



# 1 Dynamic Infrared Gas Analysis from Longleaf Pine Fuelbeds Burned in a Wind

## 2 Tunnel: Observation of Phenol in Pyrolysis and Combustion Phases

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## 9 **0. Abstract**

10 Pyrolysis is the first step in a series of chemical and physical processes that produce flammable organic gases from wildland fuels that can result in a wildland fire. We report results using a new 11 12 time-resolved Fourier transform infrared method that correlates the measured FTIR spectrum to an infrared thermal image sequence enabling identification and quantification of gases within 13 14 different phases of the fire process. The flame from burning fuel beds composed of pine needles (Pinus palustris) and mixtures of sparkleberry, fetterbush and inkberry plants was the natural heat 15 source for pyrolysis. Extractive gas samples were analyzed and identified in both static and 16 dynamic modes synchronized to thermal infrared imaging: A total of 29 gases were identified 17 18 including small alkanes, alkenes, aldehydes, nitrogen compounds and aromatics, most previously measured by FTIR in wildland fires. This study presents one of the first identifications of phenol 19 20 associated with both pre-combustion and combustion phases, using ca. 1 Hz resolution. 21 Preliminary results indicate ~2.5x greater phenol emission from sparkleberry and inkberry 22 compared to fetterbush, with differing temporal profiles.

Keywords: Fourier transform infrared, time-resolved, biomass burning, pyrolysis, phenol,
 benzene, naphthalene, Pinus palustris, Lyonia lucida, Ilex glabra, Vaccinium arboreum

## 25 **1. Introduction**

Wildland fire is an important component of many ecosystems and has been used by humans for
several thousand years (Crutzen and Goldammer, 1993; Pyne, 1997; Scott et al., 2014). Many
North American ecosystems have evolved as a result of persistent fire (Barbour and Billings,





29 2000). The importance of fire in pine forests worldwide including the southern U.S. is well-known 30 (Agee, 2000; Christensen, 2000). In the U.S., prescribed burning is used on approximately 8 million ha annually to accomplish a variety of forestry and agricultural objectives (Melvin, 2015); 31 the impact of smoke from these fires has been studied for over 50 years (Chi et al., 1979; Biswell, 32 1989; Ward and Hardy, 1991; Hardy et al., 2001;). In the southern U.S., forest management 33 objectives include hazardous fuel reduction, site preparation, improved wildlife habitat, insect and 34 disease control, enhanced appearance and perpetuation of fire dependent species and natural 35 communities (Carter and Foster, 2004; Waldrop and Goodrick, 2012). The U.S. Department of 36 Defense (DoD) uses prescribed burning on approximately 243,000 ha annually for many of these 37 objectives as well as maintenance of critical training areas (Cohen et al., 2014). Many land 38 managers rely on fire behavior models to calculate fire movement on the landscape, energy release, 39 smoke plume development, dispersion and content (Bytnerowicz et al., 2009, Paton-Walsh et al. 40 2014). However, few fire behavior models account for the plethora of chemical reactions involved 41 42 in the fire. The heat transfer processes that take place in the fire environment are also only coarsely described. In order to improve the use of prescribed burning to accomplish refined objectives, 43 more detailed description and modeling of the physical and chemical processes in fire are needed 44 (Cohen et al., 2014). 45

The chemical phases of wildland fire, described as preheating, flaming, smoldering and glowing 46 47 (Ward, 2001) are understood in a chemical sense, but only to varying degrees: While the chemical effluents of flaming and smoldering phases have been characterized for many ecosystems and fuel 48 49 types at different scales (Ward and Radke, 1993), the physics and chemistry of the preheating (pyrolysis) phase, have fewer studies beyond the bench scale (e.g. Depew et al., 1972; 50 Dimitrakopoulos, 2001; Susott, 1982; Tihay, 2010). To improve fire application models and to 51 accomplish the desired fire effects and limit potential fugitive emissions, improved understanding 52 is thus needed for many fundamental processes, particularly for pyrolysis and ignition in 53 heterogeneous fuel beds of live and dead fuels that reflect the diversity of vegetation found 54 55 worldwide. (Guérette et al. 2018).

Prior to oxidative combustion, biomass thermally decomposes in a heated environment. To study
this decomposition, thermogravimetric analysis has been applied to a small set of plant species
deemed to represent major wildland fuel types (e.g. Burgan and Susott, 1991; Susott, 1982). Others





59 have determined caloric content of southern fuels which is related to the composition of pyrolysis 60 products. (Hough, 1969; Behm et al., 2004). However, most such prior work used dried, ground fuel samples in either an inert or oxidizing environment subject to uniform heating and heat 61 transfer, (Kibet et al. 2012) thereby eliminating the effects of moisture and heat transfer which are 62 key fire behavior variables. While pyrolysis and combustion of wildland fuels is known to be a 63 complex process (Zhou and Mahalingam, 2001), they are often modeled using simple 64 approximations in the relevant computer codes using the dominant gases of H<sub>2</sub>, CO, CO<sub>2</sub> and CH<sub>4</sub>. 65 Heat transfer in a wildland setting is less efficient than in thermogravimetric analysis: The amount 66 and composition of pyrolyzed species produced depend strongly on heating rate and temperature 67 and typically consists of oxidized small-molecule gases such as CO, CO<sub>2</sub>, or H<sub>2</sub>O, as well as non-68 69 oxidized or partially oxidized species such as  $H_2$ ,  $CH_4$ ,  $C_xH_y$ ,  $C_xH_yO_z$  as well as tars. The products of primary pyrolysis may react in the gas phase at elevated temperatures (i.e., secondary pyrolysis), 70 71 which may affect the amount of tar remaining.

72 This work is part of a larger project to measure and model pyrolysis gases from common wildland 73 fuels found on DoD installations in the southern United States. (Weise et al. 2018). The project includes bench-level, laboratory-scale and field plot burns; integrating the results of the field and 74 75 laboratory measurements with the modeling results to identify potential improvements that can enhance our understanding of pyrolysis and ignition in wildland fuels. During the course of the 76 project Fourier transform infrared (FTIR) technology has been used on several occasions to non-77 78 intrusively measure the composition and concentration of the pyrolysis gases. This includes 79 identifying the gases liberated by: i) heating single leaf samples from several common southern 80 fuels using different heating modes in a pyrolyzer and in a simple flat-flame burner system, (Amini et al., 2019; Safdari et al., 2020) ii) heating shrubs in prescribed burns at Ft. Jackson, South 81 Carolina, (Scharko et al., 2019a, b) and iii) heating nursery plants with flames from longleaf pine 82 needle fuel beds inside a wind tunnel (Aminfar et al., 2019). In order to achieve the goal that the 83 results be applicable to prescribed burns, a key focus has been linking the bench scale, wind tunnel 84 and field data to the models using realistic values and identities for the pyrolysis gases. Chemical 85 analysis of the foliage and results of the bench scale tests so far suggest that describing wood 86 pyrolysis may not be suitable for foliage fuels (live and dead) (Jolly et al., 2012; Jolly et al., 2016; 87 Matt et al., 2020); to date pyrolysis and ignition of wildland fuels have typically been based on 88 results for only cellulose or wood (e.g. Varhegyi et al., 1994; Di Blasi, 2008). We have extended 89





the number of southern fuels examined under more realistic conditions (Amini et al., 2019; Safdari et al., 2018) including bench-scale measurements of burning individual leaves of plants reported in this paper. A subset of the bench-scale plant species has been burned in a small wind tunnel to bridge from the bench-scale to pyrolysis measurements made in the field. The wind tunnel measurements were set to emulate the larger scale experiments with FTIR instruments and canister samples in 0.1 ha prescribed burns at Ft. Jackson in May 2018.

96 Flames in laboratory and field experiments tend to be turbulent in nature meaning there is inevitably some cross-contamination of the pyrolysis gases with flame gases produced in the 97 98 combustion reactions. It is necessary to decouple the phases so as to better understand the discrete pyrolysis and combustion processes. The same can also be said of the many studies conducted at 99 larger facilities such as the Fire Sciences Laboratory (FSL) in Montana, USA. The FSL has long 100 path length optical cell coupled to an FTIR (Yokelson et al., 1996, Burling et al., 2010), as well as 101 102 many other powerful analytical methods such as proton-transfer mass spectrometry (Christian et al., 2004; Warneke et al., 2011; Yokelson et al., 2013). The FTIR and many other systems at the 103 Missoula FSL have made first detections for dozens of chemical species and pioneered the science 104 of biomass burning in many regards. But because the sampling platform is 4 m above the floor, 105 there is mixing of gases from different phases. The combustion and smoldering phases are typically 106 easier to differentiate, primarily via the intrinsic diagnostic of the modified combustion efficiency 107 108 (MCE, Ward and Hao, 1991) which is a measure that is not independent of the composition of 109 smoke (Weise et al 2020). The same ambiguity as to the nature of the phase of fire also applies to 110 extractive methods whereby a sampling device attempts to capture pre-combustion phase gases. Such sampling systems, typically connected to a field canister are effective but are subject to 111 vagaries of sniffer gas inlet placement, i.e. proximity to the pyrolyzing plant. (Scharko et al. 112 2019a,b). 113

In this paper we describe use of an FTIR with a dynamic probe to temporally isolate, identify and
quantify some of the early-stage/pyrolysis gases from burns at a mid-scale laboratory facility.
During three measurement episodes, experiments were conducted at the Riverside Fire Lab (RFL)
in a wind tunnel using fuel beds composed of longleaf pine needles and the live plants inkberry
(*Ilex glabra* (L.) A. Gray), fetterbush (*Lyonia lucida* (Lam.) K. Koch), sparkleberry (*Vaccinium arboreum* L.) and blueberry (*V. darrowii* Camp). Multiple methods were used such as quantum-





120 cascade lasers (Phillips et al., 2020), gas chromatography-mass spectrometry as well as FTIR with the overall objectives of: i) using careful chemometric extraction from the acquired data to see 121 what pyrolysis species can be identified by the techniques; ii) using the various methods to 122 123 determine the degree of oxidation or combustion, i.e. pyrolysis characterization; iii) making first attempts to quantify the rates of evolution of pyrolysis products for certain plant species; and 124 ideally; iv) determining if differences exist between the pyrolysis emissions / temporal profiles for 125 different plant species. We take advantage of the time-resolved capabilities, as well as high 126 resolution specificity offered by IR spectroscopy and couple these to the flame/solid fuel 127 temperature diagnostics of an IR camera to analyze the emissions from a subset of 21 RFL burns. 128

## 129 **2. Experimental**

## 130 2.1 Wind Tunnel and Experimental Configuration

During November 2017, February 2018 and November 2018 a total of 88 laboratory scale burns 131 were conducted at the USDA Forest Service Pacific Southwest Research Station in Riverside, 132 California. The laboratory includes a wind tunnel ca. 3 m long and 1 m wide which was set up to 133 simulate a forest floor of litter and live plants. Fuel beds composed of 1 kg of dry longleaf pine 134 135 needles and various combinations of fetterbush, sparkleberry, blueberry and inkberry (Fig. 1a) were burned under either "no wind" or 1 m s<sup>-1</sup> conditions. Fuel moisture content and mass loading, 136 ambient temperature and relative humidity in the tunnel were varied between experiments; fuel 137 138 beds were ignited with a line fire which propagated the length of the fuel bed as seen in Figure 1b. 139 Multiple analytical techniques were used to study the fire characteristics as well as the gas effluents: thermocouples, Schmidt-Boelter flux sensor, nadir thermal IR camera and background-140 oriented Schlieren photography (Aminfar et al 2019) to estimate heat transfer / air flow around the 141 plants, canister samples analyzed by GC/FID, quantum cascade (QC) infrared laser spectroscopy, 142 143 (Phillips et al., 2020) as well as broadband Fourier transform infrared (FTIR) spectroscopy. A schematic overview of the experimental setup is seen in Figure 1c. 144







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Figure 1. a) Overhead view of wind tunnel down its length with longleaf pine needles and interspersed
 inkberry plants; b) flame front progressing down the wind tunnel with FTIR extraction tube visible; c)
 cartoon (top view) of experimental layout with laser and FTIR systems.

