic imaging
233 and texture analysis. *Micron*, 68, 36–46. doi.org/10.1016/j.micron.2014.09.002, 2015.
- 234 Matsuda, S., and Kawashima, S.: Relationship between laser light scattering and physical properties of airborne
235 pollen. *J. Aerosol Sci.*, 124, 122–132, 2018.
- 236 Miki, K., Kawashima, S., Fujita, T., Nakamura, K., and Clot, B.: Effect of micro-scale wind on the measurement
237 of airborne pollen concentrations using volumetric methods on a building rooftop. *Atmos. Environ.*, 158,
238 1–10. doi.org/10.1016/j.atmosenv.2017.03.015, 2017.
- 239 Miki, K., Kawashima, S., Clot, B., and Nakamura, K.: Comparative efficiency of airborne pollen concentration
240 evaluation in two pollen sampler designs related to impaction and changes in internal wind speed. *Atmos.*
241 *Environ.*, 203, 18–27, doi.org/10.1016/j.atmosenv.2019.01.039, 2019.
- 242 Mitsumoto, K., Yabusaki, K., and Aoyagi, H.: Classification of pollen species using autofluorescence image
243 analysis. *J. Biosci. Bioeng.*, 107, 90–94, 2009.
- 244 Mitsumoto, K., Yabusaki, K., Kobayashi, K., Aoyagi, H.: Development of a novel real-time pollen-sorting counter
245 using species-specific pollen autofluorescence. *Aerobiologia*, 26, 99–111, doi10.1007/s10453-009-
246 9147-1, 2010.
- 247 O'Connor, D.J., Healy, D.A., and Sodeau, J.R.: The on-line detection of biological particle emissions from selected
248 agricultural materials using the WIBS-4 (Waveband Integrated Bioaerosol Sensor) technique. *Atmos.*
249 *Environ.*, 80, 415–425, doi.org/10.1016/j.atmosenv.2013.07.051, 2013.
- 250 Oteros, J., Pusch, G., Weichenmeier, I., Heimann, U., Möller, R., Röseler, S., Traidl-Hoffmann, C., Schmidt-
251 Weber, C., and Buters, J.T.M.: Automatic and online pollen monitoring. *Int. Arch. Allergy Immunol.*,
252 167, 158–166, doi: 10.1159/000436968, 2015.
- 253 Punyasena, S.W., Tchong, D.K., Wesseln, C., and Mueller, P.G.: Classifying black and white spruce pollen using
254 layered machine learning. *New Phytol.*, 196, 937–944, doi: 10.1111/j.1469-8137.2012.04291.x, 2012.
- 255 Ranzato, M., Taylor, P.E., House, J.M., Flagan, R.C., LeCun, Y., and Perona, P.: Automatic recognition of
256 biological particles in microscopic images. *Pattern Recognit. Lett.*, 28, 31–39,
257 doi:10.1016/j.patrec.2006.06.010, 2007.
- 258 Richardson, S.C., Mytilinaios, M., Foskinis, R., Kyrou, C., Papayannis, A., Pyrri, I., Giannoutsou, E., and
259 Adamakis, I.D.S.: Bioaerosol detection over Athens, Greece using the laser induced fluorescence
260 technique. *Sci. Total Environ.*, 696, 133906, doi.org/10.1016/j.scitotenv.2019.133906, 2019.
- 261 Rodríguez-Damián, M., Cernadas, E., Formella, A., Fernández-Delgado, M., and De Sá-Otero, P.: Automatic
262 detection and classification of grains of pollen based on shape and texture. *IEEE Trans. Syst. Man Cybern.*
263 *Syst.*, 36, 531–542, doi:10.1109/TSMCC.2005.855426, 2006.
- 264 Šaulienė, I., Šukienė, L., Daunys, G., Valiulis, G., Vaitkevičius, L. Matavulj, P., Brdar, S., Panic, M., Sikoparija,
265 B., Clot, B., Crouzy, B., and Sofiev, M.: Automatic pollen recognition with the Rapid-E particle counter:
266 the first-level procedure, experience and next steps. *Atmos. Meas. Tech.*, 12, 3435–3452, 2019.
- 267 Surbek, M., Esen, C., Schweiger, G., and Ostendorf, A.: Pollen characterization and identification by elastically
268 scattered light. *J. Biophotonics*, 4, 49–56, doi:10.1002/jbio.200900088, 2011.
- 269 Swanson, B.E., and Huffman, J.A.: Development and characterization of an inexpensive single-particle
270 fluorescence spectrometer for bioaerosol monitoring. *Opt. Express*, 26, 3646–3660, 2018.
- 271 Takahashi, Y., Kawashima, S., Fujita, T., Ito, C., Togashi, R., and Takeda, H.: Comparison between real-time
272 pollen monitor KH-3000 and Burkard sampler. *Arerugi*, 50, 1136–1142, 2011.

