

We thank the reviewer for her/his time and comments, which are reproduced in *black italic font* below. Our responses are shown in regular font. Revised text (as it appears in the manuscript) is shown in blue font. Line numbers are those of the revised manuscript with changes accepted.

Anonymous Referee #1 Received and published: 5 June 2020

This paper discusses the optimization of a HONO calibration source suitable for field work. For many instruments, the source needs to be relatively free of compounds that could interfere with the measurements of HONO. Current methods of HONO production often produce concentrations that vary with operating conditions (humidity, temperature, flow rates), as well as requiring long periods before stable outputs are achieved. Because of issues of stability and reproducibility, the output of HONO calibration methods need to be confirmed in the field with a secondary HONO instrument, often an absolute measurement technique such as absorbance spectroscopy or a chemiluminescent NO_x analyzer. Ideally, a field calibration source would provide stable and reproducible concentrations of HONO that could be quantified in the laboratory and reliably reproduced in the field, such that a secondary instrument would not be needed for source quantification.

The source described in this paper is based on the design of Febo et al. (1995), which has been used extensively by other groups as outlined in the manuscript. The authors describe changes in the design of the source that appear to minimize impurities such as ClNO and NO_x, and characterize the output of the source using FTIR and TDCRDS instruments. They find that their “optimized source” can produce concentrations of HONO with approximately 97% purity and is stable after 1.5 hours. The authors report HONO concentrations from the source ranging between 1.5 and 3 ppm from FTIR experiments and 11 and 50 ppb from TD-CRDS experiments (with some dilution occurring). The paper does provide some new information regarding the production of a stable, high purity HONO source for instrument calibrations and is likely of interest to the atmospheric chemistry community. While worthy of eventual publication, the paper does not provide sufficient information to give the reader confidence in reproducing the measurements. More experimental details regarding the individual flow rates, humidity, and temperatures should be provided. It is also not clear whether the source routinely produces concentrations of HONO with the stated purity over a range of operating conditions. The authors should include measurements of the HONO concentrations and impurities produced by the source over a range of temperatures and humidities that may be encountered when the source is used in the field. The authors should also provide information on the reproducibility of the source using the same operating conditions after it has been turn off. Illustrating that the output of the source is stable and reproducible from day-to-day might suggest that the source could be robust enough to be used for instrument calibrations without requiring a secondary instrument for quantification in the field.

Response: We appreciate the reviewer's point of view and agree that the manuscript would be strengthened by including a summary of our results with the optimized source in both the lab and the field.

We added the following on line 152 (to the materials and methods section):

"2.4 Field deployment

The HONO source was utilized during the "Study of nitrogen oxides in winter downwind from oil and gas sands" (SNOWDOGS) field campaign in Fort McKay, Alberta, Canada, in January 2020. Its output was quantified in parallel by TD-CRDS and a Thermo 42i-Y NO-NO_y CL instrument equipped

with a Mo converter heated to 325 °C. This converter and the TD-CRDS quartz inlets were mounted on the roof of a trailer which housed the instruments and the HONO source. The TD-CRDS sampled at a flow rate of ~1.2 slpm per channel through ~5 cm short, 300 µm i.d. stainless steel flow restrictors placed inline after the heated quartz sections and before the Teflon filter assembly. The HONO source output was delivered via a 5 m long, 1/4" (0.64 cm) o.d. and 3/16" (0.48 cm) i.d. FEP Teflon tube to both instruments."

We expanded the text on line 232:

"3.3 Source stability

The source output gradually decreased over a time scale of weeks of continuous use, which was rationalized by the visible depletion of the HCl permeation tube. However, the source output remained stable and reproducible on shorter time scales. An example FTIR time series is shown in Figure 5, which was acquired after 1 month of intermittent use. After a 1.5 hours stabilization period, the source produced a stable output of 1.57 ppmv of HONO (from >1.0 ppmv of HCl) with a precision of ±35 ppbv.

3.4 Source purity and day-to-day reproducibility

The optimized source routinely delivered HONO in high purity (range 96.0% - 98.7%), as long as scrubbed, moisture-containing room air was used as diluent gas (Table 2). When dry cylinder gas was used instead, the source output was stable but contained a larger amount of undesired side products; for example, on July 25, 2019, the output contained 4.6% NO₂ and 8.8% NO + ClNO (Table 2). When used in the field, the performance was similar: with scrubbed air, the HONO purity was 98.5%, whereas with dry cylinder gas, the purity was merely 81.3% (Table 2 and Figure S5)."

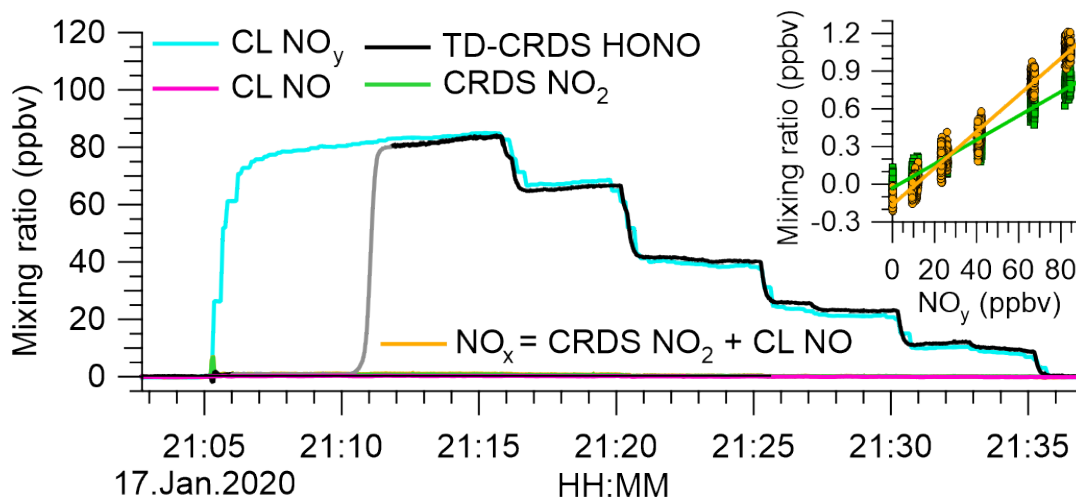
We added Table 2, which summaries our analysis results, on line 414:

"**Table 2.** Summary of TD-CRDS analyses of the HONO source output. The RH of the diluent gas was in the range of 15% to 35%, except for experiments conducted with cylinder gases which are shown below the dashed line. The range of mixing ratios stated is for output after dilution. The stated errors are from regression analyses of plots of NO₂, NO_x or NO_x+ClNO vs. NO_y and are at the 1σ level. n/d = not determined.

| Date | Setting | Diluent gas | Range (ppbv) | HONO (%) | NO ₂ (%) | NO _x (%) | NO _x +ClNO (%) |
|--------------|---------|-------------------------|--------------|-------------|---------------------|---------------------|---------------------------|
| Jan 16, 2019 | Lab | Scrubbed air | 0 - 140 | *97.2±0.1 | n/d | 2.80±0.07 | n/d |
| May 3, 2019 | Lab | Scrubbed air | 0 - 7 | 96.0±0.2 | 1.9±0.2 | 4.0±0.2 | (4.0±0.2)** |
| May 6, 2019 | Lab | Scrubbed air | 0 - 93 | 96.4±0.4 | 1.9±0.4 | 3.6±0.4 | (3.6±0.4)** |
| May 10, 2019 | Lab | Scrubbed air | 0 - 21 | *97.6±0.1 | 1.10±0.03 | 2.40±0.05 | n/d |
| May 17, 2019 | Lab | Scrubbed air | 0 - 38 | 98.7±0.1 | 0.01±0.02 | 0.92±0.01 | 1.25±0.01 |
| May 22, 2019 | Lab | Scrubbed air | 0 - 14 | 97.7±0.1 | 1.29±0.06 | 1.54±0.06 | 2.28±0.04 |
| Jan 17, 2020 | Field | Scrubbed air | 0 - 80 | 98.5±0.1 | 1.00±0.05 | 1.50±0.08 | (1.50±0.08)** |
| Mar 19, 2020 | Lab | Scrubbed air | 0 - 320 | *98.72±0.01 | n/d | 1.28±0.01 | n/d |
| Jul 25, 2019 | Lab | Cylinder O ₂ | 0 - 95 | 86.60±0.02 | 4.60±0.03 | n/d | 13.4±0.02 |
| Jan 30, 2020 | Field | Cylinder N ₂ | 0 - 50 | 81.3±0.1 | 5.90±0.09 | n/d | 18.7±0.1 |

* Upper limit. ** The TD-CRDS quartz inlet temperature was ramped between 300 °C and 600 °C; the TD-CRDS mixing ratios at an inlet temperature of 300 °C matched the NO_x mixing ratios observed with the room temperature inlet, indicating the absence of ClNO."