149 Gas samples were pumped into the cell / FTIR instrument via a stainless steel probe mounted above the plants (Fig. 1b). Gas from additional probes was pumped into canisters for offline 150 151 analysis using gas chromatography. Sixty-six or seventy-four live plants were distributed within the longleaf pine needles in ceramic holders. Figure 1a shows the configuration of the fuel bed 152 with instrumentation for *in situ* analysis. Plant species were prepared on site and samples of dry 153 154 and live fuel were clipped to determine fuel moisture content prior to each burn set. The 155 experiments were set under varying fuel bed and environmental conditions. See Table 1 for the 156 conditions in the 21 experiments presented in this paper.

**Table 1.** Summary of burn schedule for November 2018 studies including burn number, date and time, fuel description, acquisition method and resolution used for wind tunnel experiments under 1 m s<sup>-1</sup> imposed air flow. Geometric mean flame spread rate =  $0.01 \text{ m s}^{-1}$ . The FTIR acquisition methods are described in the text.

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Burn number	Date (2018)	Local ignition time (PDT)	Local finish time (PDT)	Plant species	Acquistion method	Resolution (cm <sup>-1</sup> )
76	30-Oct	11:48:01	11:52:00	inkberry	static	0.6
77	30-Oct	14:19:10	14:23:37	fetterbush	dynamic	2.0
78	30-Oct	15:12:30	15:16:33	sparkleberry	static	0.6
79	30-Oct	16:17:00	16:21:10	inkberry	dynamic	2.0
80	31-Oct	9:32:00	9:35:45	sparkleberry	static	0.6
81	31-Oct	10:35:00	10:38:52	fetterbush	dynamic	1.0
82	31-Oct	11:30:30	11:35:15	sparkleberry	static	0.6
83	31-Oct	13:19:00	13:22:58	inkberry	dynamic	1.0
84	31-Oct	14:12:15	14:16:30	fetterbush	static	0.6
85	31-Oct	15:30:30	15:34:24	fetterbush	dynamic	2.0
86	1-Nov	9:30:00	9:33:02	sparkleberry	dynamic	1.0
87	1-Nov	10:40:00	10:42:49	inkberry	dynamic	1.0
88	1-Nov	11:40:00	11:42:59	fetterbush	static	0.6
89	1-Nov	13:35:00	13:38:48	inkberry	static	0.6
90	1-Nov	14:45:00	14:49:47	sparkleberry	static	0.6
92	2-Nov	9:30:00	9:34:05	inkberry	dynamic	0.6
93	2-Nov	10:41:15	10:45:44	fetterbush	dynamic	1.0
94	2-Nov	11:28:15	11:32:28	sparkleberry	static	0.6
95	2-Nov	13:42:45	13:46:17	sparkleberry	static	0.6
97	2-Nov	15:38:38	15:41:40	sparkleberry	dynamic	0.6

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### 163 2.2 Instrumentation

Gases were extracted from the burns via 3/8" stainless steel tubing, HEPA filtered to eliminate tar 164 and char contamination and pumped into an 8-meter White cell (Bruker A136, 2.2 liter volume)<sup>1</sup> 165 166 housed inside a Bruker Tensor 37 spectrometer (Figure 1c). The extractive probe was placed directly above a plant as close as possible to the foliage. To prevent analyte/tar condensation, both 167 transfer tubing and the gas cell were heated to ~50 °C using heating tape/voltage regulator and a 168 169 cell heating shroud, respectively. A thermocouple was suspended into the White cell to record the gas temperature for subsequent spectral analysis, with pressure gauge mounted atop the cell. Prior 170 171 to data collection, the White cell was aligned using the FTIR's Ge/CaF<sub>2</sub> beamsplitter and W-lamp source. Once aligned, these were replaced with a Ge/KBr beamsplitter and mid-IR globar source, 172 173 along with a mercury cadmium telluride detector, configuring the Tensor 37 to record spectral data from 7500 to 500 cm<sup>-1</sup>. 174

175 The FTIR system was tested for leaks, followed by a gas cell path length calibration using purified isopropyl alcohol (IPA - Sigma Aldrich 99.5%). Ten spectra with IPA pressures between 0.6 and 176 10.5 Torr were recorded to 0.1 Torr using an MKS KF15 pressure transducer. The integrated area 177 of the 3515-3290 cm<sup>-1</sup> spectral domain (Bruker OPUS 5.5 software) along with recorded 178 179 temperatures and pressures were used to create a Beer-Lambert plot (Scharko et al. 2019a). Using the integrals from the ten recorded spectra the cell path length was determined to be  $6.5 \pm 0.2$  m. 180 Wavelength calibration of the infrared data was achieved after the fact using a series of 30 water 181 rotational-vibrational lines from the PNNL gas-phase database. (Sharpe et al. 2002; Williams et 182 183 al., 2013). FTIR interferograms were acquired using double-sided, forward-backward acquisition; these were apodized using a Blackman-Harris 3-Term function and phase corrected with Mertz's 184 method prior to Fourier transformation. 185

186 2.3 Infrared spectral acquisition

187 Two data acquisition modes were used to analyze the burn gases: an extractive (or static) mode 188 and a dynamic mode. In the extractive mode the gas flowing through the White cell was isolated; 189 the inlet/outlet valves were simultaneously closed such that the emitted gases were isolated in the

<sup>&</sup>lt;sup>1</sup> The use of trade or firm names in this publication is for reader information and does not imply endorsement by the U.S. Department of Agriculture of any product or service.





190 cell at a desired pressure (ca. 740-700 Torr for high pressures, and 430-400 Torr for lower pressure 191 measurements). The valves were closed just prior to the flame front reaching the probe, attempting to capture pre-combustion phases including evaporation and pyrolysis. The goal of the extractive 192 193 mode was to obtain a higher fidelity "snapshot" for a given point in time of the burn; more data were averaged longer at higher spectral resolution allowing for detection of more gaseous species 194 with higher sensitivity. (Scharko et al., 2019a). The dynamic mode measurements had fewer scans 195 at lower resolution to capture changing chemical identities/composition corresponding to different 196 197 fire phases (pyrolysis, flaming combustion, smoldering combustion), achieving temporal resolutions of the order of ca. 1 Hz. 198

Of the 21 burns, 10 were recorded using the static method. The static experiment spectra were 199 200 recorded using the full resolution of the spectrometer (0.6 cm<sup>-1</sup>), a 2 mm Jacquinot stop and double 201 sided, forward-backward acquisition. Due to the higher resolution and lower light throughput, acquisition time was extended by averaging multiple scans for a full 30 minutes, resulting in vastly 202 improved signal/noise ratios (SNR). For analysis of such complicated gas-phase mixtures, infrared 203 spectral resolutions of 1.0 cm<sup>-1</sup> or better have been demonstrated to be advantageous (Burling et 204 al., 2010, Akagi et al., 2014, Scharko et al., 2019a). While one goal was to isolate gases to include 205 only the pyrolysis and pre-combustion phases, one vagary of the technique involved the timed 206 closing of the valves relative to arrival of the flame front approaching the inlet. If the valves were 207 shut too early, the captured emissions would consist of only (warmed) ambient gas before onset 208 of thermal degradation of the solid fuel, as opposed to the desired pyrolysis phase. Conversely, if 209 shut too late, flaming, or possibly even smoldering conditions would be sampled. 210

The second method was the dynamic mode whereby the OPUS software was used to continuously 211 collect interferograms throughout the duration of the burn, capturing the chemical compositions 212 213 associated with different phases, e.g. volatilization, heating, pyrolysis, flaming or smoldering combustion. Fourier transformation of the interferograms occurred after the burns to yield faster 214 acquisition times. The dynamic acquisition mode was used in combination with thermal IR video 215 imaging recorded from above the flame bed to help synchronize spectral acquisition to the various 216 217 burn phases for a total of 11 burns. Instead of averaging for 30 minutes, the dynamic method allowed for 40-80 continuous interferometer scans (differing on the duration of the burn), and 218 vielded a spectrum every 1.5 s for data taken at 1.0 cm<sup>-1</sup> resolution, every 0.79 s for data at 2.0 cm<sup>-1</sup> 219 <sup>1</sup> resolution, and every 2.5 s for  $0.6 \text{ cm}^{-1}$  resolution spectra. Data acquisition began as the flame 220





front encroached upon the extractive probe and continued until the flame had passed. Due to the faster acquisition rate these spectra are significantly noisier than the data collected using the extractive method. To compare results from the static and dynamic modes, fires 87 and 89 will be presented. The 2 m length fuel beds for both experiments 87 and 89 consisted of 1 kg longleaf pine needles with interspersed inkberry plants.

For time synchronization it was necessary to quantify the time lag from the time the emissions 226 enter the extractive probe to midpoint in their flow through the White cell. A flow rate test was 227 thus conducted using freon gas, CFC-11 (trichlorofluoromethane) which is comparable in 228 molecular weight to the heavier gases detected by the FTIR. Figure 2 shows such a test of CFC-229 11 being introduced with spectra recorded every 0.79 s using 2.0 cm<sup>-1</sup> resolution. The time from 230 231 introduction of the freon at the extractive probe (t=0, scan 0) to first appearance in scan 4 (maroon trace) was 3.2 seconds. The freon spectra had maximized at scan 6 (red trace) for a total  $\Delta t = 4.8$ 232 s lag from the probe to the instrument. With this information, FTIR time stamped data were then 233 adjusted to reflect the 4.8 second time delay which was used when correlating the spectral data to 234 235 the visual and thermal IR video images.



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Figure 2: Dynamic spectra recording introduction of CFC-11 from extractive probe to gas cell. Scan 0 (not shown) represents start of spectral acquisition/freon release near probe. Spectra produced every 0.79 s. First observation of freon occurs with scan 4; maximum absorbance of CFC-11 and stabilization occurs at scan 6.

A combination of software was used for the post-acquisition spectral analysis and confirmation of the species observed during the campaign. The MALT5 software (Griffith, 2016) utilizing both

<sup>240 2.4</sup> Spectral Analysis





HITRAN line-by-line data (Gordon et al., 2017) as well as the PNNL 50 °C gas-phase reference 243 spectra (Kochanov, 2019; Johnson et al., 2006, 2010) as input libraries was used to identify and 244 quantify vapor-phase chemicals in the spectra. Spectra were compiled into parameter files and 245 analyzed by the MALT software using parameters including pressure, temperature, pathlength, 246 resolution, as well as estimated initial values for chemical mixing ratios. The software generates a 247 spectrum to simulate the measured spectrum, adjusting mixing ratios until the residual between 248 the simulated and measured spectra is minimized. To confirm the species were actually present, 249 250 each spectrum generated by MALT was input to OPUS and subtracted from the measured spectrum; the target compound was purposefully omitted from the subtraction process to visually 251 inspect if the omitted compound was in fact present (see e.g. Figure 5). 252

253 **3. Results and Discussion** 

254 3.1 Analysis of Static Spectra

Ten spectra were recorded from different burns using the static mode with the gas cell valves 255 closed simultaneously; gases were sampled prior to arrival of the flame front (Figure 1c). A total 256 of 29 compounds were detected and confirmed using MALT5 and OPUS 5.5. Along with CO, 257 258 CO<sub>2</sub> and nitrogen compounds, the gas-phase species are largely lightweight hydrocarbons (HCs), volatile organic compounds (VOCs) and oxygenated volatile organic compounds (OVOCs). Table 259 2 provides a summary of all compounds observed during the static measurements and is broken 260 down into subcategories of chemical classes by rows labeled a-e, with ambient gases such as CO 261 262 and  $CO_2$  in group a, alkanes and alkenes in group b, alcohols, aldehydes and carboxylic acids in group c, aromatic species in group d, and N-bearing compounds in group e. The benefits of the in 263 situ laboratory static measurements were controlled gas sample collection with FTIR analysis and 264 265 longer scan times for increased SNRs at higher spectral resolution. Valves were shut before the 266 flame front arrived allowing for minimal mixing of air and flame gases near the extractive probe. In this manner the targeted pyrolysis phase was likely to be sampled with a greater mole fraction 267 rather than that of the combustion phase. The gases listed in Table 2 have previously been 268 269 observed in smoke in either field or laboratory settings, and some have been linked to pyrolysis 270 (Scharko, 2019a,b; Burling et al., 2010, 2011; Christian et al., 2003, 2004; Gilman et al., 2015; 271 Goode et al., 1999, 2000; Hatch et al., 2017; Selimovic et al., 2018; Stockwell et al., 2014; 272 Yokelson et al., 1996, 1997; Akagi et al., 2013, 2014; Alves et al., 2010; Hurst et al., 1994a, b;





- Karl et al., 2007; Paton-Walsh et al., 2010). Compounds associated with the pyrolysis phase and observed in several of the static measurements include acetic acid, ethene ( $C_2H_4$ ), allene, 1,3butadiene, acetaldehyde, formic acid, formaldehyde, acrolein, benzene, furan, furaldehyde,
- 276 naphthalene and phenol.
- 277 As seen in Table 2, ammonia gas (NH<sub>3</sub>) was also detected at fairly low mixing ratios in the 278 laboratory scale experiments, which had previously not been detected in the Ft. Jackson field 279 study: The lack of NH<sub>3</sub> detection in those studies was ascribed to the known adsorptivity of the compound as it may have adhered to either the transfer canister walls, the extractive probe, or the 280 281 White Cell, all at ambient temperatures as used in those studies (Scharko et al., 2019; Roscioli et al., 2015; Stockwell et al, 2014; Yokelson et al., 2003; Neuman et al., 1999). Adhesion losses 282 were minimized in the present experiments by a) measuring the gas parcel directly without storage 283 284 and b) heating transfer lines and gas cell to ~55 °C.