- 273 Tcheng, D.K., Nayak, A.K., Fowlkes, C.C., and Punyasena, S.W.: Visual recognition software for binary
274 classification and its application to spruce pollen identification. PLoS ONE, 11, e0148879.
275 doi:10.1371/journal.pone.0148879, 2016.
- 276 Wang, Q., Nakamura, S., Gong, S., Suzuki, M., Nakajima, D., Takai, Y., Lu, S., Sekiguchi, K., and Miwa, M.:
277 Release behaviour of *Cryptomeria japonica* pollen allergenic cry j1 and cry j2 in rainwater containing
278 air pollutants. Int. J. Sustain. Dev. Plann., 9, 42–53, doi:10.2495/SDP-V9-N1-42-53, 2014.
- 279 Zhang, Y., Fountain, D.W., Hodgson, R.M., Flenley, J.R., and Gunetileke, S.: Towards automation of palynology
280 3: pollen pattern recognition using Gabor transforms and digital moments. J. Quat. Sci., 19, 763–768,
281 doi: 10.1002/jqs.875, 2004.
282

283 Figure 1 Schematic drawing of device setup. Laser irradiates pollen particle inside the KH-3000-01.

284 Figure 2 Light scattering distribution data from *Alnus* and *Artemisia* used for estimation test.

285 Figure 3 Light scattering plots for *Alnus* pollen – fresh and after 1h, 2h, 6h, and 10 days storage.

286 Figure 4 Light scattering distribution of various pollen taxa.

287 Figure 5 Fitted curve for side scattering (top row) and probability density curve (second row) for *Alnus* (left)

288 and *Artemisia* (right) and fitted curve for forward scattering (third row) and probability density curve (bottom

289 row) for *Alnus* (left) and *Artemisia* (right).

290 Figure 6 Results of automatic counting of *Alnus* and *Artemisia*. Red and black dots represent actual and

291 estimated numbers of pollen grains, respectively. The pair of red and black dots with the same shape are in the

292 same test set.

293

294 Table 1 Multiple comparison between *Alnus* data stored for various periods.

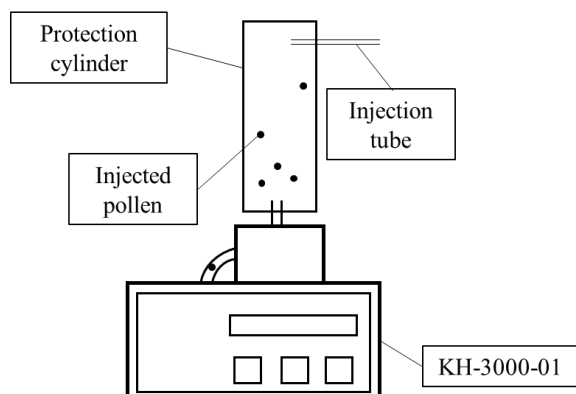
295 Table 2 Multiple comparison between each pollen taxon