To the SI, we added



"Figure S5. Sample field analysis of the HONO source output. The TD-CRDS and CL instruments sampled scrubbed "zero" air before 21:05 and after 21:36. The HONO source output was added at 21:05:30. Only the CL NO_y responded because the TD-CRDS quartz inlet temperature was 200 °C. At 21:10, the temperature of the quartz inlet was increased from 200 °C to 600 °C to quantify HONO. The absence of an inflection point and prior agreement with the NO_x measurement implies the absence of ClNO. At 21:16 and every ~5 min thereafter, the HONO output concentration was decreased by slightly opening the bypass valve. The insert shows scatter plots of NO₂ and NO_x (calculated by adding CL NO and CRDS NO₂ data) against NO_y. Slopes and offsets were (0.96±0.05)% and -(30±3) pptv for NO₂ (points shown in green) and (1.46±0.08)% and -(164±4) pptv for NO_x (data points shown in orange), respectively."

Finally, we revised the statement given in the abstract on line 15:

"The source produces gas streams containing HONO in air in > ~~95%~~ 97% purity relative to other nitrogen oxides." to account for the observation of (96±0.2)% purity on May 3, 2019 (Table 2).

Specific comments:

Methods: As mentioned in the paper, previous studies have demonstrated that the production of HONO by reaction R4 requires the gas stream to be humidified. However, the authors do not state the humidity of the gas stream used in their experiments.

Response: We added the following on line 88: "For the experiments shown in this manuscript, the relative humidity of the diluent gas stream was in the range of 15% to 35%. "

Page 4, line 88: Is there any reason for the 20 ccm dilution with O₂?

Response: The additional flow is probably not essential, but we found it practical to keep part of the setup under flow of a clean gas - our laboratory air contains surprisingly high levels of NO_x delivered from the building's air intake, plus the continued flow would remove any impurities that might build up. The use of a second mass flow controller was also advantageous as it allowed the residence time and HCl concentration to be changed on the fly without disturbing the permeation tube setup (which can take quite a long time to return to a stable output when flows are changed).

We modified the text on line 90 as follows:

"The permeation tube was placed inside a glass chamber (VICI Dynacalibrator Model 120) whose temperature was controlled at 25.0 °C and which was continuously flushed at a flow rate of ~0.15 L min⁻¹ with room air scrubbed using activated charcoal for a period of several days [into a fume hood](#). The use of scrubbed air ensured that the gas stream contained water to maintain efficient HONO production (Schiller et al., 2001).

The HCl output was diluted in a gas stream of O₂ (~20 mL min⁻¹); [this gas stream also served to continuously flush the connecting tubing when not in use.](#)"

If so, why O₂ and not N₂, zero air, or scrubbed room air?

Response: The reviewer is correct that either N₂, zero air or scrubber air could have been used. We chose O₂ because an O₂ cylinder was already in place for the O₃ generator. No changes were made.

Page 4, last paragraph: How much dilution was typical? What are final flow rates? More experimental details should be provided.

Response: We diluted the output by factors between 5 and 200, depending on how much HONO we desired to deliver on any particular day at a flow rate marginally larger than the TD-CRDS inlet sample flow. We modified the paragraph as follows:

"To deliver HONO in atmospheric concentrations (i.e., < 10 ppbv), a portion of the source output was directed towards waste with the aid of a pump and a needle valve. The output concentration could be rapidly changed ([typically by factors between 5 to 200](#)) by adjusting the position of the needle valve. The remaining output was diluted using scrubbed air and directed towards the instruments [at a final flow rate slightly larger than the sampling requirement of the instruments.](#)"

Fig. 1: The position of the needle valve and dilution do not match the written description. The figure only shows the input of the TD-CRDS being diluted and not the FTIR, while the text describes dilution before both instruments.

Response: As stated on line 107, the needle valve was used to deliver HONO in atmospheric concentrations (i.e., < 10 ppbv). The FTIR sampled the output without dilution, and the Figure is accurate as shown. We clarified this on line 111:

"Gas streams exiting the HONO sources (prior to dilution) were analysed using an FTIR spectrometer"

The schematic would benefit with typical flow rates.

Response: This information has been added to Figure 1.

Page 7, line 161: In this discussion, the authors are probably referring to reaction R5, and not R6.

Response: We thank the reviewer for catching this error. It has been corrected (line 177). We also corrected another error of this type on line 165.

Page 7, line 169: It appears that the improvement in source 2 is only a reduction of ClNO contamination, as both the NO and NO₂ concentrations increased. This should be clarified.

Response: The reviewer's observation is correct. The issue with both source 1 and source 2 was that there was too much HONO produced, which can generate side products by either R-1 or R5. In source 2, more HONO was produced than in source 1 (because the rate of R5 was reduced) which increased the rate of NO and NO₂ produced via R-1.

We added the following on line 185: "... an improvement over source 1 in terms of unwanted ClNO production but still inadequate."

Page 8, equation 2: Check the equation – there appears to be an error translating the fonts.

Response: Apologies - this was an error converting word (.docx) to pdf format. Equation (2) is

$$[\text{Observed}]_{\text{total}} = [\text{NO}_x] + [\text{ClNO}] \left(1 - e^{-A_{\text{ClNO}} \times e^{\frac{-E_{\text{A,ClNO}}}{RT}} \times t_{\text{res}}} \right) + [\text{HONO}] \left(1 - e^{-A_{\text{HONO}} \times e^{\frac{-E_{\text{A,HONO}}}{RT}} \times t_{\text{res}}} \right)$$

This has been corrected in the revised document.

Fig. 5: The authors should define the lighter shaded points in both plots. Are the lighter shades indicating points while the source is stabilizing? If so, why are there lighter shade points at the end of the time series?

Response: Our apologies - the two lighter shade points at the end of the time series should have been in dark shade as well; this has been corrected. We also modified the caption as requested.

"Figure 5. Time series of HONO and HCl mixing ratios derived from FTIR analysis of the undiluted HONO source output. The NaNO₂ was placed in line at 10:10. Data acquired during the initial period are shown in light grey colour. The shaded areas represent the average $\pm 2\sigma$ after the source output had stabilized (after 11:45; data points shown in black colour). The 1σ precision of the HONO data was ± 35 ppbv and that of the HCl data was ± 42 ppbv. The HCl mixing ratios are an underestimate because the widths of their absorption lines are less than the FTIR's resolution of 0.5 cm^{-1} ."

The authors should clarify how they determined that the output had stabilized.

Response: In the caption of Figure 5, we had stated "The shaded areas represent the average $\pm 2\sigma$ after the source output had stabilized (after 11:45; data points shown in black colour)."

Since we didn't apply a mathematically rigorous criterion to determine output stability, we changed the above to:

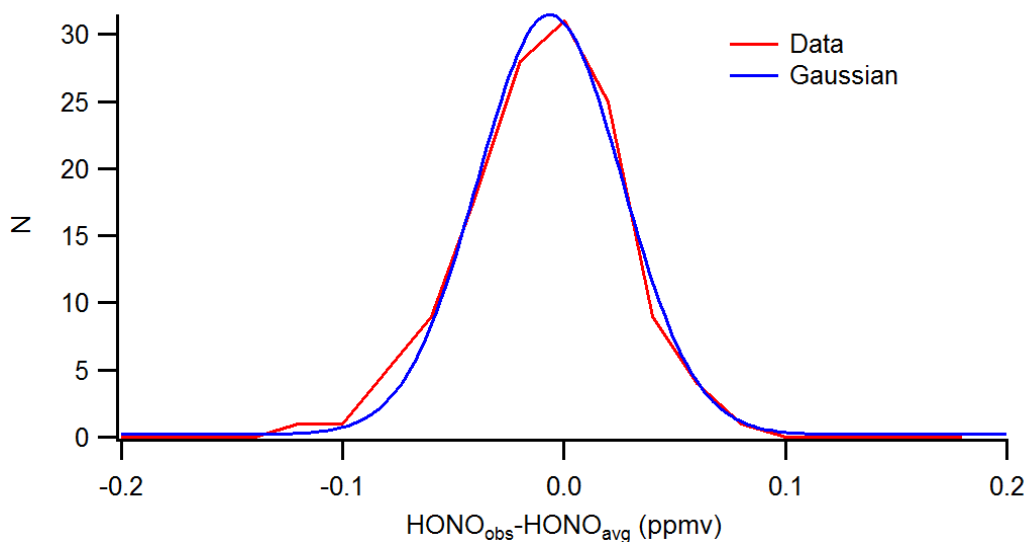
"The shaded areas represent the average $\pm 2\sigma$ after the source output had ~~had~~ was judged to have stabilized (after 11:45; data points shown in black colour)."

We had judged whether the output was stable (or not) by visual inspection but also had performed a few simple statistical tests for corroboration.

A stable output is achieved when the output mixing ratio is not changing. For the data shown in black, this criterion is met since the slope of a linear regression fit is $-(0.4 \pm 1.9) \times 10^{-6}\text{ s}^{-1} \approx 0$.