Table 2. Mixing ratio of chemicals from spectra collected using the static acquisition method. Burns are labeled
 by number and plant species. Compound mixing ratios are reported in ppm (with the exception of H<sub>2</sub>O and CO<sub>2</sub>
 reported as percents) and categorized by (a) background ambient compounds, (b) simple hydrocarbons, (c)

288	oxygenated organic compounds,	(d) aromatics, a	and (e) N-bearing species.
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		Burn 76	Burn 78	Burn 80	Burn 82	Burn 84	Burn 88	Burn 89	Burn 90	Burn 94	Burn 95
		inkberry	sparkleberry	sparkleberry	sparkleberry	fetterbush	fetterbush	inkberry	sparkleberry	sparkleberry	sparkleberry
	% H <sub>2</sub> O	1.24	1.05	3.23	2.03	3.54	3.08	3.46	1.82	6.21	4.10
_	% CO <sub>2</sub>	0.06	0.09	2.06	0.48	1.51	2.06	1.36	0.34	4.60	2.08
a	со	1.45	3.90	808	192	1089	1057	391	160	7506	2651
	N <sub>2</sub> O	0.35	0.34	1.21	0.50	1.28	1.79	0.44	0.41	3.22	1.78
	CH <sub>4</sub>	2.27	2.21	45.3	10.7	54.5	50.3	15.4	11.3	682	198
	$C_2H_2$	0.01	0.06	23.8	4.52	23.4	23.2	8.82	5.62	351	96.5
	$C_2H_4$	0.07	0.05	29.3	7.05	39.9	39.3	9.66	6.52	452	133
	C <sub>2</sub> H <sub>6</sub>			0.83			2.76		4.00E-04	24.2	6.29
b	C <sub>3</sub> H <sub>6</sub>			4.02	0.99	5.55	5.48	0.75	0.77	61.3	18.1
	allene	0.17		0.64	0.29	1.12	1.21	0.25	0.12	8.69	2.30
	1,3-butadiene			1.63	0.37	1.98	2.07	0.26	0.43	28.1	7.57
	isobutene			0.75		0.74	0.52			3.16	1.07
	isoprene			1.78	0.39	1.72	1.43	0.31	0.32	11.7	4.22
	CH <sub>3</sub> OH	0.89	0.24	6.81	1.53	6.92	9.44	1.66	0.93	42.3	18.0
	C <sub>2</sub> H <sub>5</sub> OH	1.37									
	acetic acid	0.07		5.93	3.55	13.4	13.8	11.0	2.49	13.4	9.62
	formic acid			15.9	5.14	32.35	35.3	9.20	3.64	130	73.6
C	acetaldehyde			5.87	1.69	7.62	8.65	1.51	0.94	73.6	22.6
	acrolein			2.59	1.29	3.99	4.35	0.98	0.00	26.0	9.53
	crotonalehyde			1.51	0.54			0.73	0.17	9.97	5.64
	formaldehyde		0.08	13.6	4.31	21.3	22.5	5.41	3.33	114	52.8
	benzene			4.08	2.23	5.19	4.24	1.93	1.48	61.3	18.6
_	furan			0.75		0.39	0.54			3.07	1.16
d	furfural			0.65	0.06				0.13	3.34	1.24
	naphthalene			4.48	1.06	3.60	4.80	3.40	0.82	14.6	1.42
	phenol			0.90	0.30	1.36	1.63	1.75	0.37	2.19	1.63
	NH <sub>3</sub>	0.10	0.29	0.19	1.29	1.79	0.88	1.08	0.41	0.58	0.57
ρ	HCN			5.84	2.19	8.25	6.94	3.36	1.69	64.2	21.0
C	HNCO			1.89	0.67	2.61	2.94	1.27	0.70	5.37	1.98
	HONO		0.11	9.40	2.53	9.72	12.7	8.92	1.75	26.9	11.3

289





290

291 When comparing the RFL laboratory scale experiments to the 2018 Ft. Jackson field scale 292 experiments, it is evident that field scale values via the static mode are greater than those of the 293 laboratory, even though the laboratory experiment attempted to replicate Ft. Jackson fuel beds and 294 scenarios. In most cases, a comparison of compounds found in the RFL laboratory burns and the 295 Ft. Jackson 2018 field burns finds Ft. Jackson mixing ratios approximately 4 to 10x greater than those of the RFL 2018 tunnel data. Field scale measurements typically yield more emissions than 296 experiments conducted in the laboratory (Yokelson, 2013; Scharko, 2019b, Weise et al. 2015). 297 298 However, while the mixing ratios may differ or be larger/smaller, the information describing the 299 composition of the mixture is relative in nature and is contained in log-ratios of the various gases. But analysis of the data as compositional data (Aitchison 1986) is beyond the scope of the present 300 301 paper. Table 3 displays the minimum and maximum mixing ratio values in ppm for five compounds from the Ft. Jackson studies presented in Scharko et al. (2019a) versus the present 302 303 RFL laboratory results. Of the five species compared, acetaldehyde, acrolein, and allene all follow the trend of having Ft. Jackson results being significantly higher than the RFL studies by a factor 304 305 of ~4. Naphthalene, a polycyclic aromatic hydrocarbon (PAH) was the only exception to this trend, having comparable mixing ratio values in the two studies. This anomaly could be attributed to one 306 of naphthalene's pyrolysis formation route as suggested by Fairburn et al., where a single ringed 307 308 aromatic compound undergoes a Diels-Alder reaction of an alkene (Fairburn et al., 1990; Liu et al., 2017). Of the four compounds compared, naphthalene is the only one to be derived from a 309 310 secondary reaction, whereas acetaldehyde and acrolein are derived directly from the pyrolysis of cellulose (Stein et al., 1983), while allene is a compound known to be a precursor of aromatic 311 312 compounds and soot (Frenklach et al., 1983, 1988). As noted, most compounds detected in the 313 RFL laboratory studies yielded ~4 to 10x lower mixing ratios compared to the field scale studies 314 at Ft. Jackson. Along with naphthalene, however, acetic acid, formaldehyde, isoprene and isobutene were also found to have comparable mixing ratios to those reported in the Ft. Jackson 315 studies. This could be due to the four compounds being products of secondary reactions, or 316 317 fragmentation, of species such as lignin, xylan and glucomannan (Collard and Blin, 2014). It 318 should be noted that of the five novel compounds detected in Scharko et al. (2019b), only four 319 were detected in these laboratory scale experiments. Methyl nitrite was not observed (Table 3). This is attributed to the field experiment being on the Ft. Jackson base where there is known to be 320





- 321 unexploded ordinance (Scharko et al., 2019b) or possibly due to lower concentration levels that
- 322 are below the detection limits of the present laboratory-scale experiment.

323 Table 3. Calculated minimum and maximum mixing ratios (ppm) for the 10 canister measurements taken at the

**324** Ft. Jackson field measurements (Scharko et al., 2019) along with the minimum and maximum mixing ratios

(ppm) for the 10 static measurements during the RFL laboratory experiment of acetaldehyde, acrolein, allene,
 methyl nitrite and naphthalene.

Target Compound	Ft. Jackson calculated mixing ratio (ppm)		RFL calculat mixing ratio (ppm)		
	min	max		min	max
acetaldehyde	34.5	264.8		0.94	73.6
acrolein	14.7	125.7		0.98	26
allene	2.2	37.8		0.12	8.69
methyl nitrite	2.3	21		-	-
nonhthalana		10.0		0.00	110

327

328 It is clear that the static method as deployed was not perfect at either isolating strictly the pyrolysis 329 phase gases or capturing extremely high fractions of combustion gases. The method relied heavily on valves being closed prior to the flame front using visual cues as opposed to using other 330 techniques, e.g. thermal IR. While not readily visible to the human eye, radiant and convective 331 332 heating (as determined by Background-Oriented Schlieren measurements - Aminfar et al 2019) occurred well in advance of the flame front, suggesting this as a possible alternate visualization 333 of pyrolysis gas release (Aminfar 2019). In any case, there is a narrow temporal window for the 334 pre-combustion phase making the valve-close time extremely important. For example, spectra 335 from burns 76 and 78 show largely the detection of only ambient air compounds indicating the 336 337 valves were closed too early. Conversely, in other samples there is clearly some mixing of both upstream and downstream air before the gas enters the extractive probe. Despite the shortcomings 338 of the static method most of the attempts to obtain pre-combustion gases were successful as 339 340 evidenced in part by the chemical composition of the isolated gases.

341

#### 342 *3.2 Spectral-Thermal Correlation to Isolate Pyrolysis Phase*

Dynamic IR data and visual image acquisition proved advantageous to resolve the different phases
of the experiments (e.g. pyrolysis, flaming combustion, smoldering combustion). This was





345 important since MCE is a function of the gas composition and is not unique to phase (i.e., the same value of MCE results if the same relative amounts of CO and CO<sub>2</sub> are observed, whether in the 346 pyrolysis, flaming or smoldering combustion phases) are less appropriate due to the lack of arrival 347 of the flame front and onset of combustion. MCE, defined as  $\Delta CO_2/(\Delta CO + \Delta CO_2)$ , has many 348 times been used to distinguish phases of combustion, namely flaming vs. smoldering although 349 Ward and Radke (1993) recommended combustion efficiency as the preferred descriptor of the 350 combustion system. MCE has not been used to identify pyrolysis nor should it be for the non-351 uniqueness described previously. Recent studies have introduced more sophisticated techniques to 352 analyze smoke emissions data with compositional data methods (Weise et al. 2020). However, 353 since primary and secondary pyrolysis occurs both prior to and after the onset of combustion or 354 oxidation, methods such as the MCE are not appropriate. We were not able to use the metric 355 suggested by Sekimoto et al. (2018), namely high temperature vs. low temperature pyrolysis as 356 357 determined from the acetylene-to-furan ratio due to weak furan signals in the present study due to 358 shortened scan times. The analysis was further exacerbated because furan's strongest vibrational band, the  $v_{19}$  vibrational band near 745 cm<sup>-1</sup>, corresponding to the C-H out-of-plane bend 359 (Shimanouchi, 1972), was obscured by saturated carbon dioxide lines and thus MALT was not 360 361 able to generate a satisfactory fit for this microwindow.

The pre-flame arrival gases were identified by either of two methods: The first method involved a 362 simple time subtraction of 4.8 s from the recording of the infrared spectrum time stamp and 363 associating that time to the corresponding visual and FLIR thermal infrared video images (Fig. 3). 364 This provided a relatively accurate verification that the gases being investigated were emitted prior 365 to the onset of combustion as seen in Table 4. The second method used the FTIR spectra directly: 366 demarcations for the flame front were denoted by the maximal value obtained for both CO and 367  $CO_2$  concentrations, i.e. greatest fraction of gas from the combustion phase. From this value the 368 369 FTIR's scans were selected for pyrolysis corresponding to the ~10 seconds before arrival (~0.1 m 370 distance) of the flame front.



371





Figure 3. Burn 87 inkberry on longleaf pine needle fuel bed – FLIR thermal imaging for burn progression.
 (a) frame corresponding to FTIR scan 5 signaling the pre-combustion phase, (b) frame corresponding to
 FTIR scan 16, flame front nearing sample probe, (c) frame corresponding to FTIR scan 21, inkberry bush
 consumed by flame, (d) frame corresponding to FTIR scan 44, flame front has passed the probe. Dark
 circles are ceramic plant holders.