296 Table 3 Results of estimation of number of pollen grains of *Alnus* and *Artemisia* and errors of each estimation.

297

298

299

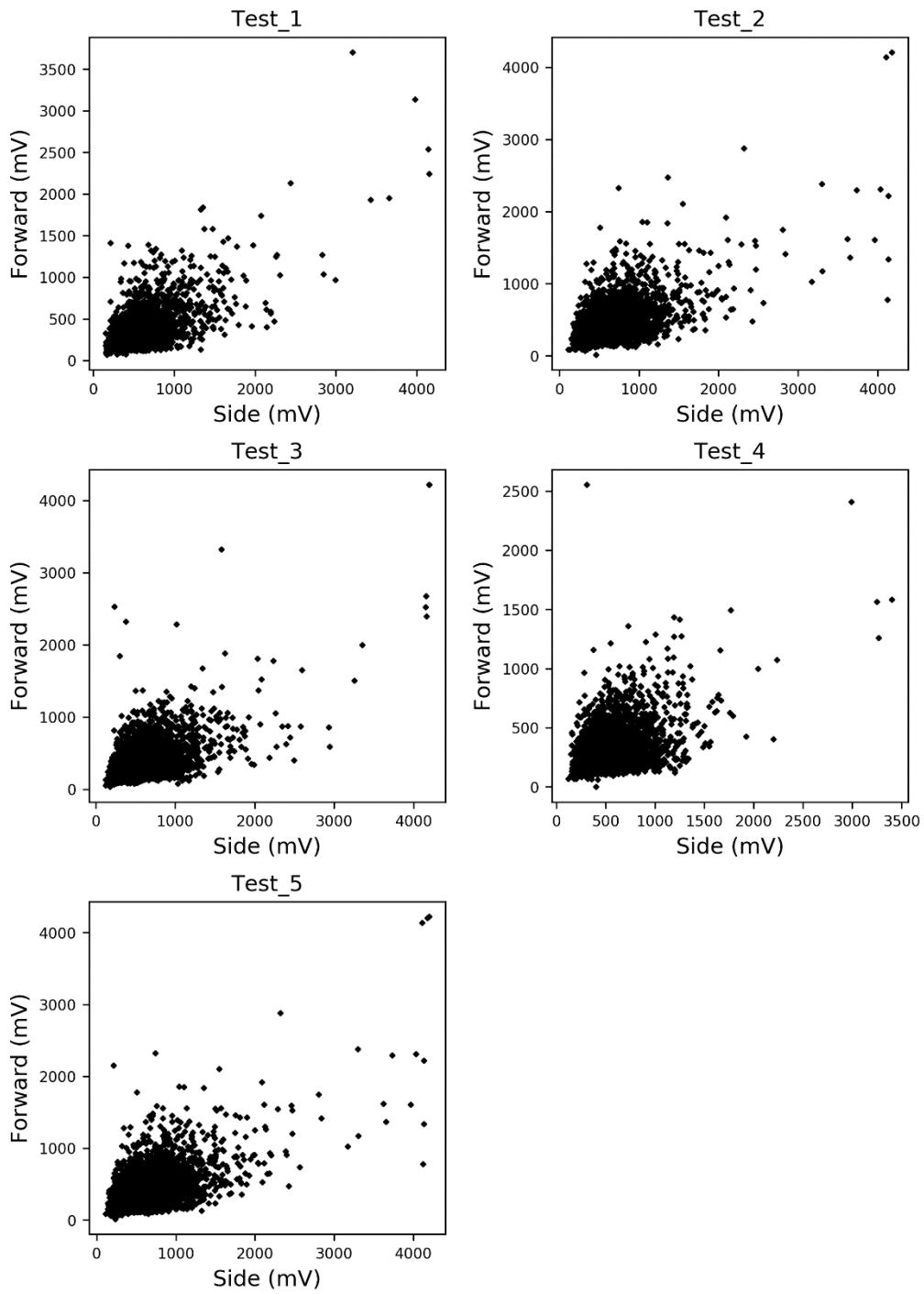


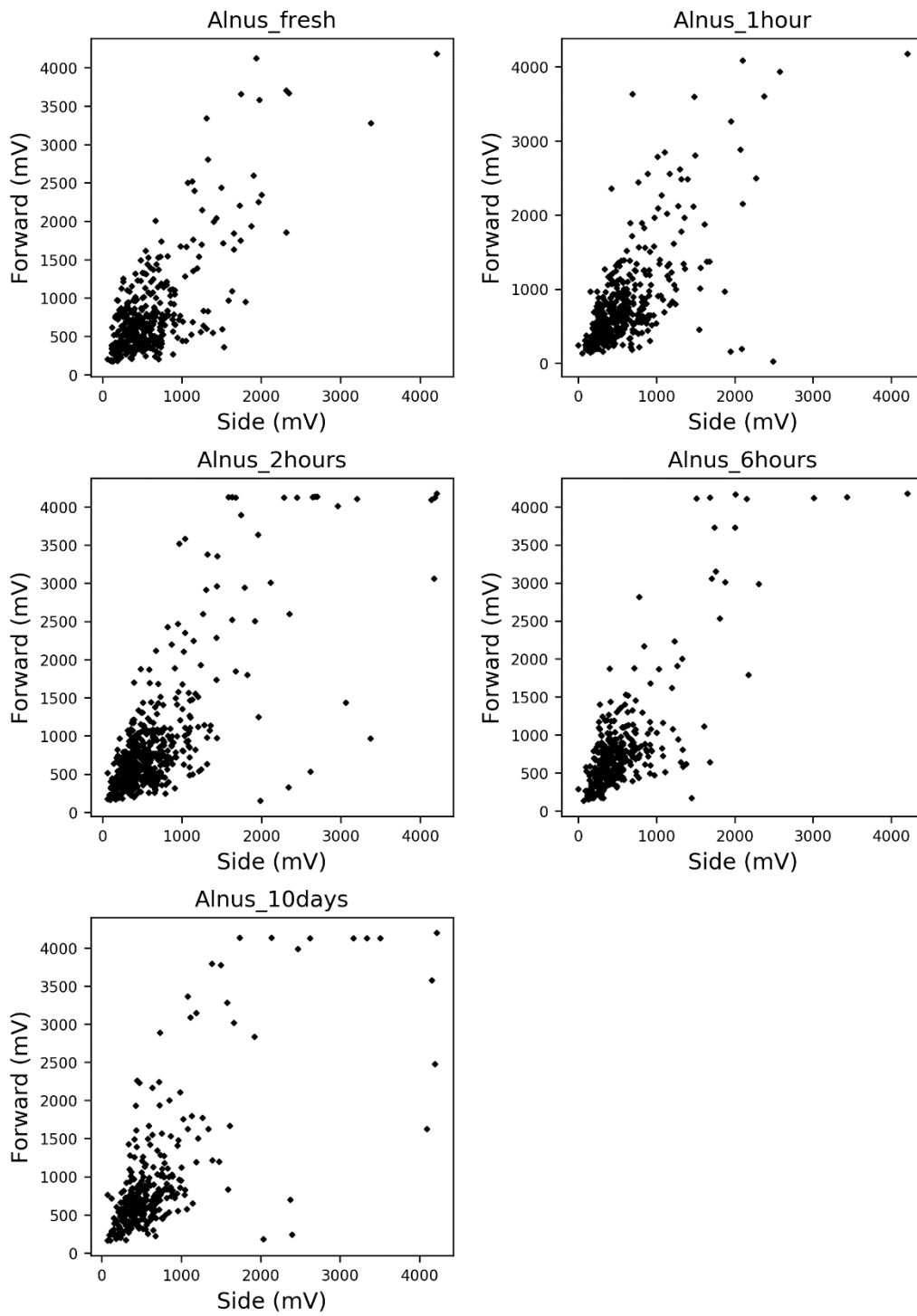
300

301 Fig. 1

Miki and Kawashima

302





310

311

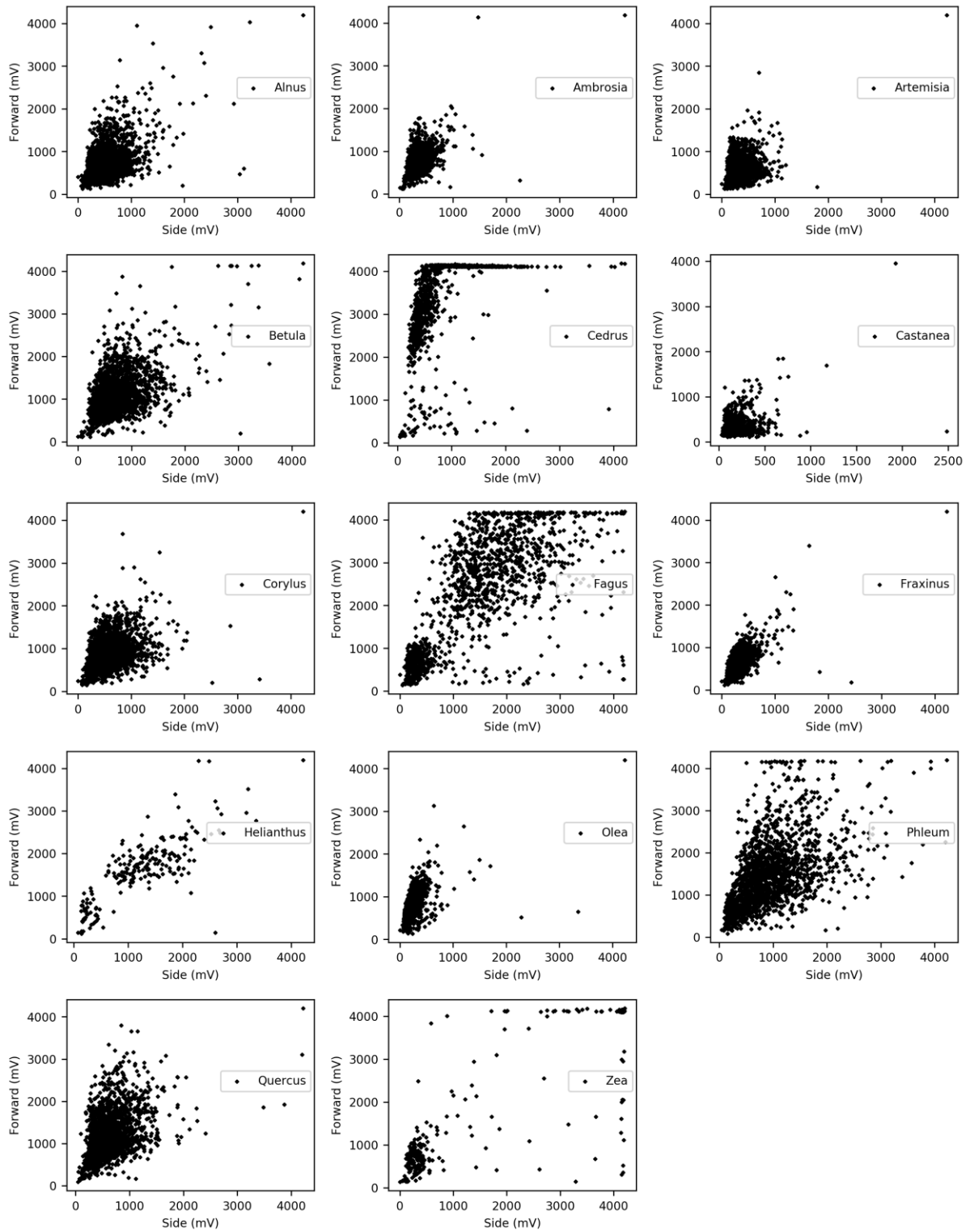
312

313 Fig.3

314

315

Miki and Kawashima