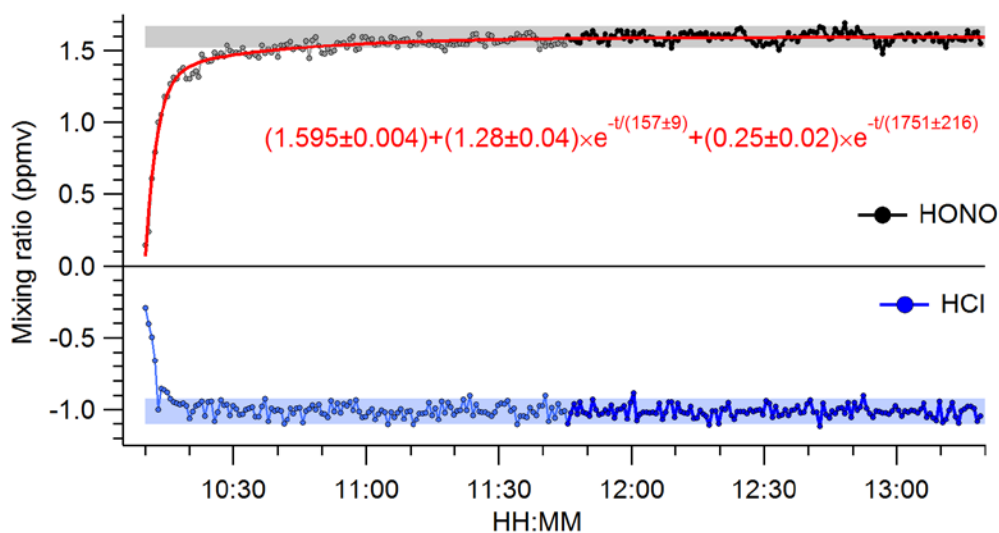
We also calculated the average (μ) and standard deviation (σ) for this period which were 1.596 and ± 0.036 ppmv. The residuals during this time period were normally distributed (i.e., reproduced by a Gaussian equation, see figure below).

The $\mu \pm 2\sigma$ interval is shown as a grey area in Figure 5; since the noise is normally distributed, this interval is expected to encompass 95% of the data; for the data shown in black, 6 of the 132 (~4.5%) are outside this confidence interval.



The above confirms that the output during the chosen period was stable (i.e., confirms that "the source output had stabilized" as we had stated in the caption of Figure 5), but it does not justify the choice of the left limit to what is called a stable period in Figure 5 (which we had estimated at 11:45:30).

Expanding on the above discussion, we applied a double exponential fit to the HONO data in Figure 5:



The first time constant of this fit was (157 ± 9) s, corresponding to the mixing time of gas in the FTIR multipass cell (assuming plug flow 450 cm^3 at a flow rate of $170 \text{ cm}^3 \text{ min}^{-1} \sim 160$ s). The second time constant was 29.3 ± 3.6 min. The fit is within 1% of the limiting mean at 11:46:56 in Figure 5, which is very close to the "eye-ball" estimate.

We thank the reviewer for her/his time and comments, which are reproduced in *black italic font* below. Our responses are shown in regular font. Revised text (as it appears in the manuscript) is shown in blue font. Line numbers are those of the revised manuscript with changes accepted.

Anonymous Referee #2 Received and published: 12 June 2020

The authors present an optimized HONO calibration source that is based on the design of Febo et al., 1995, where HCl vapor is passed over solid NaNO₂. They used a permeation tube to achieve lower (< 4 ppmv) HCl mixing ratios which is key to achieve higher purity as ClNO formation is slowed down. They quantified the impurities by using FTIR spectroscopy and TD-CRDS and found the optimized design of the source to be of > 97 % purity. With the optimized design (lower HCl supply) they were able to generate HONO concentrations in the low ppm range and by further dilution in the lower ppb range. The stabilization time of 1.5 h was short compared to other source designs.

Furthermore, the source is portable and after stabilization, HONO mixing ratios are readily tunable. Due to its instability, there are no permeation tubes or standard gases for HONO available and it must be produced in situ. Therefore, to calibrate gas phase mixing of HONO such a compact and easy deployable source is of interest for the atmospheric chemistry community. The study is well performed and the manuscript well written. Therefore, I support publication after considering the minor comments given below.

Response: We appreciate the reviewer's supportive comments.

General comments:

As surfaces are unavoidable in laboratory setups, the role of surface reactions should be discussed. Although I guess that HONO formation from NO₂ impurities is not of importance, there are also heterogeneous decomposition reactions for HONO that form NO (and NO₂) and might therefore important to keep impurities low. See esp. (Finlayson-Pitts et al., 2003).

Response: We agree with the reviewer that surface chemistry is an important consideration. Finlayson-Pitts et al. (2003) discussed the mechanism of heterogeneous conversion of NO₂ (to HONO and HNO₃) on Pyrex which involves formation and wall-adsorption of NO₂ dimer (i.e., N₂O₄) as the initial steps. Another paper by the same group (Syomin and Finlayson-Pitts, 2003) discusses HONO decomposition on glass surfaces, which was linked to wall-adsorbed HONO reacting with wall-adsorbed HNO₃.

We believe this chemistry is limited in our setup as we have taken several steps to avoid it becoming factor. First of all, the chemistry described by Finlayson-Pitts et al. (2003) and Syomin and Finlayson-Pitts (2003) was observed in a static system on a time scale of hours or required cell conditioning with HNO₃; in our system, the 50 mL reaction vessel was continuously flushed (at a flow of ~150 mL min⁻¹ from the zero air generator when in use and continuously with ~20 mL min⁻¹ from the O₂ cylinder), such that there was little time for this chemistry to create much of an effect. In addition, formation of N₂O₄ scales with the square of the NO₂ concentration, which was low in our experiments. More importantly, the Pyrex reaction vessel was heated, which promotes partitioning of wall-adsorbed species to the gas-phase and reduces the extent of surface chemistry. Furthermore, the equilibrium

constant for the reaction $2\text{NO}_2 \rightleftharpoons \text{N}_2\text{O}_4$ is temperature-dependent favoring N_2O_4 dissociation at higher temperature. The NASA-JPL compilation recommends an equilibrium constant of $5.9 \times 10^{-29} \text{ cm}^3 \text{ molecule}^{-1} e^{(6643/T)}$ (Burkholder et al., 2015). When the temperature is increased from room temperature to 50 °C, the equilibrium constant decreases by a factor of ~6.

The following was added to the manuscript on line 97:

"In the second source ("source 2"), the NaNO_2 was placed in a two-neck, 50 mL Pyrex round bottom flask which was covered in aluminium foil to prevent photolysis of nitrite and nitrous acid and was externally heated to a temperature of 50 °C using a water bath and mechanically agitated using a magnetic stir bar as described by Febo et al. (1995). Heating this vessel promotes dissociation of molecular clusters such as N_2O_4 and partitioning of wall-adsorbed molecules to the gas-phase, which, if present, can drive HONO decomposition on borosilicate glass surfaces (Syomin and Finlayson-Pitts, 2003)."

Another consideration is surface chemistry in the connecting tubing. We constructed all our connecting tubing out of inert Teflon material and kept it under continuous flow (with an inert gas such as N_2 or O_2) when not in use (a detail we added to the manuscript - see our response to the question on L88 below).

Regarding the FTIR measurements: Can the authors provide more details about the reference spectrum used for HONO (spectrum of cis or trans isomer or total spectrum? taken at which temperature?). The spectral features will change with temperature as the amounts of cis and trans isomers of HONO change with temperature (e.g. Barney et al., 2000). Furthermore, please provide temperature and humidity values of the gas stream if possible.

Response: The reference spectrum was obtained from the Pacific Northwest National Laboratory digital library (Sharpe et al., 2004). According to their meta data file, they acquired this spectrum at 25 °C and atmospheric pressure and with dry nitrogen as diluent gas and generated HONO in situ from the reaction of NaNO_2 with HCl. We would hence expect the same distribution of HONO isomers as we would have generated in our experiments, which were conducted at room temperature.

We added more detail to section 2.2 ("Analysis of HONO source output by FTIR") on line 115 as requested by the reviewer:

"Room-temperature spectra were acquired continuously at a time resolution of 30 s. Mixing ratios of trace gases were determined from fits (by least squares error minimization over a selected wavelength range) of room-temperature reference spectra from the Pacific Northwest National Laboratory (Sharpe et al., 2004), the pressure inside the multi-pass cell (which was equal to that of the room) was monitored using a pressure transducer (Omegadyne PX419-015A5V)."