The FTIR time-resolved scans (including derived chemical mixing ratios) synchronized to the RFL 377 378 time-stamped thermal IR temperature images provide insight into the chemical composition of each burn. As an example, Table 4 pairs data from the two systems for Burn 87. FTIR scan number, 379 380 FTIR time stamp, RFL FLIR recorded temperature near the extractive probe, and a selection of chemical concentrations are shown. The table demonstrates that spectral data for FTIR scans 0-8 381 saw no significant detections above ambient levels as corroborated by the FLIR images displaying 382 temperatures range from 40-80 °C (see Table 4 and Figure 3a); the extractive probe is still in the 383 low temperature region. The gradual increase in mixing ratios for most compounds (excluding 384 ammonia, which is primarily a smoldering gas) begins after FTIR scan 9. The magenta and orange 385 colored domains seen in Figure 3b indicate the encroaching flame front and a rise in thermal 386 temperatures. The frames corresponding to FTIR scans 16-19 display IR temperatures between 387 388 175 and 220 °C. In this temperature range compounds associated with the pyrolysis phase such as acetaldehyde, acetic acid and allene (shown in Table 4) are not only manifest in the IR spectra, but 389 their mixing ratios rise rapidly. Shortly thereafter the greatest mixing ratios of CO<sub>2</sub> occur at scans 390





- 20 through 22, indicating the flaming stage; this is corroborated by thermal IR video of the inkberry plant beginning to be fully consumed in flames (Fig 3c). As the flame front progressed down the tunnel, temperatures near the plant holder began to drop with the onset of the smoldering phase as indicated by lower mixing ratios as well as the thermal IR visual seen in Fig. 3d. [We note in Figure 3 that the temperature directly near/above the holders is much cooler due to minimal duff cover and the plants being green.] The video stopped recording at scan 48, when the flame reached
- 397 the end of the fuel bed although the FTIR continued to collect interferograms to monitor
- 398 smoldering from the fire.

399

- 400 Table 4: Burn 87 inkberry amongst pine needle fuel bed FTIR scan summary synchronized to FLIR temperature data.
- 401 Scan number, FTIR time stamp, along with FLIR video emissions temperature at extractive probe accounting for time
- 402 delay and mixing ratios from carbon dioxide (CO<sub>2</sub>), carbon monoxide (CO), ethene (C<sub>2</sub>H<sub>4</sub>), acetic acid (CH<sub>3</sub>COOH), 403 formaldehyde (HCHO), acetaldehyde (CH<sub>3</sub>CHO), and phenol (C<sub>6</sub>H<sub>6</sub>O).





			4.8 s Delaved	FLIR							
	FTIR Scan	FTIR time	FLIR Video	temperature	CO,	со	C₂H₄	сн₃соон	нсно	CH3CHO	C <sub>6</sub> H <sub>6</sub> O
	Number	stamp	Time Stamp	at inlet (°C)	(ppm)	(ppm)	(ppm)	(ppm)	(ppm)	(ppm)	(ppm)
	scan 0	10:41:23.69	10:41:18.89	41.4	1548	49.5	2.4	0.0	0.6	-	-
	scan 1	10:41:25.20	10:41:20.40	44.2	1912	74.5	1.9	1.7	0.9	-	-
	scan 2	10:41:26 70	10:41:21 90	51.0	1562	63.7	2.3	1 1	0.3	-	-
Ħ	scan 3	10.41.28 21	10.41.23 41	54.9	1290	58.4	11	0.5	0.5	-	-
bie	scan 4	10:41:29.71	10:41:24.91	53.4	1882	63.1	1.2	1.2	0.8	-	-
, in the second	scan 5	10:41:31.21	10:41:26.41	63.2	1946	57.0	1.5	1.1	0.4	-	-
4	scan 6	10:41:32.72	10:41:27.92	72.8	3811	102	4.7	1.8	1.2	-	-
	scan 7	10.41.34 22	10.41.29 42	77.6	4722	138	2.7	19	1 5	-	-
	scan 8	10:41:35.73	10:41:30.93	86.2	3553	109	0.9	2.2	1.4	-	-
	scan 9	10:41:37 23	10:41:32 43	116.9	2957	97	1.0	2.2	0.7	_	
50	scan 10	10:41:37:23	10:41:32.43	132.1	3360	138	1.0	2.4	1.7	14	_
ysi	scan 11	10:41:40 24	10:41:35.33	115 7	7476	246	1.0	2.0	3.6	0.0	_
2	scan 12	10:41:41 74	10:41:35.44	121.2	10274	240	 	2.0	1.0	4.2	0.7
<u>ъ</u>	scan 12	10:41:41.74	10:41:30.34	121.3	10274	201	11 /	11 5	4.0	4.2	1.6
	scan 1/	10:41:44 75	10:41:30.45	159.0	11833	635	23.5	14.5	13.5	3.3	1.0
tio	scan 15	10:41:44.75	10:41:33:33	162.9	16090	1214	23.5	14.5	29.7	3.3	1.5
liza	scan 16	10:41:40.25	10:41:41.45	105.8	25757	2214	40.5	16.2	54.4	17.6	1.5
lati	scan 17	10:41:47.70	10:41:42.30	191.2	21956	2217	120	14.4	70.5	24.0	1.0
Vo]	scan 19	10:41:49.20	10:41:44.40	220.1	41201	2913	260	14.4	0.5	24.0	1.4
-	scan 10	10.41.50.77	10:41:45.97	220.1	61166	2070	200	13.1	92.0	34.5	1.4
	Scall 19	10.41.52.27	10:41:47.47	219.0	70222	0220	433	12.5	121	44.5	2.1
	scan 20	10:41:53.77	10:41:48.97	296.1	79332	11354	/4/	13.1	1/8	80.7	2.9
	scan 22	10:41:55.28	10:41:50.48	201.7	54381	12054	1025	17.0	200	140	4.7
	scan 22	10:41:56.78	10:41:51.98	450.1	64077	12954	1025	15.3	100	103	6.0
- U	scan 23	10:41:58.29	10:41:53.49	429.9	41495	8620	530	15.4	123	63.3	6.3
usti	scan 24	10:41:59.79	10:41:54.99	516.5	25879	3453	257	15.5	/4.5	33.9	6.8
- Â	scan 25	10:42:01.29	10:41:56.49	514.5	15965	3110	52.2	15.1	45.7	19.6	5.8
-5-	scan 26	10:42:02.80	10:41:58.00	460.1	11819	2416	53.2	15.6	35.2	12.0	5.0
- <u>2</u> 0	scan 27	10:42:04.30	10:41:59.50	453.7	8566	18/5	30.0	14.7	27.4	6.9	5.2
Ē	scan 28	10:42:05.81	10:42:01.01	448.0	5795	1320	14.5	13.2	21.8	3.0	5.6
E	scan 29	10:42:07.31	10:42:02.51	440.2	5235	1302	11.0	14.6	20.0	5.1	4.5
	scan 30	10:42:08.81	10:42:04.01	484.7	3626	916	5.8	15.1	15.0	5.2	4.2
	scan 31	10:42:10.32	10:42:05.52	470.4	2368	570	3.3	11.5	11.1	2.7	3.8
	scan 32	10:42:11.82	10:42:07.02	497.5	1636	3//	1.6	10.8	9.4	0.3	3./
	scan 33	10:42:13.33	10:42:08.53	477.4	1684	399	1.0	9.2	8.8	2.1	3.4
	scan 34	10:42:14.83	10:42:10.03	450.1	1986	519	0.9	10.4	9.5	-1.5	3.0
	scan 35	10:42:16.33	10:42:11.53	397.9	1968	518	0.7	9.9	9.3	2.8	3.2
	scan 36	10:42:17.84	10:42:13.04	410.2	1901	495	1.4	9.6	8.8	0.9	2.8
-	scan 37	10:42:19.34	10:42:14.54	401.6	1936	516	1.5	9.2	9.4	0.0	2.7
tio	scan 38	10:42:20.85	10:42:16.05	358.3	1935	513	1.6	9.0	9.5	1.1	2.8
sng	scan 39	10:42:22.35	10:42:17.55	341.8	1753	439	1.8	9.7	8.9	1.1	2.3
Ē	scan 40	10:42:23.85	10:42:19.05	320.6	1438	345	1.4	10.4	8.3	-0.5	2.9
ů	scan 41	10:42:25.36	10:42:20.56	305.9	1224	277	-0.1	10.9	7.4	1.4	2.8
iig	scan 42	10:42:26.86	10:42:22.06	295.0	1377	324	1.4	11.6	8.2	-2.0	2.4
der	scan 43	10:42:28.37	10:42:23.57	272.5	1629	411	1.1	11.8	8.4	0.3	2.4
nol	scan 44	10:42:29.87	10:42:25.07	258.3	1366	325	0.8	12.8	7.8	2.5	2.7
S	scan 45	10:42:31.37	10:42:26.57	260.1	1059	212	0.4	11.9	6.7	3.4	2.7
	scan 46	10:42:32.88	10:42:28.08	238.5	1037	212	0.8	13.1	7.1	0.5	2.3
	scan 47	10:42:34.38	10:42:29.58	223.9	1094	236	0.9	14.4	6.7	1.1	1.2
	scan 48	10:42:35.89	10:42:31.09	226.7	1117	248	1.2	13.1	7.0	-0.4	2.7

404 405

406 As stated, a second method was also used to analyze/corroborate the different stages of the burn, 407 whereby mixing ratios of  $CO_2$ , CO and  $C_2H_4$  were analyzed to find their burn maxima (Viatte et

408 al., 2015). The CO<sub>2</sub> elevated mixing ratios (esp. relative to CO) are associated with the hottest,





flaming stage of biomass burns (Yokelson et al., 1996). To temporally isolate the flaming stage, the MCE criteria was employed and values of 88-95% indicative of smoldering were found for the region. Having identified the flaming stage, the pyrolysis stage was estimated by subtracting 6-8 seconds from that spectrum with maximal CO/CO<sub>2</sub> emissions, corresponding to ~4 FTIR scans (at 1 cm<sup>-1</sup> resolution). The agreement between the two methods was quite good and helped to demarcate the stages as seen in Table 4.

415

416 Figure 4 displays the infrared spectral progression of Burn 87, longleaf pine needles with inkberry, bed at 1.0 cm<sup>-1</sup> resolution looking at two different spectral regions. The CO (and  $CO_2$ ) profiles are 417 seen in Fig. 4a. Noted on the z-axis is scan 22; scans 20-22 are the time frames where maximal 418  $CO_2$  and CO emissions were observed; the region is also denoted by red spectral traces. Once the 419 flaming stage had been identified, the pyrolysis phase was then demarcated; in the pyrolysis phase 420 CO was evident (partially from upwind mixing) and was beginning to significantly increase; the 421 422 stage is indicated by orange traces (scans 16-19) in Figure 4. Other stages assigned were noted as the pre-flame stage where  $\triangle CO$  and  $\triangle CO_2$  were near zero in the FTIR data and are seen as scans 423 0-8 with purple traces. Blue traces correspond to the smoldering phase of combustion, where  $CO_2$ 424 425 mixing ratios decreased, the flame front had passed the extractive probe and MCE values were on the order of 85-75%. The spectral profile and mixing ratios of ethene ( $C_2H_4$ ) were also used to 426 evaluate the time-resolved FTIR data. (Johnson et al. 1993) This lightweight hydrocarbon is a 427 product of primary pyrolysis and if detected can be used to determine certain stages of the burn 428 (e.g. Yang et al., 2007). Figure 4b displays primarily the  $v_7$  band of ethene at 949.4 cm<sup>-1</sup> 429 (Shimanouchi, 1972). Ethene reached its maxima mixing ratio at scan 22 (red traces) before it 430 quickly disappeared, being a pyrolysis gas that was oxidized by the flame. It was first seen to 431 appear as early as scan 13 (green traces) but became clearly evident in scan 16 (orange traces, 432 433 pyrolysis phase) and continued to grow. The rapid disappearance of  $C_2H_4$  upon combustion is similar to that of formaldehyde and acetaldehyde (Table 4) whose concentrations also dropped 434 435 after scan 23, but the disappearance is juxtaposed with acetic acid whose values remained ~constant throughout the flaming and smoldering phases. As seen in the IR data, the  $C_2H_4$  gas 436 437 signal corroborated that ethene is a key product of the primary pyrolysis phase. Other compounds 438 showing significant signals in this time domain and described as pyrolysis gases include acrolein





- 439 and allene. (Scharko 2019a; Akagi et al., 2013; Frenklach et al. 1983, 1988; Stein et al., 1983;
- 440 Koss et al., 2018; Brilli et al., 2014).