316

317

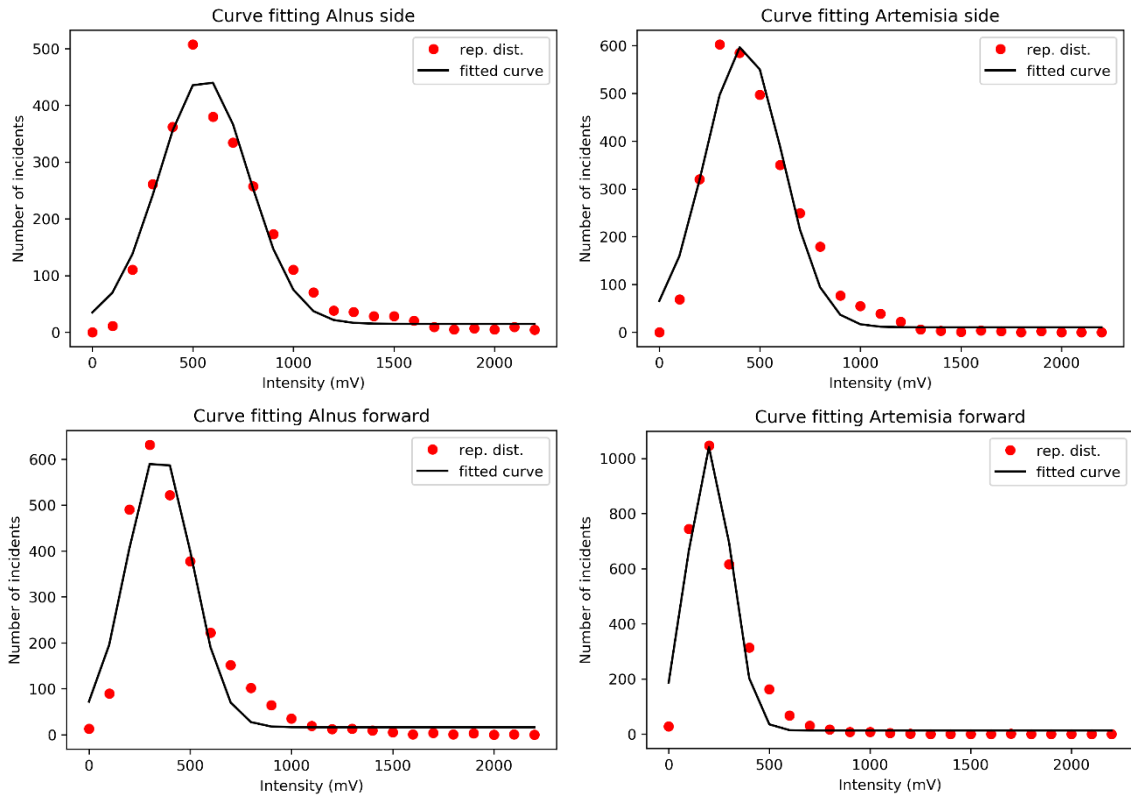
318 Fig.4

319

320

Miki and Kawashima

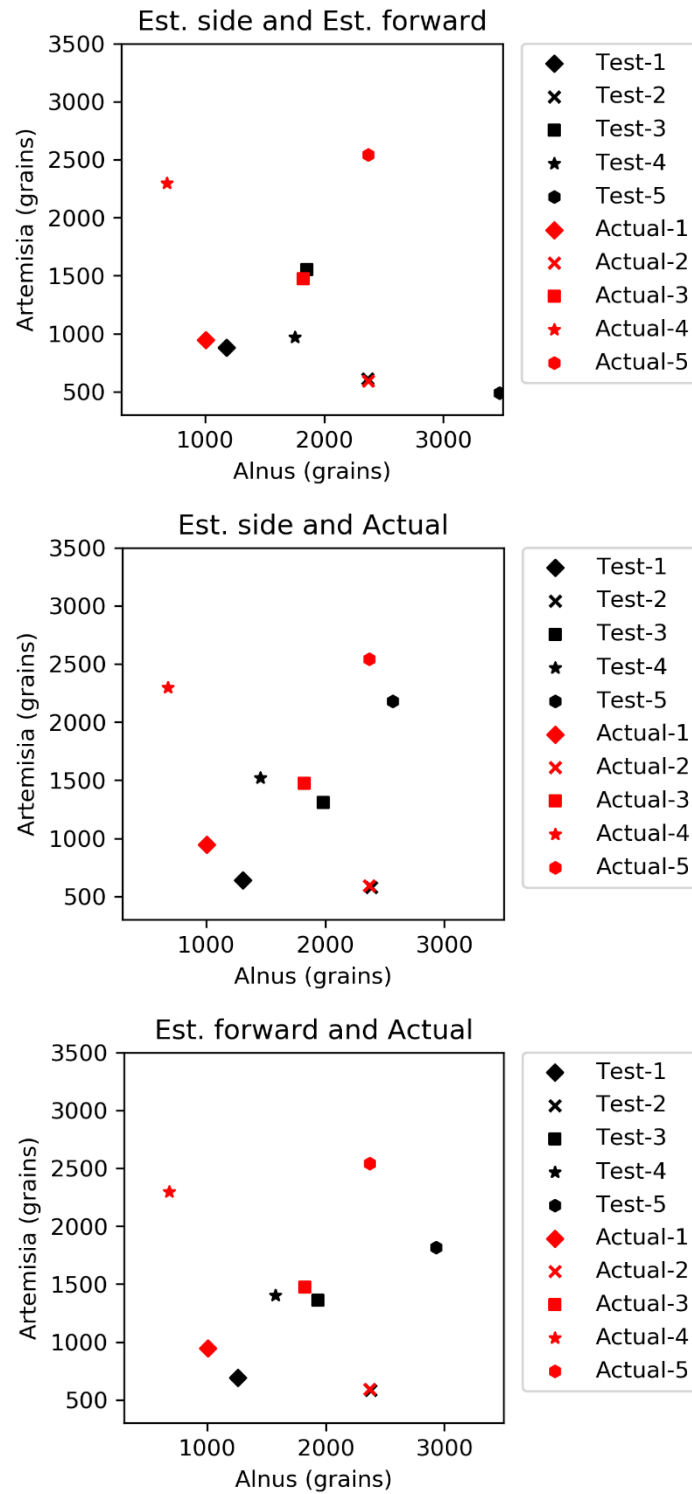
321



322

323 Fig.5

Miki and Kawashima



325

326 Fig.6

Miki and Kawashima

327

328

329

330

331

332

Table 1 Multiple comparisons between each time step (*Alnus*)

333

Side				
	1hour	2hour	6hour	10day
fresh	1.00	0.38	1.00	1.00