Because the bulk of the diluent gas was generated by passing room air through a scrubber, the relative humidity of the gas was approximately the same as the room (~30%; accounting for dilution with dry oxygen and neglecting H_2O co-emitted by the HCl perm tube, RH is estimated at ~25% for the data shown in Figure 2). Since the background spectrum was acquired with this H_2O background, the FTIR only shows drifts in H_2O . The following was added on line 187:

"A sample FTIR spectrum of this source's output is shown in Figure 2. With a freshly prepared HCl permeation tube and an estimated relative humidity in the round bottom flask of ~25%, > ~2.5 ppmv HCl were consumed and ~3.0 ppmv of HONO were produced. "

Specific comment:

L88: Why diluting with 20 mL min⁻¹ flow of oxygen?

Response: The additional flow is probably not essential, but we found it practical to keep part of the setup under flow of a clean gas - our laboratory air contains surprisingly high levels of NO_x delivered from the building's air intake, plus the continued flow would remove any impurities that might build up. The use of a second mass flow controller was also advantageous as it allowed the residence time and HCl concentration to be changed on the fly without disturbing the permeation tube setup (which can take quite a long time to return to a stable output when flows are changed).

We modified the text as follows:

"The permeation tube was placed inside a glass chamber (VICI Dynacalibrator Model 120) whose temperature was controlled at 25.0 °C and which was continuously flushed at a flow rate of ~0.15 L min⁻¹ with room air scrubbed using activated charcoal for a period of several days into a fume hood. The use of scrubbed air ensured that the gas stream contained water to maintain efficient HONO production (Schiller et al., 2001). The HCl output was diluted in a gas stream of O₂ (~20 mL min⁻¹); this gas stream also served to continuously flush the connecting tubing when not in use."

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A compact, high-purity source of HONO validated by Fourier Transform Infrared and Thermal Dissociation Cavity Ring-down Spectroscopy

Nicholas J. Gingerysty¹ and Hans D. Osthoff¹

¹ Department of Chemistry, University of Calgary, 2500 University Drive N.W., Calgary, Alberta, Canada T2N 1N4

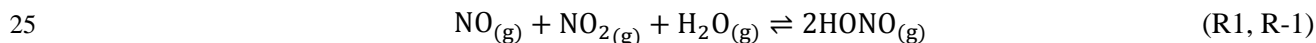
5 Correspondence to: Hans D. Osthoff (hosthoff@ucalgary.ca)

Abstract

A well-characterized source of nitrous acid vapour (HONO) is essential for accurate ambient air measurements by instruments requiring external calibration. In this work, a compact HONO source is described in which gas streams containing dilute concentrations of HONO are generated by flowing
10 hydrochloric acid (HCl) vapour emanating from a permeation tube over continuously agitated dry sodium nitrite (NaNO₂) heated to 50 °C. Mixing ratios of HONO and potential by-products including NO, NO₂ and nitrosyl chloride (ClNO) were quantified by Fourier Transform Infrared (FTIR) and thermal dissociation cavity ring-down spectroscopy (TD-CRDS). A key parameter is the concentration of HCl, which needs to be kept small (< 4 ppmv) to avoid ClNO formation. The source produces gas streams containing HONO in
15 air in > 95.7% purity relative to other nitrogen oxides. The source output is rapidly tuneable and stabilizes within 90 min. Combined with its small size and portability this source is highly suitable for calibration of HONO instruments in the field.

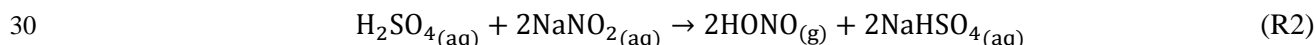
1 Introduction

20 The generation of nitrous acid vapour ($\text{HONO}_{(g)}$) free of other nitrogen oxides has been a long-standing challenge to atmospheric chemists (Table 1). Such sources are needed in kinetic studies, for absorption cross-section measurements, and for the calibration of field instruments. Nitrous acid is not suitable for permeation devices since HONO is difficult to prepare in low concentration and in high purity and disproportionates to nitric oxide and nitrogen dioxide via equilibrium (1).

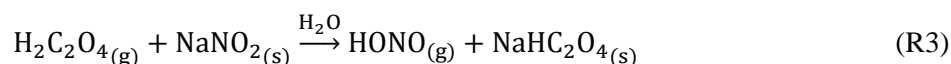


In fact, early experiments relied on the NO, NO₂ and H₂O vapour equilibrium to form HONO in situ (King and Moule, 1962; Cox, 1974; Stockwell and Calvert, 1978).

Cox and Derwent prepared HONO vapours in ~50% purity by flowing nitrogen over an aqueous solution containing 0.1 M sodium nitrite with and 1.4%-2.5% sulfuric acid (Cox and Derwent, 1976).

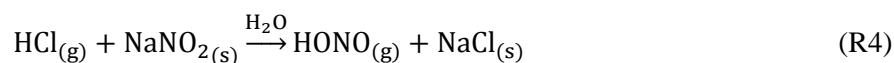


Braman and de la Cantera sublimed oxalic acid onto solid sodium nitrite and were able to produce gas streams in >50% and up to 90% purity as long as water was present (Braman and De la Cantera, 1986).



Subsequent refinements included a flow reactor design in solutions of sulfuric acid and sodium nitrite were
35 dynamically mixed, producing a stable output in >90% purity (Taira and Kanda, 1990).

Febo et al. (1995) developed a method in which gas-phase hydrochloric acid emitted from a permeation device was quantitatively reacted in a humidified gas stream with solid sodium nitrite, producing a stable and highly pure (>99.5%) HONO output.



40 The output of sources based on R4 has been characterized by differential optical absorption spectroscopy (DOAS) (Febo et al., 1995; Stutz et al., 2000), Fourier transform infrared (FTIR) spectroscopy (Brust et al., 2000; Schiller et al., 2001), tuneable diode laser absorption spectroscopy (TLDAS) (Schiller et al., 2001), incoherent cavity-enhanced absorption spectroscopy (IBBCEAS) (Roberts et al., 2010), and long-path optical absorption photometry (LOPAP) (Ren et al., 2010). Stutz et al. (2000) noted that concentrations of

45 HONO should be kept low ($< 10^{14}$ molecules cm^{-3} ; ~ 4 ppmv) to avoid disproportionation of HONO via R-1.

Perez et al. (2007) examined a source based on the design by Febo et al. (1995) using thermal-dissociation chemiluminescence (TD-CL) detection of NO and reported that their source co-emitted nitrosyl chloride (ClNO); they opted to generate HONO from reaction of H_2SO_4 aerosol with $\text{NaNO}_2(\text{s})$ instead. In contrast,
50 Roberts et al. (2010) observed little or no production of ClNO ($< 4.5\%$) and quantitative conversion of HCl to HONO, at odds with Perez et al. (2007) but in agreement with the original work by Febo et al. (1995). As far as we know, this inconsistency is unexplained to date.

As part of our development of an IBBCEAS for HONO measurement, our lab recently constructed a HONO source using an aqueous solution of NaNO_2 buffered with oxalate to pH 3.74 (Jordan and Osthoff, 2020).
55 This source generated HONO in trace amounts suitable to our needs but co-emitted NO and NO_2 in similar ratios as reported by Braman and De la Cantera (1986). Furthermore, this type of source could not be turned off (i.e., had to remain under continuous N_2 flow) and required long times to stabilize, motivating us to develop an alternative HONO generation method. Ambient air HONO measurement techniques that need to be externally calibrated often poorly agree with each other (Crilley et al., 2019), further motivating the
60 development of a well-characterized, compact and portable HONO source and to improve the understanding of conditions needed to suppress formation of undesired side products such as ClNO.

In this work, we describe the implementation of a compact, high-purity HONO generation device based on R4 for field deployment and laboratory experiments and characterized its output by FTIR and thermal dissociation cavity ring-down spectroscopy (TD-CRDS). Conditions to avoid generation of impurities such
65 as ClNO are identified. The performance of this source in comparison to existing methods is discussed.

2 Materials and methods

2.1. Generation of gas streams containing HONO

A practical challenge is the generation of stable and dilute gas streams containing HCl. Calibration gas
70 cylinders are the most straightforward method to use for this purpose but may require periods of up to 10 days to stabilize (Roberts et al., 2010). Furthermore, low-concentration HCl gas cylinders are expensive to source, such that we chose to generate HCl gas through the use of relatively inexpensive permeation devices.

In the setup by Febo et al. (1995), which was also implemented by Brust et al. (2000), Stutz et al. (2000) and Ren et al. (2010), a gas stream of N₂ is passed through Teflon tubing immersed in an HCl bath. This setup involved a rather large 1 L vessel containing liquid HCl which is somewhat impractical and potentially hazardous in a field setting. With portability in mind, we decided to construct a permeation tube containing HCl(l). Literature does not inform as to suitable tube dimensions, which were determined through trial-and-error.