441

442 Figure 4. Burn 87, inkberry with longleaf pine needles: a) CO and CO<sub>2</sub> spectral profile from 2250-2000 cm<sup>-1</sup>. 443 Purple traces indicate the ambient stage, green and orange traces indicate the pre-combustion/pyrolysis stage, 444 red spectral traces indicate the flaming stage and blue traces indicate smoldering. b) Largely C<sub>2</sub>H<sub>4</sub> spectral 445 waterfall plot from 1000-900 cm<sup>-1</sup> with accompanying C<sub>2</sub>H<sub>4</sub> reference spectrum as black trace.

446

447 The two methods to determine the pyrolysis, flaming, combustion and smoldering phases yielded congruous results: The isolated burn stages determined from method one, in which FTIR gas-448 phase data were synchronized to the FLIR thermal imaging, and from method two, using the FTIR 449 time-resolved data only, were found to be virtually identical. This is evidenced by linking the scans 450





determined to be in the pyrolysis phase (scans 16-19) using method two as seen in Fig. 4, with the
temperature data recorded by the FLIR using method one and seen in Table 4. For these scans,
the temperature profile ranges from 175-220 °C, corresponding to temperatures associated with
first stages of pyrolysis.

455 For most analytes biomass burning gas mixing ratios, the concentration values observed at the peak of the dynamic measurements values were significantly greater than for the concentrations 456 457 recorded in the static measurements. The dynamic experiments were of course carried out for the duration of the burn, whereas the static burns (in an effort to characterize pre-combustion phases) 458 459 attempted to isolate a specific time when the pyrolyzate concentrations were maximized. Analyzing the data using the dynamic technique allowed for confirmation of certain compounds 460 such as naphthalene, allene, acetaldehyde, and acrolein as compounds that appeared during the 461 pyrolysis phase. These compounds, which have been previously detected as pyrolysis gases using 462 463 FTIR for field plot burns, (Scharko et al. 2019) were again observed during these laboratory scale 464 tests and in almost all cases appeared before the flame front encroached on the sampling probe.

## 465 *3.3 Dynamic Detection of Phenol in the Pre-combustion Through Smoldering Stages*

In the present study phenol ( $C_6H_6O$ ) was detected during several burns; its origin ascribed to the 466 pyrolysis of lignin(s) (Kibet et al. 2012, Hawthorne et al. 1989) and has been mostly observed 467 using other techniques such as gas chromatography mass spectrometry (GC-MS). (Saiz-Jimenez 468 et al. 1986) Phenol and phenolic compounds are also known to contribute to the formation of 469 secondary organic aerosols. (Yee et al., 2013) It has been observed in simple pyrolysis 470 471 experiments, emanating from both pine and spruce species (e.g. Saiz-Jimenez and De Leeuw, 472 1986, Ingemarsson et al., 1998). In addition to simply pyrolytic emissions, phenol has also been 473 identified as a common component of tar as a pyrolysis product. In biomass burning, phenol has 474 been observed using both FTIR and other methods, (Gilman et al. 2015, Yokelson et al. 2013), e.g. proton-transfer mass spectrometry (PTR-MS) and GC-MS. In 2013 phenol was detected in a 475 476 closed cell, airborne FTIR field experiment but not in an open-path FTIR lab experiment 477 (Yokelson et al. 2013). The absence of  $C_6H_6O$  in the lab experiment was attributed to the lack of consumption of rotten wood as fuel. In those studies, airborne phenol emissions measured in the 478 field with closed-cell FTIR were also noted as being 2 to 4x greater than the phenol emissions 479 480 captured by PTR-MS in the laboratory.





481 In the present experiments, phenol was detected in 8 of the 10 static measurements (recall that two 482 of the static measurements only showed ambient gases due to early closure of the valves). Figure 5 demonstrates the static spectrum from Burn 89, corresponding to the burning of longleaf pine 483 with inkberry. Seen in Fig. 5 are the experimental spectrum (blue trace) and also the reference 484 spectrum of acetic acid (green trace). After subtraction of the CH<sub>3</sub>COOH vapor spectrum, the 485 residual contained two small peaks which were readily identified as phenol vapor via the  $v_{15}$ 486 vibrational band near 1176.2 cm<sup>-1</sup>, as well as the  $v_{16}$  band at 1150.2 cm<sup>-1</sup> (Keresztury et al., 1998). 487 The phenol reference spectrum from the PNNL spectral library (red trace) was then subtracted 488 from that residual (purple trace) with an overall residual that is mostly noise (black trace). [For the 489 dynamic spectra the process is repeated for each of the individual spectral time slices, represented 490 by scan number using the concentration of phenol determined by the MALT program.] To confirm 491 the spectral analysis, in each case the mixing ratio calculated by MALT was converted to a 492 493 spectrum by multiplying by the appropriate concentration path length factor; the predicted 494 spectrum was visually compared to the actual data.



495

Figure 5. Static spectrum obtained from Burn 89 (1 kg longleaf pine needles with inkberry). The blue trace
 is the FTIR experimental spectrum, the green trace the reference spectrum of acetic acid, the purple trace
 the residual after acetic acid subtraction, the red trace the reference spectrum of phenol and the black trace
 residual after phenol optimization/subtraction.





500 Phenol was also detected using the dynamic method and Figure 6 displays a series of dynamic 501 spectra recorded for Burn 87. The spectra in the left frame (a) are individual spectra after the acetic acid (CH<sub>3</sub>COOH) spectral component has been subtracted from the spectrum for each time slice, 502 all recorded at 1.0 cm<sup>-1</sup> resolution with 54 total measurements recorded at  $\Delta t=1.5$  seconds. While 503 the spectral noise is still significant, the presence of phenol peaks, particularly the  $v_{16}$  Q-branch at 504 1176.5 cm<sup>-1</sup> and the  $v_{15}$  peak at 1150.2 cm<sup>-1</sup>, are evident. Optimization for the phenol mixing ratio 505 in each spectrum allowed for its calculation in individual time slices and the derived phenol-only 506 spectra are presented as a waterfall plot in the right frame (b). The first clear evidence of phenol is 507 seen in scans 14 to 18, before reaching a maximum concentration of 6.9 ppm in scan 24; this is 508 observed in the right frame of Figure 6, coinciding approximately with maximal CO<sub>2</sub> concentration 509 (scan 22), indicating the greatest ratio of smoke/ambient air in the gas cell. 510



511



**Figure 6.** Burn 87 –longleaf pine needles with inkberry fuel bed during dynamic mode. Measured and scaled burn spectra showing the progression of phenol during the time resolved study. Acetic acid and water spectral features have been removed in Frame a) with the phenol-only derived mixing ratio spectra in Frame b.

Figure 6 displays the rapid increase of phenol vapor due to the approaching flame front from scan 516 14 (t = 22.5 s) to its maximal mixing ratio in scan 24 (t = 36 s) followed by a longer gradual phenol 517 decay with time. This can be juxtaposed with the ethene mixing ratios (seen in Figure 4) that fall 518 519 to nearly zero with the onset of combustion; the ethene is consumed by the flame propagation. 520 Prior to scan 14 in Figure 6b, minimal phenol is observed relative to the noise level and are thus fit as zero concentration. Phenol contributions for scans 16-19 can be associated with the pyrolysis 521 phase of the burn and not combustion. Phenol is one of the major products of 1,2-benzenediol 522 pyrolysis with maximum yield reported at 800 °C (Ledesma et al. 2002; Thomas et al. 2007). It is 523 important to note that both temperature and rate of heating influence the composition and yield of 524 525 pyrolysis products. Evidenced in Figure 6, the thermal imaging associated with FTIR scan 17





shows a temperature of  $\sim 200$  °C which is indicated in Kibet et al. (2012) to be within the 526 temperature range of pyrolysis of lignin: 200 to 400 °C. Shortly thereafter, phenol mixing ratios 527 rapidly increase and reach a maximal mixing ratio of 6.9 ppm at scan 24. At scan 24 the flame 528 529 front has already reached the extractive probe and thus the maximum intake of smoke and ambient air is achieved; temperature of the fuel bed is ca. 600 °C, consistent with the flaming phase. The 530 gradual decay in phenol production as the flame front passes could be due to several factors: (i) an 531 increased temperature required for complete combustion of the C<sub>6</sub>H<sub>6</sub>O (ii) residence time of 532 phenol, (iii) phenol production in the smoldering phase as a tar/char, (iv) adsorption to walls of the 533 stainless steel tubing and cell. A cross-section of Figure 6 shows the rapid onset of phenol 534 production at the temperatures followed by a gradual decay in concentration: this is indicative of 535 phenol production throughout the burning of inkberry as a species. 536



537

Figure 7. Temporal mixing ratios of phenol for different shrub species. Phenol mixing ratios plotted over time indicated by scan number for Burn 97 sparkleberry (red), Burn 87 inkberry (green), and Burn 20 fetterbush (blue) all on longleaf pine straw bed.

The shape of the temporal profile yields information as to the production of phenol throughout the evolution of burning. Figure 7 shows the progression of phenol concentration following its first observed presence in the burn. These graphs are effectively a cross-section of Figure 6, showing the progression of the height of the phenol peak (directly correlated to phenol concentration) throughout the burn (with time being represented by scan number in both cases). The level of





546 phenol generation was observed to vary between plant species. Temporal profiles of phenol 547 concentration were constructed for burns with three different species: sparkleberry, inkberry, and fetterbush. These plots illustrate a range of behavior with inkberry and sparkleberry having similar 548 temporal profiles and similar maxima of ca. 6.5 ppm and fetterbush having a different temporal 549 profile. It is important to note that Burn 97 was measured at 0.6 cm<sup>-1</sup>, while burns 87 and 93 were 550 measured at 1.0 cm<sup>-1</sup>, although the profile of burn 97 is consistent with that of burn 87. [We do not 551 believe the small change in resolution affects the recovered mixing ratios.] Demonstrated in Figure 552 7, trace amounts of phenol appear at the onset of combustion and throughout the pyrolysis phase. 553 Phenol reaches its highest concentrations, however, during the flaming stage as all three temporal 554 profiles reach maximum during the latter stages of the burn. Moreover, phenol remains throughout 555 the duration of the burn and is not consumed by secondary reactions, as is e.g. ethene. For these 556 three burns, fetterbush was observed to have the lowest maximum concentration of phenol, 3.1 557 558 ppm, of the three species, while sparkleberry and inkberry had similar maxima (as well as similar 559 temporal profiles).

560 The observed differences of phenol in both temporal profile and overall peak concentrations could arise due to differences in leaf structure and shape, or possibly due to differences in leaf/plant 561 composition. Pyrolytic production of phenol has been previously attributed to multiple 562 components of plant composition, including phenol content, lignin content and the amount of 563 cellulose in each plant species. Therefore, varying phenols, lignin, and cellulose in these plant 564 565 species could be the source of phenol concentration variability for each burn. The physical composition of multiple plant species, including inkberry and fetterbush was analyzed by Matt and 566 Dietenberger (2020); it was shown that inkberry has 2.6 times the percentage of phenol by 567 composition (9.0%) than fetterbush (3.4%) Although sparkleberry was not included in that study, 568 it can be suggested that the compositions of inkberry and sparkleberry are similar due to observed 569 phenol in this experiment as well as plant characteristics. Sparkleberry, a member of the Vaccinium 570 genus, contains many species collectively known as blueberries which are known to contain high 571 levels of phenolic compounds in the fruits (e.g. Prior et al 1998). This study and the results of Matt 572 and Dietenberger support the present hypothesis that peak concentrations of phenol are highest for 573 sparkleberry and inkberry due to higher phenolic content in the plants. 574

575 4. Summary





576 The analytical methods used in this study attempt to provide a detailed view of prescribed burning by enlisting two different FTIR acquisition modes, static and dynamic. By capturing a "snapshot" 577 of a single burn experiment, used in the static method, one can discern the gases with higher 578 579 specificity and in turn decipher complex spectra by use of chemometrics to extract compounds with high concentrations leaving behind a residual to be analyzed. Lower resolution may hinder 580 these efforts and allow compounds that are present at lower mixing ratios to be obscured by higher 581 absorbing compounds, e.g., carbon dioxide, water, and ethene. In this study we were able to detect 582 additional compounds e.g. phenol, benzene, and allene with greater confidence. However, in 583 gaining specificity there is a loss of time resolution and this is where the dynamic method becomes 584 advantageous. The FTIR dynamic acquisition method when synchronized to thermal imaging, 585 while lower in sensitivity, allows for an overall profile of the burn and can help assign phases to 586 the dynamic stages of the flame. That is, the dynamic method in conjunction with thermal IR 587 588 imaging provides a more detailed description as temperature and chemical composition profiles 589 can be correlated and assigned to certain phases of the burns. In this study pyrolysis, flaming and smoldering combustion were identified using these new techniques which can aide in the 590 improvement of fire behavior models used by land managers to conduct prescribed fires. 591

592 *Data availability*. Data are not publicly available as data release has not been authorized by sponsor593 of this research.