1hour	—	1.00	1.00	1.00
2hour	—	—	0.71	1.00
6hour	—	—	—	1.00

Forward				
	1hour	2hour	6hour	10day
fresh	1.00	1.00	1.00	1.00
1hour	—	1.00	0.84	1.00
2hour	—	—	1.00	1.00
6hour	—	—	—	0.31

334

335

336

337

Miki and Kawashima

338

339

340

341

342

343

Table 2 Multiple comparisons between each pollen taxon

Side	<i>Ambrosia</i>	<i>Artemisia</i>	<i>Betula</i>	<i>Castanea</i>	<i>Cedrus</i>	<i>Corylus</i>	<i>Fagus</i>	<i>Fraxinus</i>	<i>Helianthus</i>	<i>Olea</i>	<i>Phleum</i>	<i>Quercus</i>	<i>Zea</i>
<i>Alnus</i>	0.34	*	*	*	*	1.00	*	*	*	1.00	*	*	*
<i>Ambrosia</i>	—	*	*	*	*	*	*	0.08	*	*	*	*	*
<i>Artemisia</i>	—	—	*	*	*	*	*	0.06	*	*	*	*	*
<i>Betula</i>	—	—	—	*	*	*	*	*	*	*	0.06	1.00	*
<i>Castanea</i>	—	—	—	—	*	*	*	*	*	*	*	*	*
<i>Cedrus</i>	—	—	—	—	—	*	*	*	*	*	*	*	*
<i>Corylus</i>	—	—	—	—	—	—	*	*	*	0.49	*	*	*
<i>Fagus</i>	—	—	—	—	—	—	—	*	*	*	*	*	0.59
<i>Fraxinus</i>	—	—	—	—	—	—	—	—	*	*	*	*	*
<i>Helianthus</i>	—	—	—	—	—	—	—	—	—	*	*	*	1.00
<i>Olea</i>	—	—	—	—	—	—	—	—	—	—	*	*	*
<i>Phleum</i>	—	—	—	—	—	—	—	—	—	—	—	1.00	*
<i>Quercus</i>	—	—	—	—	—	—	—	—	—	—	—	—	*