Three HONO sources were constructed and evaluated. In the first ("source 1"), a 24 cm long, polytetrafluoroethylene (PTFE) permeation tube (Chromatographic Specialties C111LE; wall thickness 1.5 mm; outer diameter (o.d.) 7.8 mm) was filled with 2.5 mL of 37% HCl (Sigma-Aldrich) and sealed at both ends with PTFE plugs held in place by stainless steel compression rings. The permeation tube was placed inside a glass chamber (VICI Dynacalibrator Model 120) whose temperature was controlled at 25.0 °C and which was continuously flushed at a flow rate of ~0.15 L min⁻¹ with room air scrubbed using activated charcoal for a period of several days into a fume hood. The use of scrubbed air ensured that the gas stream contained water to maintain efficient HONO production (Schiller et al., 2001). For the experiments shown in this manuscript, the relative humidity of the diluent gas stream was in the range of 15% to 35%.

The HCl output was diluted in a gas stream of O₂ (~20 mL min⁻¹); this gas stream also served to continuously flush the connecting tubing when not in use. Roughly 1.5 g of solid NaNO₂ was placed inside a 50 cm long loop of 3/16" (0.476 cm) inner diameter (i.d.) and ¼" (0.635 cm) o.d. fluorinated ethylene propylene (FEP) Teflon tubing downstream from the HCl addition point. This section could be manually bypassed with a pair of 3-way valves (Entegris) to turn HONO production on or off. A Teflon filter (Pall, 2 µm pore size and 47 mm diameter) inside a Teflon filter holder (Cole-Parmer) was placed downstream of the NaNO₂ powder.

In the second source ("source 2"), the NaNO₂ was placed in a two-neck, 50 mL Pyrex round bottom flask which was covered in aluminium foil to prevent photolysis of nitrite and nitrous acid and was externally heated to a temperature of 50 °C using a water bath and mechanically agitated using a magnetic stir bar as described by Febo et al. (1995). Heating this vessel promotes dissociation of molecular clusters such as N₂O₄ and partitioning of wall-adsorbed molecules to the gas-phase, which, if present, can drive HONO decomposition on borosilicate glass surfaces (Syomin and Finlayson-Pitts, 2003).

The third source ("optimized source") used the same setup as source 2, except that a shorter permeation tube (length 6 cm) containing 0.35 mL of HCl was used. This setup is depicted in Figure 1. The overall dimensions are 28 cm × 28 cm × 36 cm and its modular design ensures easy transport to and from the field.

To deliver HONO in atmospheric concentrations (i.e., < 10 ppbv), a portion of the source output was directed towards waste with the aid of a pump and a needle valve. The output concentration could be rapidly changed (typically by factors between 5 to 200) by adjusting the position of the needle valve. The remaining output was diluted using scrubbed air and directed towards the instruments at a final flow rate slightly larger than the sampling requirement of the instruments.

2.2 Analysis of HONO source output by FTIR

Gas streams exiting the HONO sources (prior to dilution) were analysed using an FTIR spectrometer (Bruker Tensor 27) equipped with a liquid nitrogen cooled mercury cadmium telluride (MCT) detector and a White multi-pass gas cell with a 6.4 m optical path length and an internal volume of 0.75 L (Gemini Scientific Instruments, Venus series) in a similar fashion as described earlier (Taha et al., 2013). Room-temperature spectra ~~Spectra~~ were acquired continuously at a time resolution of 30 s. Background spectra were recorded with HCl inline but with the NaNO₂ bypassed, which resulted in the observed spectra showing the change in trace gas concentrations.

Mixing ratios of trace gases were determined from fits (by least squares error minimization over a selected wavelength range) of room-temperature reference spectra from the Pacific Northwest National Laboratory (Sharpe et al., 2004), multiplied by the respective mixing ratios as variables, to the observed spectra. The reference spectra are provided in units of ppmv at atmospheric pressure per meter of optical length, necessitating a correction factor of 6.4 accounting for the actual path length and a pressure correction factor, for which the pressure inside the multi-pass cell (which was equal to that of the room) was monitored using a pressure transducer (Omegadyne PX419–015A5V).

The FTIR limits of detection (LODs) are specific to each molecule (due to differing absorption cross-sections) and are in the 100 to 300 parts-per-billion (by volume; ppbv; 10⁻⁹) range. For example, the 1σ precision of NO₂ data was ±100 ppbv, yielding a 3σ LOD of 300 ppbv.

The spectral resolution of the FTIR was ~0.5 cm⁻¹, which does not suffice to fully resolve the absorption lines of HCl, H₂O and NO; their FTIR derived mixing ratios are hence lower limits.

2.3 Analysis of HONO source output by TD-CRDS

The (diluted) HONO source output was also analysed using a four-channel TD-CRDS instrument (Odame-Ankrah, 2015). Briefly, mixing ratios of NO₂ are quantified by absorption at 405 nm. Ring-down time constants in the absence (τ_0) and presence (τ) of NO₂ are converted to concentrations (N) using equation (2) where c is the speed of light, σ is the NO₂ absorption cross-section, and R_L is a correction factor (~1.2) accounting for mirror purge flows (Paul and Osthoff, 2010).

$$N = \frac{R_L}{c\sigma} \left(\frac{1}{\tau} - \frac{1}{\tau_0} \right) \quad (1)$$

Concentrations are converted to mixing ratios using the ideal gas law. Other NO_y components are converted to NO₂ in separate channels and are quantified by difference.

For this study, the instrument was operated as follows: Mixing ratios of NO₂ were monitored using a room temperature, 1/4" (0.64 cm) o.d. and 1/8" (0.32 cm) i.d. FEP Teflon inlet. Mixing ratios of NO_x were quantified on a second channel with a room temperature Teflon inlet and by adding O₃ (mixing ratio after addition ~3 ppmv) to titrate NO to NO₂ (Odame-Ankrah, 2015; Fuchs et al., 2009). A third channel was operated with a 1/4" (0.64 cm) o.d. quartz inlet heated to 600 °C. Ozone was added between the quartz inlet and the CRDS cell to quantify NO_y, including HONO (Jordan and Osthoff, 2020; Wild et al., 2014; Perez et al., 2007). The fourth inlet was operated at 350 °C and with added O₃. This channel quantified NO_x + ΣPAN + ΣAN + CINO + CINO₂ and was used as a HONO reference channel. Each channel sampled at a flow rate of ~0.8 slpm through ~5 cm short, 1/16" i.d. Teflon flow restrictors placed inline after the heated quartz sections (if existing) and before a 50 mm Teflon (Pall Teflo, 2 µm pore size) housed in a Teflon filter holder (Cole Parmer). The laser pulse repetition rate was 1500 Hz, and 1500 ring-down events were averaged to produce 1 s data.

2.4 Field deployment

The HONO source was utilized during the "Study of nitrogen oxides in winter downwind from oil and gas sands" (SNOWDOGS) field campaign in Fort McKay, Alberta, Canada, in January 2020. Its output was quantified in parallel by TD-CRDS and a Thermo 42i-Y NO-NO_y CL instrument equipped with a Mo converter heated to 325 °C. This converter and the TD-CRDS quartz inlets were mounted on the roof of a trailer which housed the instruments and the HONO source. The TD-CRDS sampled at a flow rate of ~1.2 slpm per channel through ~5 cm short, 300 µm i.d. stainless steel flow restrictors placed inline after the heated quartz sections and before the Teflon filter assembly. The HONO source output was delivered via a 5 m long, 1/4" (0.64 cm) o.d. and 3/16" (0.48 cm) i.d. FEP Teflon tube to both instruments.

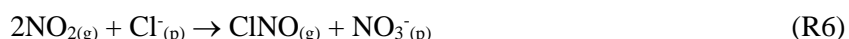
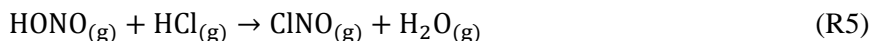
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3 Results

3.1 Development of a compact, high-purity HONO source

The first design tested (source 1) produced an output of ~38 ppmv of HONO from a similar amount (>38 ppmv) of HCl (Figure S1A). This HONO mixing ratio is a factor of ~10 larger than the maximum recommended by Stutz et al. (2000) to avoid partial conversion of HONO to NO₂ and NO via [R5R-1](#). Consistently, the source output contained ~4.0 ppmv of NO₂ (Figure S1A) and an analogous amount of NO (Figure S1B). Furthermore, the spectrum contained an absorption feature around 1808 cm⁻¹, reproduced by ~15.5 ppmv of ClNO.

Nitrosyl chloride can be produced by reaction of HONO with gas-phase or wall-adsorbed HCl (Zhang et al., 1996; Wingen et al., 2000) or by reaction of two equivalents of NO₂ with particulate phase Cl⁻ (Weis and Ewing, 1999), though the relatively low mixing ratios of NO₂ in this system makes this reaction less likely.