594 *Competing interests.* No competing interests.

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### 600 **REFERENCES**

- 601 Agee, J. K.: Fire and pine ecosystems, in Ecology and Biogeography of Pinus, edited by D. M.
- 602 Richardson, pp. 193–218, Cambridge University Press, Cambridge, U.K., 2000.
- Aitchison, J.: The statistical analysis of compositional data, Chapman and Hall, London; NewYork., 1986.
- Akagi, S. K., Yokelson, R. J., Burling, I. R., Meinardi, S., Simpson, I., Blake, D. R.,
- 606 McMeeking, G. R., Sullivan, A., Lee, T., Kreidenweis, S., Urbanski, S., Reardon, J., Griffith, D.
- W. T., Johnson, T. J., and Weise, D. R.: Measurements of reactive trace gases and variable O3
- formation rates in some South Carolina biomass burning plumes, Atmos. Chem. Phys., 13, 1141-
- 609 1165, 2013.
- 610 Akagi, S. K., Burling, I. R., Mendoza, A., Johnson, T. J., Cameron, M., Griffith, D. W. T., Paton-
- 611 Walsh, C., Weise, D. R., Reardon, J., and Yokelson, R. J.: Field measurements of trace gases
- emitted by prescribed fires in southeastern US pine forests using an open-path FTIR system,
- 613 Atmos. Chem. Phys., 14, 199-215, 2014.
- Alves, C. A., Gonçalves, C., Pio, C. A., Mirante, F., Caseiro, A., Tarelho, L., Freitas, M. C., and
- Viegas, D. X.: Smoke emissions from biomass burning in a Mediterranean shrubland, Atmos.
  Environ., 44, 3024-3033, 2010.
- 617 Aminfar, A., Cobian-Iñiguez, J., Ghasemian, M., Espitia, N. R., Weise, D. R. and Princevac, M.:
- Using Background-Oriented Schlieren to Visualize Convection in a Propagating Wildland Fire,
- 619 Combustion Science and Technology, 1–21, doi:<u>10.1080/00102202.2019.1635122</u>, 2019.
- Aminfar, A.: Application of Computer Vision to Transport Phenomena, Ph.D., University ofCalifornia, Riverside., 2019.
- Amini, E., Safdari, M.-S., DeYoung, J. T., Weise, D. R. and Fletcher, T. H.: Characterization of
- 623 pyrolysis products from slow pyrolysis of live and dead vegetation native to the southern United
- 624 States, Fuel, 235, 1475–1491, doi:10.1016/j.fuel.2018.08.112, 2019.
- Barbour, M. G. and Billings, W. D., Eds.: North American terrestrial vegetation, 2. ed.,
  Cambridge Univ. Press, Cambridge., 2000.
- 627 Behm, A., Duryea, M. L., Long, A. J. and Zipperer, W. C.: Flammability of native understory
- 628 species in pine flatwood and hardwood hammock ecosystems and implications for the wildland-
- 629 urban interface, International Journal of Wildland Fire, 13(3), 355–365, doi:<u>10.1071/WF03075</u>,
  630 2004.
- 631 Bist, H. D., Brand, J. C. D. and Williams, D. R.: The Vibrational Spectrum and Torsion of
- 632 Phenol, J. Mole. Spec, 24, 402-412, 1967.
- Biswell, H. H.: Prescribed burning in California wildlands vegetation management, Berkeley,
- 634 CA: University of California Press; p. 255, 1989.





- Brilli, F., Gioli, B., Ciccioli, P., Zona, D., Loreto, F., Janssens, I. A., and Ceulemans, R.: Proton
- 636 Transfer Reaction Time-of-Flight Mass Spectrometric (PTR-TOF-MS) determination of volatile
- organic compounds (VOCs) emitted from a biomass fire developed under stable nocturnal
- 638 conditions, Atmos. Environ., 97, 54-67, 2014.
- Burgan, R. E. and Susott, R. A.: Influence of sample processing techniques and seasonal
- 640 variation on quantities of volatile compounds of gallberry, saw-palmetto and wax myrtle,
- 641 International Journal of Wildland Fire, 1(1), 57–62, doi:<u>10.1071/WF9910057</u>, 1991.
- 642 Burling, I. R., Yokelson, R. J., Griffith, D. W. T., Johnson, T. J., Veres, P., Roberts, J. M.,
- 643 Warneke, C., Urbanski, S. P., Reardon, J., Weise, D. R., Hao, W. M. and de Gouw, J.:
- 644 Laboratory measurements of trace gas emissions from biomass burning of fuel types from the
- southeastern and southwestern United States, Atmos. Chem. Phys., 10(22), 11115–11130,
- 646 doi:<u>10.5194/acp-10-11115-2010</u>, 2010.
- 647 Burling, I. R., Yokelson, R. J., Akagi, S. K., Urbanski, S. P., Wold, C. E., Griffith, D. W. T.,
- Johnson, T. J., Reardon, J., and Weise, D. R.: Airborne and ground-based measurements of the
- trace gases and particles emitted by prescribed fires in the United States, Atmos. Chem. Phys.,
  11, 12197-12216, 2011.
- Bytnerowicz, A., Arbaugh, M. A., Andersen, C. K. and Riebau, A. R., Eds.: Wildland fires and
  air pollution, Elsevier, Amsterdam; Boston., 2009.
- Carter, M. C. and Foster, C. D.: Prescribed burning and productivity in southern pine forests: a
  review, Forest Ecol. Manage., 191, 93–109, 2004.
- 655 Christensen, N. L.: Vegetation of the Southeastern Coastal Plain, in North American Terrestrial
- Vegetation, edited by M. G. Barbour and W. D. Billings, pp. 397–448, Cambridge University
  Press, New York, NY., 2000.
- 658 Christian, T. J., Kleiss, B., Yokelson, R. J., Holzinger, R., Crutzen, P. J., Hao, W. M., Shirai, T.,
- and Blake, D. R.: Comprehensive laboratory measurements of biomass-burning emissions: 2.
- 660 First intercomparison of open-path FTIR, PTR-MS, and GC-MS/FID/ECD, J. Geophys. Res.
- 661 Atmos., 109, 2004.
- 662 Chi, C. T., Horn, D. A., Zanders, D. L., Opferkuch, R. E., Nyers, J. M., Pierovich, J. M., Lavdas,
- L. G., McMahon, C. K., Nelson, R. M., Jr., Johansen, R. W. and Ryan, P. W.: Source
- Assessment: Prescribed Burning, State of the Art, Environmental Protection Technology, United
   States Environmental Protection Agency, Research Triangle Park, NC. [online] Available from:
   nepis.epa.gov, 1979.
- 667 Cohen, S., Hall, J. and Hiers, J. K.: Fire Science Strategy, Strategic Environmental Research and
- 668 Development Program, Resource Conservation and Climate Change Program Area, Washington,
- 669 D.C. [online] Available from: <u>https://serdp-</u>
- 670 estcp.org/content/download/30210/291748/file/Fire%20Science%20Strategy.pdf, 2014.





- 671 Collard, F.X. and Blin, J.: A review on pyrolysis of biomass constituents: Mechanisms and
- 672 composition of the products obtained from the conversion of cellulose, hemicelluloses and
- 673 lignin. Renewable and Sustainable Energy Reviews, 38, pp.594-608, 2014.
- 674 Crutzen, P. J. and Goldammer, J. G., Eds.: Fire in the environment: the ecological, atmospheric,
- and climatic importance of vegetation fires: report of the Dahlem Workshop, held in Berlin, 15-
- 676 20 March 1992, Wiley, Chichester, England; New York., 1993.
- 677
- 678 Depew, C. A., Mann, M. J. and Corlett, R. C.: A Laboratory Simulation of Wood Pyrolysis
- 679 Under Field Conditions, Combustion Science and Technology, 6(4), 241–246,
- 680 doi:<u>10.1080/00102207208952326</u>, 1972.
- 681 Di Blasi, C.: Modeling chemical and physical processes of wood and biomass pyrolysis, Progress
- in Energy and Combustion Science, 34(1), 47–90, doi:<u>10.1016/j.pecs.2006.12.001</u>, 2008.
- 683
- Dimitrakopoulos, A. P.: Thermogravimetric analysis of Mediterranean plant species, Journal of
   Analytical and Applied Pyrolysis, 60(2), 123–130, doi:10.1016/S0165-2370(00)00164-9, 2001.
- 686
- Fairburn, J. A., Behie, L. A., and Svrcek, W. Y.: Ultrapyrolysis of n-hexadecane in a novel
  micro-reactor, FUEL, 69, 1537-1545, 1990.
- 689
- Frenklach, M., Taki, S., Durgaprasad, M. B., and Matula, R. A.: Soot formation in shock-tube
  pyrolysis of acetylene, allene, and 1, 3-butadiene, Combust. Flame, 54, 81–101, 1983.
- 692
- Frenklach, M., Yuan, T., and Ramachandra, M. K.: Soot formation in binary hydrocarbon
  mixtures, Energy Fuels, 2, 462–480, 1988.
- 695

Gilman, J. B., Lerner, B. M., Kuster, W. C., Goldan, P. D., Warneke, C., Veres, P. R., Roberts, J.
M., de Gouw, J. A., Burling, I. R., and Yokelson, R. J.: Biomass burning emissions and potential
air quality impacts of volatile organic compounds and other trace gases from fuels common in

- 699 the US, Atmos. Chem. Phys., 15, 13915-13938, 2015.
- 700
- 701 Griffith, D. W. T.: MALT5 User guide Version 5.5.9 2016.
- 702
- Goode, J. G., Yokelson, R. J., Susott, R. A., and Ward, D. E.: Trace gas emissions from
- laboratory biomass fires measured by open-path Fourier transform infrared spectroscopy: Fires
   in grass and surface fuels, J. Geophys. Res. Atmos., 104, 21237-21245, 1999.
- 706
- 707 Goode, J. G., Yokelson, R. J., Ward, D. E., Susott, R. A., Babbitt, R. E., Davies, M. A., and Hao,
- 708 W. M.: Measurements of excess O3, CO2, CO, CH4, C2H4, C2H2, HCN, NO, NH3, HCOOH,
- 709 CH3COOH, HCHO, and CH3OH in 1997 Alaskan biomass burning plumes by airborne Fourier
- transform infrared spectroscopy (AFTIR), J. Geophys. Res. Atmos., 105, 22147-22166, 2000.
- 711
- 712 Gordon, I. E., Rothman, L. S., Hill, C., Kochanov, R. V., Tan, Y., Bernath, P. F., Birk, M.,
- 713 Boudon, V., Campargue, A., Chance, K. V., Drouin, B. J., Flaud, J.-M., Gamache, R. R.,
- Hodges, J. T., Jacquemart, D., Perevalov, V. I., Perrin, A., Shine, K. P., Smith , M.-A. H.,
- 715 Tennyson, J., Toon, G. C., Tran, H., Tyuterev, V. G., Barbe, A., Császár, A. G., Devi, V. M.,