* $p < 0.05$

344

Forward	<i>Ambrosia</i>	<i>Artemisia</i>	<i>Betula</i>	<i>Castanea</i>	<i>Cedrus</i>	<i>Corylus</i>	<i>Fagus</i>	<i>Fraxinus</i>	<i>Helianthus</i>	<i>Olea</i>	<i>Phleum</i>	<i>Quercus</i>	<i>Zea</i>
<i>Alnus</i>	*	*	*	*	*	1.00	*	*	*	*	*	1.00	*
<i>Ambrosia</i>	—	0.95	*	*	*	*	*	1.00	*	*	*	*	*
<i>Artemisia</i>	—	—	*	*	*	*	*	1.00	*	*	*	*	*
<i>Betula</i>	—	—	—	*	*	*	*	*	*	*	1.00	1.00	*
<i>Castanea</i>	—	—	—	—	*	*	*	*	*	1.00	*	*	*
<i>Cedrus</i>	—	—	—	—	—	*	*	*	1.00	*	*	*	*
<i>Corylus</i>	—	—	—	—	—	—	*	*	*	*	*	1.00	*
<i>Fagus</i>	—	—	—	—	—	—	—	*	0.14	*	*	*	1.00
<i>Fraxinus</i>	—	—	—	—	—	—	—	—	*	*	*	*	*
<i>Helianthus</i>	—	—	—	—	—	—	—	—	—	*	*	*	*
<i>Olea</i>	—	—	—	—	—	—	—	—	—	—	*	*	*
<i>Phleum</i>	—	—	—	—	—	—	—	—	—	—	—	0.10	*
<i>Quercus</i>	—	—	—	—	—	—	—	—	—	—	—	—	*

* $p < 0.05$

345

346

347

348

349

Miki and Kawashima

350

351

352

353

354

355

356 Table 3 Results of estimation of number of pollen grains of *Alnus* and *Artemisia* and errors of each estimation.

		Test 1		Test 2		Test 3	
		<i>Alnus</i> (error)	<i>Artemisia</i> (error)	<i>Alnus</i> (error)	<i>Artemisia</i> (error)	<i>Alnus</i> (error)	<i>Artemisia</i> (error)
Estimation	Side and Forward	1183 (17%)	881 (6.8%)	2367 (0.17%)	612 (3.2%)	1855 (1.8%)	1552 (5.4%)
	Total and Side	1310 (30%)	642 (32%)	2386 (0.63%)	577 (2.7%)	1984 (8.8%)	1310 (11%)
	Total and Forward	1259 (25%)	694 (27%)	2378 (0.30%)	585 (1.4%)	1932(6.0%)	1362 (7.5%)
Actual		1008	945	2371	593	1823	1472

		Test 4		Test 5	
		<i>Alnus</i> (error)	<i>Artemisia</i> (error)	<i>Alnus</i> (error)	<i>Artemisia</i> (error)
Estimation	Side and Front	1753 (157%)	968 (58%)	3469 (57%)	489 (81%)
	Total and Side	1458 (114%)	1520 (34%)	2567 (16%)	2179 (14%)
	Total and Front	1577 (132%)	1402 (39%)	2929 (33%)	1817 (29%)
Actual		681	2297	2205	2542

357

358

359

360

361

Miki and Kawashima