The FTIR spectrum contained ~50 ppmv more H₂O than the reference spectrum, i.e., more moisture than emitted by the permeation tube with NaNO₂ bypassed. While this is consistent with ClNO production via [R6R5](#), the H₂O data are likely not meaningful since the H₂O mixing ratios changed slowly over time (data not shown), possibly because of slow equilibration with the inner walls of the tubing and multi-pass cell and perhaps also because of water's presence in the FTIR optical path outside the multi-pass cell at a concentration that may have drifted.

Febo et al. (1995) reported lower production of NO_x by mechanically agitating the NaNO₂ using a stir bar (to break up pockets of high [HONO]) and by heating the reaction vessel to 50 °C; these steps were incorporated in the second design (source 2). The FTIR spectrum of its output showed a consumption of > ~42 ppmv HCl and production of ~48 ppmv HONO, ~8.0 ppmv of NO₂, ~6.5 ppmv of ClNO, ~8 ppmv of NO, and ~50 ppmv of H₂O (Figure S2), an improvement over source 1 [in terms of unwanted ClNO production](#) but still inadequate.

A shorter HCl permeation tube was used in the third design, with the expectation that the lower HCl concentrations results in reduced generation of side products via R5 and R6. A sample FTIR spectrum of this source's output is shown in Figure 2. With a freshly prepared HCl permeation tube [and an estimated](#)

relative humidity in the round bottom flask of ~25%, > ~2.5 ppmv HCl were consumed and ~3.0 ppmv of HONO were produced. The mixing ratios of undesired side products (i.e., NO, NO₂, ClNO, and HNO₃) were below their respective FTIR detection limits (Figures 2, S3 and S4).

3.2 Analysis of source output by TD-CRDS

To better constrain the mole fractions of impurities, the source output was analysed by TD-CRDS. Figure 3 shows an example experiment. The TD-CRDS sampled scrubbed air before 21:38 and after 21:58 to determine the ring-down time constants in the absence of absorbers, τ_0 . In the time periods in between, varying amounts of the HONO output were sampled. There was little response other than in the NO_y channel, the only channel sensitive to HONO. Scatter plots of NO₂, NO_x and NO_x + Σ PAN + Σ AN + ClNO + ClNO₂ ("HONO ref") against NO_y (Figure 3, inserts) have slopes of (1.29±0.06)%, (1.54±0.06)% and (2.28±0.04)%, respectively, from which a source purity of >97.3% was deduced.

When concentrations changed, the 90%-10% rise (and fall) times of the TD-CRDS was < 3 s, a time constant identical to changes in NO_x concentration and indicating a "well-behaved" inlet, i.e., the absence of inlet memory effects and fast equilibration with the inner walls of the inlet. The fast response contrasts with the rise (and fall) times of HNO₃ of > 180 s (data not shown). Furthermore, the response in a channel sampling from an inlet heated to 600 °C without added O₃ was the same as the that of the NO₂ channel (data not shown), consistent with the absence of HNO₃ observed by FTIR spectroscopy (Figure S4).

A quartz inlet temperature scan when the TD-CRDS was sampling a constant concentration of HONO is shown in Figure 4. The experimental TD curve was reproduced by a fit to equation (2), which is based on equation (4) by Paul et al. (2009).

$$[\text{Observed}]_{\text{total}} = [\text{NO}_x] + [\text{ClNO}] \left(1 - e^{-A_{\text{ClNO}} \times e^{-\frac{E_{A,\text{ClNO}}}{RT}} \times t_{\text{res}}} \right) + [\text{HONO}] \left(1 - e^{-A_{\text{HONO}} \times e^{-\frac{E_{A,\text{HONO}}}{RT}} \times t_{\text{res}}} \right) \quad (2)$$

Here, A_{HONO} and $E_{A,\text{HONO}}$ are Arrhenius parameters for the TD of HONO taken from Tsang (1991), A_{ClNO} and $E_{A,\text{ClNO}}$ are from Baulch et al. (1981), and t_{res} is the residence time of the gas in the converter at the converter set temperature T . A fit of equation (2) to the observation (superimposed as a black line in Figure 4) using the software package Igor Pro (Wavemetrics) gave $t_{\text{res}} = (77.6 \pm 0.1)$ ms, $[\text{NO}_x] = (0.03 \pm 0.30)$ ppbv, $[\text{ClNO}] = (0.27 \pm 0.34)$ ppbv, and $[\text{HONO}] = (49.97 \pm 0.18)$ ppbv, suggesting a source purity of (99.4±0.9)%.

In our experience, the TD inflection points vary between TD-CRDS channels (and depend on sample flow rates), captured by the t_{res} parameter that is specific to each channel. For the data shown in Figure 4, HONO

fully dissociates at a temperature of ~570 °C, and the inflection point is at ~500 °C. Nitrosyl chloride is predicted to fully dissociate at ~330 °C (inflection point at ~288 °C), such that the temperature of 350 °C chosen for the HONO ref channel (Figure 3) is justified. The blue, dashed line superimposed in Figure 4 shows a simulation of 5 ppbv ClNO and 45 ppbv HONO to demonstrate the relative positions of the TD of ClNO and HONO. We attempted to verify the ClNO TD profile experimentally, but the unoptimized source's ClNO outputs were not sufficiently stable, and those attempts were abandoned. Regardless, it is clear from Figure 4 that the contribution of ClNO to the total source output is negligible.

As can be seen from the insert in Figure 4, HONO contributes a tiny amount to the TD-CRDS signal at 350 °C (~4 pptv in the simulation of 50 ppbv HONO). This contribution may differ between quartz heaters and channels and could have impacted by TD-CRDS using multiple parallel channels presented at the beginning of this section and in Figure 3, leading to an overestimation of the total amount of impurities present in that experiment. The temperature scan data shown in Figure 4, on the other hand, were collected using a single quartz heater and are thus more accurate.

3.3 Source stability

The source output gradually decreased over a time scale of weeks of continuous use, which was rationalized by the visible depletion of the HCl permeation tube. However, the source output remained stable and reproducible on shorter time scales. An example [FTIR](#) time series is shown in Figure 5, which was acquired after 1 month of intermittent use. After a 1.5 hours stabilization period, the source produced a stable output of 1.57 ppmv of HONO (from >1.0 ppmv of HCl) with a precision of ± 35 ppbv.

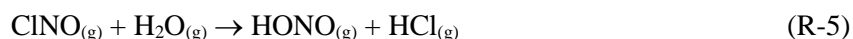
3.4 Source purity and day-to-day reproducibility

The optimized source routinely delivered HONO in high purity (range 96.0% - 98.7%), as long as scrubbed, moisture-containing room air was used as diluent gas (Table 2). When dry cylinder gas was used instead, the source output was stable but contained a larger amount of undesired side products; for example, on July 25, 2019, the output contained 4.6% NO₂ and 8.8% NO + ClNO (Table 2). When used in the field, the performance was similar: with scrubbed air, the HONO purity was 98.5%, whereas with dry cylinder gas, the purity was merely 81.3% (Table 2 and Figure S5).

4. Discussion

250 In this work, a small footprint, portable, stable, rapidly tuneable, and high-purity (>97% on average) HONO source has been described. The source achieves compactness by using a diffusion tube containing 0.35 mL concentrated HCl rather than a 1 L HCl bath. The latter is a considerable hazard since an HCl spill can cause severe burns of skin or permanent eye damage, and hydrochloric acid vapours are toxic to inhale. Hence, smaller volumes are preferred when deploying to the field. The source stabilizes within a time frame
255 of 1.5 h (Figure 5) which is fast compared to, for example, the 10 day stabilization period reported by Roberts et al. (2010). The source output is also rapidly tuneable (Figure 3) which allows expedient generation of calibration plots. These properties make it a useful setup for in-field instrument calibrations.

The combination of FTIR and TD-CRDS provided a unique and powerful tool set to analyse the purity of the source's output. This work has shown that HONO sources based on R4 can generate NO, NO₂ and ClNO
260 as by-products via R-1 and R5. Febo et al. (1995) and Roberts et al. (2010) have presented HONO sources based on R4 and demonstrated stoichiometric conversion of HCl to HONO. The results in this work suggest that devices relying solely on loss of HCl for calibration of HONO output should be used with caution as co-production of NO, NO₂, and ClNO is a real possibility. In our setup, formation of ClNO could be avoided by keeping the HCl concentration low (i.e., < 4 ppmv), which puts an important constraint on sources
265 generating HONO by R4. With this in mind, it is instructive to scrutinize the earlier work in regard to side product formation. It has generally been noted that the presence of moisture is needed to ensure a high HONO output. We suggest that water is needed in part because ClNO is prone to hydrolysis (Karlsson and Ljungstrom, 1996) such that the product distribution is shifted towards HONO.