716



I., Kyuberis, A. A., Loos, J., Lyulin, O. M., Massie, S. T., Mikhailenko, S. N., Moazzen-Ahmadi, 717 N., Müller, H. S. P., Naumenko, O. V., Nikitin, A. V., Polvansky, O. L., Rev, M., Rotger, M., 718 Sharpe, S. W., Sung, K., Starikova, D., S.A. Tashkun, S. A., Van der Auwera, J., Wagner, G., 719 Wilzewski, J., Wcisło, P., Yu, S., and Zak, E. J.: The HITRAN2016 molecular spectroscopic 720 721 database, J. Quant. Spectrosc. Radiat. Transfer, 203, 3-69, 2017. 722

Furtenbacher, T., Harrison, J. J., Hartmann, J.-M., Jolly, A., Johnson, T. J., Karman, T., Kleiner,

- 723 Guérette, É.-A., Paton-Walsh, C., Desservettaz, M., Smith, T. E. L., Volkova, L., Weston, C. J.
- and Meyer, C. P.: Emissions of trace gases from Australian temperate forest fires: emission 724
- factors and dependence on modified combustion efficiency, Atmos. Chem. Phys., 18, 3717-3735, 725 doi: 10.5194/acp-18-3717-2018, 2018. 726
- 727
- Hardy, C. C., Ottmar, R. D., Peterson, J. L., Core, J. E. and Seamon, P.: Smoke management 728 729 guide for prescribed and wildland fire; 2001 ed., PMS 420-2 National Wildfire Coordinating
- group, Boise, ID. 226 pp., 2001. 730
- 731
- 732 Hatch, L. E., Yokelson, R. J., Stockwell, C. E., Veres, P. R., Simpson, I. J., Blake, D. R.,
- 733 Orlando, J. J., and Barsanti, K. C.: Multi-instrument comparison and compilation of non-
- 734 methane organic gas emissions from biomass burning and implications for smoke-derived 735 secondary organic aerosol precursors, Atmos. Chem. Phys., 17, 1471-1489, 2017.
- 736
- Hawthorne, S. B., Krieger, M. S., Miller, D. J., and Mathiason, M. B.: Collection and 737 738 quantitation of methoxylated phenol tracers for atmospheric pollution from residential wood
- 739 stoves, Environ. Sci. Technol., 23, 470-475, https://doi.org/10.1021/es00181a013, 1989.
- 740
- 741 Hough, W. A.: Caloric value of some forest fuels of the southern United States, Research Note,
- 742 USDA Forest Service, Southeastern Forest Experiment Station, Asheville, NC. [online] 743 Available from: http://www.treesearch.fs.fed.us/pubs/2778, 1969.
- 744
- 745 Hurst, D. F., Griffith, D. W. T., Carras, J. N., Williams, D. J., and Fraser, P. J.: Measurements of
- 746 Trace Gases Emitted by Australian Savanna Fires During the 1990 Dry Season, J. Atmos.
- Chem., 18, 33-56, 1994a. 747
- 748
- Hurst, D. F., Griffith, D. W. T., and Cook, G. D.: Trace gas emissions from biomass burning in 749 750 tropical Australian savannas, J. Geophys. Res., 99, 16441-16456, 1994b.
- 751
- 752 Ingemarsson, A., Nilsson, U., Nilsson, M., Pedersen, J. R., and Olsson, J. O.: Slow Pyrolysis of Spruce and Pine Samples Studied with GC/MS and GC/FTIR/FID, Chemosphere, 36-14, 2879-
- 753
- 2889, 1998. 754
- 755
- Johnson, T. J., Simon, A., Weil, J. M. and Harris, G. W., "Applications of time-resolved step-756
- 757 scan and rapid-scan FT-IR spectroscopy: Dynamics from ten seconds to ten nanoseconds"
- 758 Applied Spectroscopy, 47, 1376, (1993).
- 759





- Johnson, T. J., Masiello, T., and Sharpe, S. W.: The quantitative infrared and NIR spectrum of
- 761 CH2I2 vapor: vibrational assignments and potential for atmospheric monitoring, Atmos. Chem.
  762 Phys., 6, 2581-2591, 2006.
- 763
- Johnson, T. J., Profeta, L. T. M., Sams, R. L., Griffith, D.W. T., and Yokelson, R. L.: An
- infrared spectral database for detection of gases emitted by biomass burning, Vib. Spectrosc., 53,
  97–102, 2010.
- 767
- Jolly, W. M., Hintz, J., Linn, R. L., Kropp, R. C., Conrad, E. T., Parsons, R. A. and Winterkamp,
- 769 J.: Seasonal variations in red pine (*Pinus resinosa*) and jack pine (*Pinus banksiana*) foliar
- physio-chemistry and their potential influence on stand-scale wildland fire behavior, Forest
- 771 Ecology and Management, 373, 167–178, doi:<u>10.1016/j.foreco.2016.04.005</u>, 2016.
- 772
- Jolly, W. M., Parsons, R. A., Hadlow, A. M., Cohn, G. M., McAllister, S. S., Popp, J. B.,
- Hubbard, R. M. and Negron, J. F.: Relationships between moisture, chemistry, and ignition of *Pinus contorta* needles during the early stages of mountain pine beetle attack, Forest Ecology
  and Management, 269, 52–59, doi:10.1016/j.foreco.2011.12.022, 2012.
- 777
- Karl, T. G., Christian, T. J., Yokelson, R. J., Artaxo, P., Hao, W. M., and Guenther, A.: The
  Tropical Forest and Fire Emissions Experiment: method evaluation of volatile organic compound
  emissions measured by PTR-MS, FTIR, and GC from tropical biomass burning, Atmos. Chem.
- 781 Phys., 7, 5883-5897, 2007.
- 782

Keresztury, G., Billes, F., Kubinyi, M. and Sundius, T.: A Density Functional, Infrared Linear
Dichroism, and Normal Coordinate Study of Phenol and its Deuterated Derivatives: Revised

- 785 Interpretation of the Vibrational Spectra, J. Phys. Chem, 102, 1371-1380, 1998.
- 786
- Kibet, J., Khachatryan, L., and Dellinger, B.: Molecular products and radicals from pyrolysis of
   lignin, Environ. Sci. Technol., 46, 12994-13001, 2012.
- 789

790 Kochanov, R.V., Gordon, I.E., Rothman, L.S., Shine, K.P., Sharpe, S.W., Johnson, T.J.,

- Wallington, T.J., Harrison, J.J., Bernath, P.F., Birk, M. and Wagner, G.: Infrared absorption
   cross-sections in HITRAN2016 and beyond: Expansion for climate, environment, and
- rest atmospheric applications. Journal of Quantitative Spectroscopy and Radiative Transfer, 230,
- pp.172-221. 2019.
- 795
- 796 Koss, A. R., Sekimoto, K., Gilman, J. B., Selimovic, V., Coggon, M. M., Zarzana, K. J., Yuan,
- 797 B., Lerner, B. M., Brown, S. S., Jimenez, J. L., Krechmer, J., Roberts, J. M., Warneke, C.,
- 798 Yokelson, R. J., and de Gouw, J.: Non-methane organic gas emissions from biomass burning:
- 799 identification, quantification, and emission factors from PTR-ToF during the FIREX 2016
- laboratory experiment, Atmos. Chem. Phys., 18, 3299, 2018.
- 801
- Ledesma, E. B., Marsh, N. D., Sandrowitz, A. K. and Wornat, M. J.: An experimental study on the thermal decomposition of catechol, Proceedings of the Combustion Institute, 29(2), 2299–
- 2306, doi:10.1016/S1540-7489(02)80280-2, 2002.
- 805





- Liu, X., Huey, L. G., Yokelson, R. J., Selimovic, V., Simpson, I. J., Müller, M., Jimenez, J. L.,
- 807 Campuzano-Jost, P., Beyersdorf, A. J., Blake, D. R., Butterfield, Z., Choi, Y., Crounse, J. D.,
- 808 Day, D. A., Diskin, G. S., Dubey, M. K., Fortner, E., Hanisco, T. F., Hu, W., King, L. E.,
- 809 Kleinman, L., Meinardi, S., Milkoviny, T., Onasch, T. B., Palm, B. B., Peischl, J., Pollack, I. B.,
- 810 Ryerson, T. B., Sachse, G. W., Sedlacek, A. J., Shilling, J. E., Springston, S., St. Clair, J. M.,
- 811 Tanner, D. J., Teng, A. P., Wennberg, P. O., Wisthaler, A., and Wolfe, G. M.: Airborne
- measurements of western US wildfire emissions: Comparison with prescribed burning and air
- quality implications, J. Geophys. Res.-Atmos., 122, 6108–6129, doi: 10.1002/2016JD026315,
  2017.
- 814 815
- 816 Melvin, M. A.: 2015 National Prescribed Fire Use Survey Report, Technical Report, Coalition of
- 817 Prescribed Fire Councils, Inc. [online] Available from:
- 818 <u>http://stateforesters.org/sites/default/files/publication-</u>
- 819 documents/2015% 20Prescribed% 20Fire% 20Use% 20Survey% 20Report.pdf, 2015.
- 820
- 821 Matt, F. J., Dietenberger, M. A. and Weise, D. R.: Summative and ultimate analysis of live
- leaves from southern U.S. forest plants for use in fire modeling, Energy Fuels, (34), 4703–4720,
  doi:10.1021/acs.energyfuels.9b04107, 2020.
- 824
- 825 Neuman, J. A., Huey, L. G., Ryerson, T. B., and Fahey, D. W.: Study of Inlet Materials for
- 826 Sampling Atmospheric Nitric Acid, Environ. Sci. Technol., 33, 1133-1136, doi:
   827 10.1021/es980767f, 1999.
- 828
- 829 Paton-Walsh, C., Deutscher, N. M., Griffith, D. W. T., Forgan, B. W., Wilson, S. R., Jones, N.
- 830 B., and Edwards, D. P.: Trace gas emissions from savanna fires in northern Australia, J.
- 831 Geophys. Res., 115, doi:10.1029/2009JD013309, 2010.
- 832
- 833 Paton-Walsh, C., Smith, T. E. L., Young, E. L., Griffith, D. W. T. and Guérette, É.-A.: New
- emission factors for Australian vegetation fires measured using open-path Fourier transform
- infrared spectroscopy- Part 1: Methods and Australian temperate forest fires, Atmos. Chem.
- 836 Phys., 14, 11313-11333, doi: 10.5194/acp-14-11313-2014, 2014.
- 837
- Phillips, M. C., Myers, T. L., Johnson, T. J., and Weise, D. R.: In-situ measurement of pyrolysis
  and combustion gases from biomass burning using swept wavelength external cavity quantum
  cascade lasers, Optics Express, 28(6), 8680-8700, 2020
- 841
- 842 Prior, R. L., Cao, G., Martin, A., Sofic, E., McEwen, J., O'Brien, C., Lischner, N., Ehlenfeldt,
- 843 M., Kalt, W., Krewer, G. and Mainland, C. M.: Antioxidant Capacity As Influenced by Total
- Phenolic and Anthocyanin Content, Maturity, and Variety of Vaccinium Species, J. Agric. Food
  Chem., 46(7), 2686–2693, doi:10.1021/if980145d, 1998.
- Pyne, S. J.: World fire: the culture of fire on earth, Pbk. ed., University of Washington Press,
  Seattle., 1997.
- 848 Rao, P. V. R. and Rao, G. R.: Vibrational analysis of substituted phenols Part I. Vibrational
- spectra, normal coordinate analysis and transferability of force constants of some formyl-,





- methoxy-, formylmethoxy-, methyl- and halogeno-phenols, Spectrochimica ACTA Part A, 58,
  3039-3065, 2002.
- 852 Roscioli, J. R., Zahniser, M. S., Nelson, D. D., Herndon, S. C., and Kolb, C. E.: New Approaches
- to Measuring Sticky Molecules: Improvements of Instrumental Response Times Using Active
- Passivation, J. Phys. Chem. A, 120, 1347-1357, doi: 10.1021/acs.jpca.5b04395, 2015.
- 855 Safdari, M.-S., Rahmati, M., Amini, E., Howarth, J. E., Berryhill, J. P., Dietenberger, M., Weise,
- D. R., and Fletcher, T. H.: Characterization of pyrolysis products from fast pyrolysis of live and
- dead vegetation native to the Southern United States, Fuel, 229, 151-166, 2018.
- 858 Safdari, M.-S., Amini, E., Weise, D. R. and Fletcher, T. H.: Comparison of pyrolysis of live
- wildland fuels heated by radiation vs. convection, Fuel, 268, 117342,
- 860 doi:<u>10.1016/j.fuel.2020.117342</u>, 2020.
- 861
- 862 Saiz-Jimenez, C. and De Leeuw, J. W.: Lignin pyrolysis products: Their structures and their
- significance as biomarkers, Organic Geochemistry, 10(4–6), 869–876, doi:10.1016/S01466380(86)80024-9, 1986.
- 865
- Scharko, N. K., Oeck, A. M., Myers, T. L., Tonkyn, R. G., Banach, C. A., Baker, S. P., Lincoln,
- 867 E. N., Chong, J., Corcoran, B. M., Burke, G. M., Ottmar, R. D., Restaino, J. C., Weise, D. R. and
- Johnson, T. J.: Gas-phase pyrolysis products emitted by prescribed fires in pine forests with a
- shrub understory in the southeastern United States, Atmos. Chem. Phys., 19(15), 9681–9698,
- 870 doi:<u>10.5194/acp-19-9681-2019</u>, 2019a.
- 871 Scharko, N. K., Oeck, A. M., Tonkyn, R. G., Baker, S. P., Lincoln, E. N., Chong, J., Corcoran,
- 872 B. M., Burke, G. M., Weise, D. R., Myers, T. L., Banach, C. A., Griffith, D. W. T. and Johnson,
- 873 T. J.: Identification of gas-phase pyrolysis products in a prescribed fire: first detections using
- 874 infrared spectroscopy for naphthalene, methyl nitrite, allene, acrolein and acetaldehyde,
- 875 Atmospheric Measurement Techniques, 12(1), 763–776, doi:<u>10.5194/amt-12-763-2019</u>, 2019b.
- 876
- Scott, A. C., Bowman, D. M. J. S., Bond, W. J., Pyne, S. J. and Alexander, M. E.: Fire on earth:
  an introduction, John Wiley & Sons, Inc, Chichester, West Sussex., 2014.
- 879
- 880 Sekimoto, K., Koss, A. R., Gilman, J. B., Selimovic, V., Coggon, M. M., Zarzana, K. J., Yuan,
- 881 B., Lerner, B. M., Brown, S. S., Warneke, C., Yokelson, R. J., Roberts, J. M., and de Gouw, J.:
- 882 High-and low-temperature pyrolysis profiles describe volatile organic compound emissions from
- 883 western US wildfire fuels, Atmospheric Chemistry & Physics, 18, 2018.
- 884
- Selimovic, V., Yokelson, R. J., Warneke, C., Roberts, J. M., Gouw, J. d., Reardon, J., and
- 886 Griffith, D. W. T.: Aerosol optical properties and trace gas emissions by PAX and OP-FTIR for
- laboratory-simulated western US wildfires during FIREX, Atmos. Chem. Phys., 18, 2929-2948,
  2018.
- 889
- 890 Sharpe, S.W., Sams, R.L., Johnson, T.J., Chu, P.M., Rhoderick, G.C., and Guenther,
- F.R.: Creation of 0.10-cm<sup>-1</sup> resolution quantitative infrared spectral libraries for gas samples,





- 892 Proc. SPIE 4577, Vibrational Spectroscopy-based Sensor Systems, doi:
- 893 https://doi.org/10.1117/12.455730, 2002.
- 894
- Shimanouchi, T.: Tables of Vibrational Frequencies, Consolidated Vol. I. National Bureau ofStandards, 1972.
- 897
- Smith, T., Paton-Walsh, C. P., Meyer, G. C., Maier, S. W., Russell-Smith, J., Wooster, M., and
   Yates, C. P.: New emission factors for Australian vegetation fires measured using open-path
- 900 Fourier transform infrared spectroscopy-Part: Australian tropical savanna fire, Atmos. Chem.
- 901 Phys. 14, 14, 11335–11352, 2014.
- 902
- 903 Smith, T., Clare Paton-Walsh, C. P. Meyer, Garry Cook, Stefan W. Maier, Jeremy Russell-
- Smith, Martin Wooster, and Cameron P. Yates. "New emission factors for Australian vegetation
   fires measured using open-path Fourier transform infrared spectroscopy-Part 2: Australian
   tropical savanna fires." (2014): 11335.
- 907
- Stein, Y. S., Antal Jr, M. J., and Jones Jr., M.: A study of the gas-phase pyrolysis of glycerol, J.
  Anal. Appl. Pyrol., 4, 283–296, 1983.
- 910
- 911 Stockwell, C. E., Yokelson, R., Kreidenweis, S. M., Robinson, A. L., DeMott, P. J., Sullivan, R.
- 912 C., Reardon, J., Ryan, K. C., Griffith, D. W. T., and Stevens, L.: Trace gas emissions from
- 913 combustion of peat, crop residue, domestic biofuels, grasses, and other fuels: configuration and
- 914 Fourier transform infrared (FTIR) component of the fourth Fire Lab at Missoula Experiment
- 915 (FLAME-4), Atmos. Chem. Phys., 14, 9727-9754, 2014.
- 916
- Susott, R. A.: Characterization of the thermal properties of forest fuels by combustible gas
  analysis, For. Sci., 28(2), 404–420, 1982.
- 919 Thomas, S., Ledesma, E. B. and Wornat, M. J.: The effects of oxygen on the yields of the
- thermal decomposition products of catechol under pyrolysis and fuel-rich oxidation conditions,
  Fuel, 86(16), 2581–2595, doi:10.1016/j.fuel.2007.02.003, 2007.
- 922
- 923 Tihay, V. and Gillard, P.: Pyrolysis gases released during the thermal decomposition of three
- 924 Mediterranean species, Journal of Analytical and Applied Pyrolysis, 88(2), 168–174,
- 925 doi:<u>10.1016/j.jaap.2010.04.002</u>, 2010.
- 926
- Varhegyi, G., Jakab, E. and Antal, M. J.: Is the Broido-Shafizadeh Model for Cellulose Pyrolysis
  True?, Energy Fuels, 8(6), 1345–1352, doi:<u>10.1021/ef00048a025</u>, 1994.
- 929 Viatte, C., Strong, K., Hannigan, J., Nussbaumer, E., Emmons, L. K., Conway, S., Paton-Walsh,
- 930 C., Hartley, J., Benmergui, J. and Lin, J.: Identifying fire plumes in the Arctic with tropospheric
- FTIR measurements and transport models, Atmos. Chem. Phys., 15, 2227-2246, doi:, 2015.
- 932 Waldrop, T. A. and Goodrick, S. L.: Introduction to prescribed fires in southern ecosystems,
- 933 Science Update, USDA Forest Service, Southern Research Station, Asheville, NC. [online]
- Available from: <u>http://www.treesearch.fs.fed.us/pubs/41316</u>, 2012.





- 935 Ward, D. E. and Hao, W. M.: Projections of emissions from burning of biomass for use in
- 936 studies of global climate and atmospheric chemistry, 19 p., Air and Waste Management
- 937 Association, Vancouver, British Columbia, Canada. [online] Available from:
- 938 http://www.treesearch.fs.fed.us/pubs/43258, 1991.
- Ward, D. E., and Hardy, C. C.: Smoke emissions from wildland fires, Environ. Int., 17, 117-134,
  1991.
- 941 Ward, D. E. and Radke, L. F.: Emissions measurement from vegetation fires: a comparative
- 942 evaluation of methods and results, in Fire in the environment: the ecological, atmospheric, and
- climatic importance of vegetation fires: report of the Dahlem Workshop, held in Berlin, 15-20
- March 1992, edited by P. J. Crutzen and J. G. Goldammer, pp. 53–76, John Wiley & Sons Ltd.
- 945 [online] Available from: <u>http://www.fs.fed.us/rm/pubs\_other/rmrs\_1993\_ward\_d001.pdf</u>, 1993.
- 946 Ward, D. E.: Combustion chemistry and smoke, in Forest Fires: Behavior and Ecological Effects,
- edited by E. A. Johnson and K. Miyanishi, pp. 55–77, Academic Press, San Diego, CA. [online]
- 948 Available from: <u>http://www.doi.org/10.1016/B978-012386660-8/50006-3</u>, 2001.
- 949 Warneke, C., Roberts, J. M., Veres, P., Gilman, J., Kuster, W. C., Burling, I., Yokelson, R., and
- 950 de Gouw, J. A.: VOC identification and inter-comparison from laboratory biomass burning using
- 951 PTR-MS and PIT-MS, Int. J. Mass Spectrom., 303, 6–14, doi:10.1016/j.ijms.2010.12.002, 2011.
- 952 Weise, D. R., Fletcher, T. H., Cole, W., Mahalingam, S., Zhou, X., Sun, L., and Li, J.: Fire
- behavior in chaparral- Evaluating flame models with laboratory data, Combust. and Flame, 191,
  500-512, <u>https://doi.org/10.1016/j.combustflame.2018.02.012</u>, 2018.
- Weise, D. R., Johnson, T. J. and Reardon, J.: Particulate and trace gas emissions from prescribed
  burns in southeastern U.S. fuel types: Summary of a 5-year project, Fire Safety Journal, 74, 71–
  81, doi:10.1016/j.firesaf.2015.02.016, 2015.
- 958 Weise, D. R., Palarea-Albaladejo, J., Johnson, T. J., and Jung, H.: Analyzing Wildland Fire
- Smoke Emissions Data Using Compositional Data Techniques, J. Geophys. Res. Atmos., 125(6),
   https://doi.org/10.1029/2019JD032128, 2020.
- 961 Williams, S.D., Johnson, T.J., Sharpe, S.W., Yavelak, V., Oates, R.P. and Brauer, C.S.:
- 962 Quantitative vapor-phase IR intensities and DFT computations to predict absolute IR spectra
- 963 based on molecular structure: I. Alkanes. Journal of Quantitative Spectroscopy and Radiative
- 964 Transfer, 129, 298-307, <u>https://doi.org/10.1016/j.jqsrt.2013.07.005</u>, 2013
- Yang, H., Yan, R., Chen, H., Lee, D. H. and Zheng, C.: Characteristics of hemicellulose,
  cellulose and lignin pyrolysis, Fuel, 86, 1781–1788, 2007.
- 967 Yee, L. D., Kautzman, K. E., Loza, C. L., Schilling, K. A., Coggon, M. M., Chhabra, P. S.,
- 968 Chan, M. N., Chan, A. W. H., Hersey, S. P., Crounse, J. D., Wennberg, P. O., Flagan, R. C., and
- 969 Seinfeld, J. H.: Secondary organic aerosol formation from biomass burning intermediates: phenol
- 970 and methoxyphenols, Atmos. Chem. Phys., 13, 8019–8043, https://doi.org/10.5194/acp-13-8019-
- 971 2013, 2013.





- 972 Yokelson, R. J., Griffith, D. W. T. and Ward, D. E.: Open-path Fourier transform infrared studies
- of large-scale laboratory biomass fires, Journal of Geophysical Research, 101(D15), 21067,
  doi:10.1029/96JD01800, 1996.
- 975 Yokelson, R. J., Susott, R., Ward, D. E., Reardon, J., and Griffith, D. W. T.: Emissions from
- smoldering combustion of biomass measured by open-path Fourier transform infrared
- 977 spectroscopy, J. Geophys. Res. Atmos., 102, 18865-18877, 1997.
- 978 Yokelson, R. J., Christian, T. J., Bertschi, I. T., and Hao, W. M.: Evaluation of adsorption effects
- on measurements of ammonia, acetic acid, and methanol, J. Geophys. Res., 108, 4649,
  doi:10.1029/2003JD003549, 2003.
- 981 Yokelson, R. J., Burling, I. R., Gilman, J. B., Warneke, C., Stockwell, C. E., Gouw, J. d., Akagi,
- 982 S. K., Urbanski, S. P., Veres, P., Roberts, J. M., Kuster, W. C., Reardon, J., Griffith, D. W. T.,
- Johnson, T. J., Hosseini, S., Miller, J. W., Cocker III, D. R., Jung, H., and Weise, D. R.:
- 984 Coupling field and laboratory measurements to estimate the emission factors of identified and
- unidentified trace gases for prescribed fires, Atmos. Chem. Phys., 13, 89-116, 2013.
- 986 Zhou, X. and Mahalingam, S.: Evaluation of reduced mechanism for modeling combustion of
- 987 pyrolysis gas in wildland fire, Combustion Science and Technology, 171(1), 39–70,
- 988 doi:<u>10.1080/00102200108907858</u>, 2001.
- 989