270 Roberts et al. (2010) observed no ClNO production, likely because they kept their concentrations relatively low (<900 ppbv) and humidified their gas streams, though they appear to be just below the threshold of possible ClNO production. Perez et al. (2007), on the other hand, observed ClNO but did not disclose details as to how they operated their source, i.e., if they worked under trace conditions and with humidified gases, though it is likely that either or both conditions were not met. Schiller et al. (2001) bubbled N₂ through
275 liquid HCl and would have certainly achieved high enough HCl concentration to generate ClNO with their setup. However, they reported achieving higher HONO output in the presence of moisture. Hydrolysis of ClNO (R-5) may be a (partial) explanation why the presence of H₂O improved the output of their HONO.

The present HONO source has a large dynamic range (0 - 1 ppmv), well above the HONO concentrations found in ambient air. For field calibrations in future, it may be desirable to construct a permeation tube with
280 lower HCl output, for example by lowering the HCl concentration in the permeation tube.

Data availability

The data used in this study are available from the corresponding author upon request (hosthoff@ucalgary.ca).

Author contributions

285 NJG and HDO designed the experiments and carried them out.

Competing interests

The authors declare that they have no conflict of interest.

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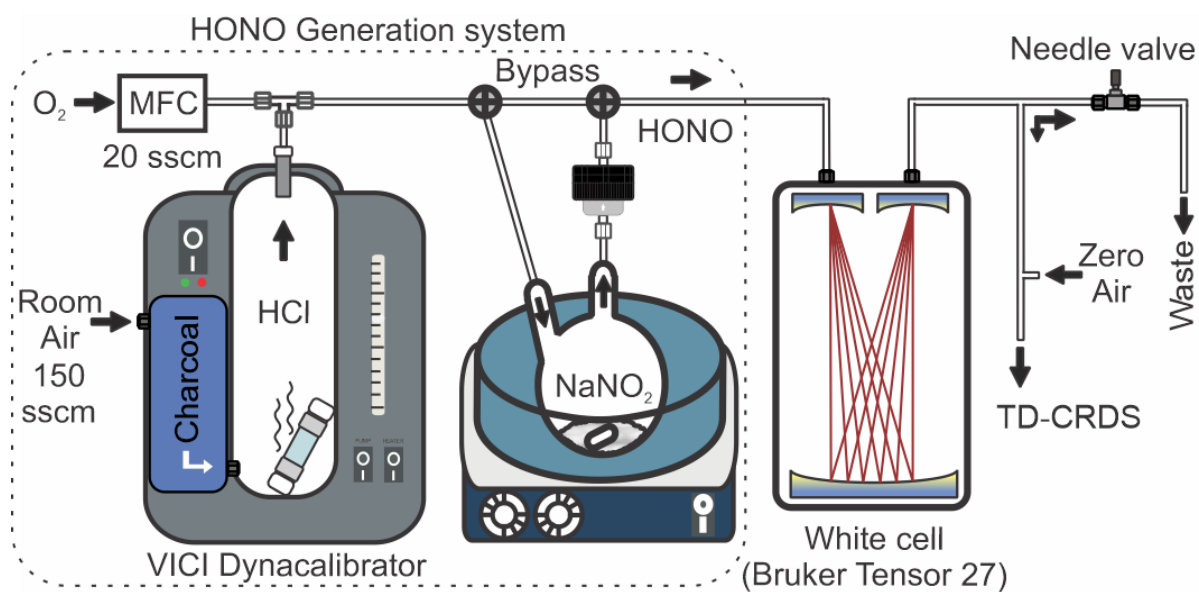
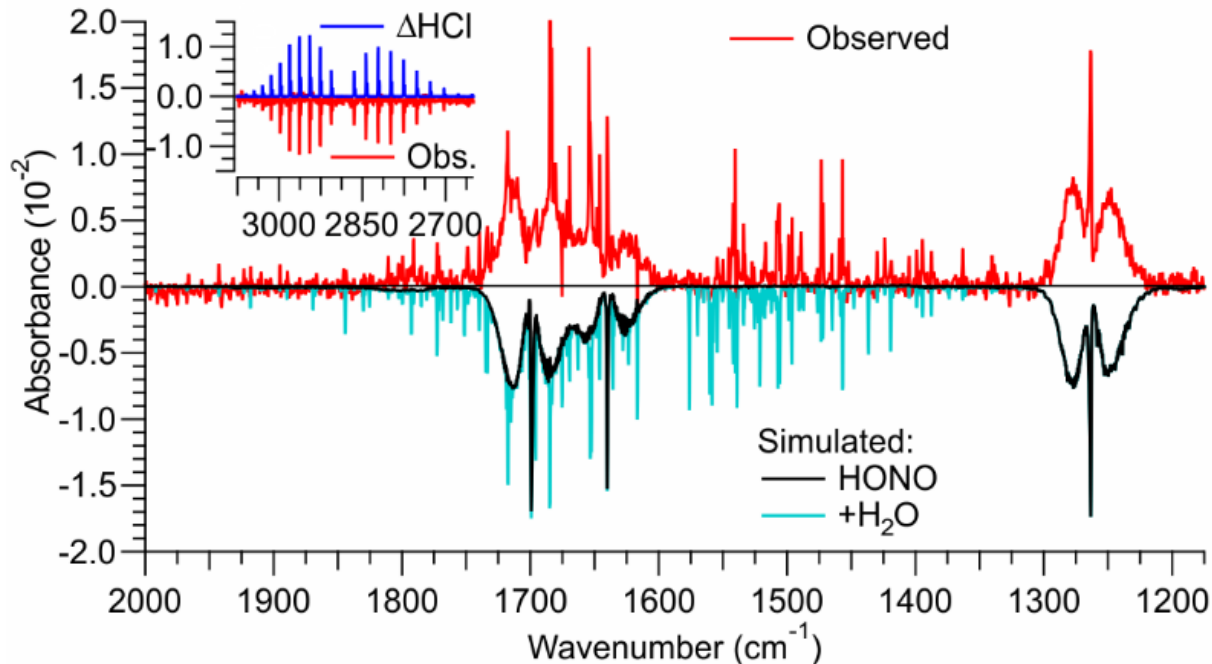


Figure 1. Schematics of the experimental setup. The high-purity HONO generation system is shown on the left (dotted border). MFC = mass flow controller. sscm = standard cubic centimetre per minute.



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Figure 2. Infrared spectrum (shown in red colour) of a gas stream containing HONO generated by reaction of $\text{HCl}_{(\text{g})}$ with $\text{NaNO}_{2(\text{s})}$ after the source was optimized. The reference spectrum was collected when NaNO_2 was bypassed, i.e., contained HCl. Literature spectra (Sharpe et al., 2004) were multiplied by the optical path length of 6.4 m and mixing ratios of the identified trace gases until they reproduced the observed spectrum. The optimized source delivered 3.0 ppmv of HONO from >2.5 ppmv of HCl. The spectrum also contained ~ 4.0 ppmv of H_2O . The HCl and H_2O concentrations are underestimates of their true concentrations since their absorption lines are narrower than the resolution of the FTIR of 0.5 cm^{-1} .

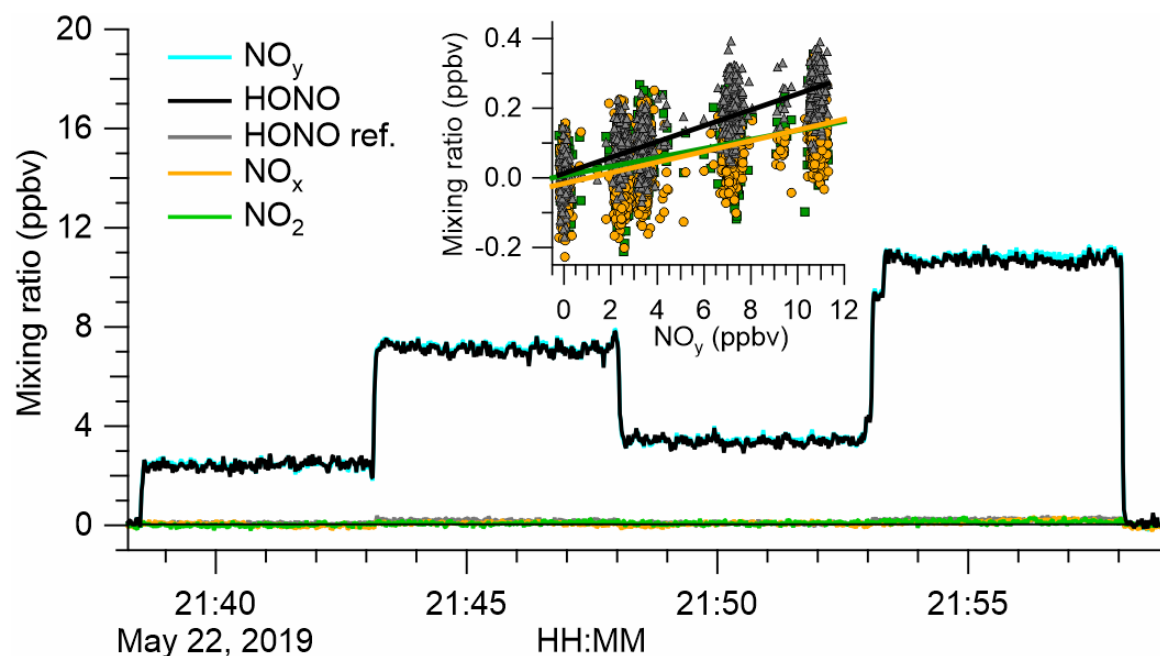


Figure 3. Analysis of the HONO source output by TD-CRDS. The TD-CRDS sampled scrubbed "zero" air before 21:38 and after 21:58. The HONO source output was varied by incrementally opening (or closing) the bypass valve. The HONO mixing ratio was calculated by subtracting the response of the "HONO reference" from the NO_y channel. The insert shows scatter plots of NO₂, NO_x and "HONO ref" against NO_y. Slopes of $(1.29 \pm 0.06)\%$ for NO₂ (points shown in green), $(1.54 \pm 0.06)\%$ for NO_x (data points shown in orange) and $(2.28 \pm 0.04)\%$ for "HONO ref" (points shown in grey) were determined, respectively.

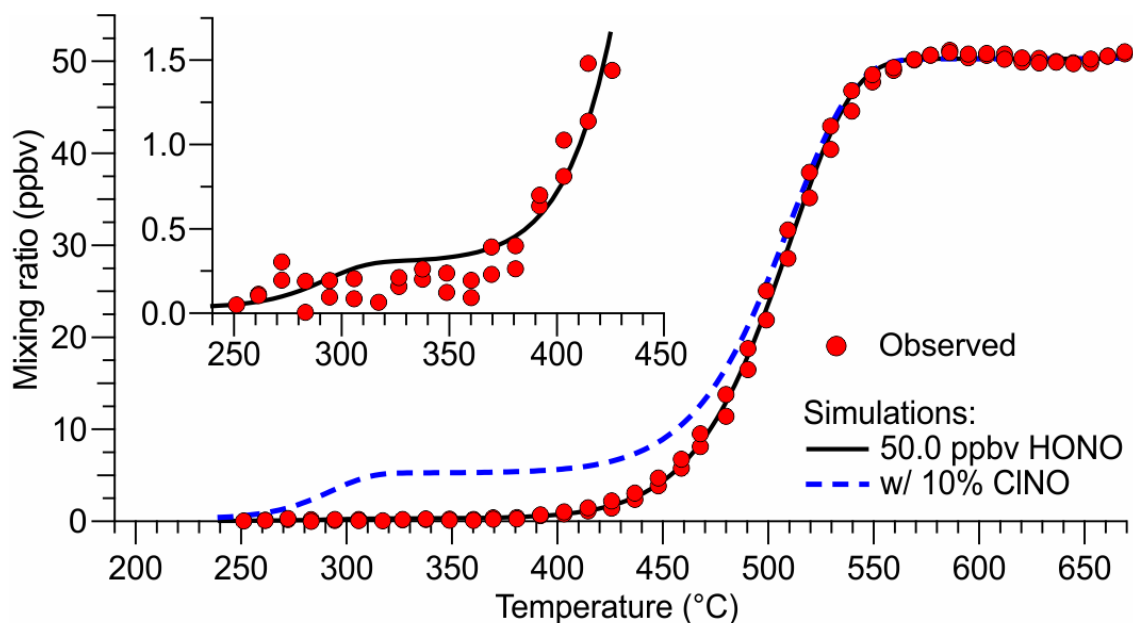


Figure 4. Inlet temperature scan when the TD-CRDS was sampling a constant concentration of HONO (data points shown in red). The TD profile was reproduced by a fit to equation (2) with mixing ratios of 400 (49.97 ± 0.18) ppbv, (0.27 ± 0.34) ppbv and (0.03 ± 0.30) ppbv for HONO, CINO and NO_x , respectively (black line). The blue dashed line shows the predicted TD curve of a hypothetical mixture containing 5.0 ppbv CINO and 45.0 ppbv HONO. The insert shows a close-up of the temperature region in which TD of CINO occurs.

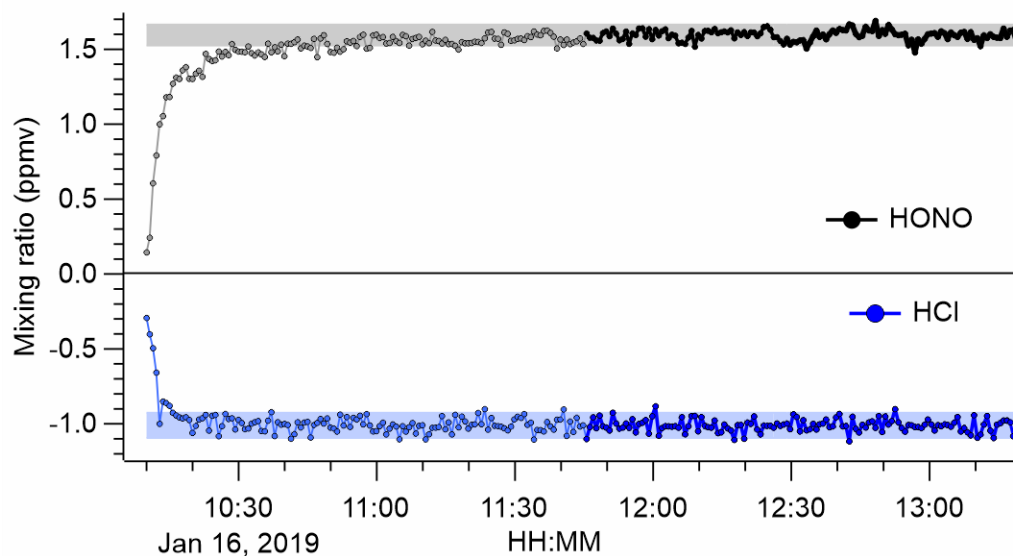


Figure 5. Time series of HONO and HCl mixing ratios derived from FTIR analysis of the undiluted HONO source output. The NaNO_2 was placed in line at 10:10. Data acquired during the initial period are shown in light grey colour. The shaded areas represent the average $\pm 2\sigma$ after the source output ~~had~~was judged to have stabilized (after 11:45; data points shown in black colour). The 1σ precision of the HONO data was ± 35 ppbv and that of the HCl data was ± 42 ppbv. The HCl mixing ratios are an underestimate because the widths of their absorption lines are less than the FTIR's resolution of 0.5 cm^{-1} .

Table 1. Selected HONO generation systems described in the literature. n/d = not disclosed

| Reference | Method | Analytical method | Result | Notes |
|--|--------|---|---|---|
| (King and Moule, 1962; Stockwell and Calvert, 1978) | (R1) | UV absorption | mixture of NO, NO ₂ and HONO | Equilibration of (R1) |
| (Cox, 1974) | (R1) | NO and "total NO _x " CL; HONO scrubbed w/ NaOH(0.1N) | mixture of NO, NO ₂ and HONO | Equilibration of (R1) |
| (Cox and Derwent, 1976) | (R2) | NO and "total NO _x " CL; HONO scrubbed w/ NaOH(0.1N) | mixture of NO, NO ₂ and ~50% HONO | Volatilization of "nitrous fumes" from solution of sulfuric acid and sodium nitrite |
| (Braman and De la Cantera, 1986) | (R3) | NO/NO _y CL analyser (heated Au tube) | mixture of NO, NO ₂ and 50% - 90% HONO | Sublimation of oxalic acid on sodium nitrite |
| (Taira and Kanda, 1990) | (R2) | NO/NO _y CL analyser (carbon converter); HONO collected on Na ₂ CO ₃ filters and analysed by IC | mixture containing 2% - 3% NO and 4% - 6% NO ₂ | Volatilization of HONO from dynamically mixed sodium nitrite and sulfuric acid solution |
| (Febo et al., 1995; Stutz et al., 2000) | (R4) | DOAS (NO ₂ , HONO); NO CL | >99.5% purity | 1 L bath, 2 m reverse diffusion tube, heated stirring reactor |
| (Brust et al., 2000) | (R4) | FTIR (NO ₂ , HONO), IC (HONO) | up to 22ppm; <2% NO ₂ | 20 cm reverse diffusion, heated reactor |
| (Schiller et al., 2001) | (R4) | TLDAS (HONO), FTIR (HONO) IC of KOH solutions | n/d | bubbled N ₂ through 0.5 M HCl followed by reaction with solid sodium nitrite |
| (Perez et al., 2007) | (R4) | TD-CL, TD-LIF | observed ClNO | same setup as Febo et al. (1995) |

| | | | | |
|------------------------|------|---|---|--|
| (Perez et al., 2007) | (R2) | TD-CL, TD-LIF | >95% | flowed H ₂ SO ₄ aerosol over NaNO ₂ sandwiched between paper filters |
| (Roberts et al., 2010) | (R4) | IBBCEAS, NO _y -CL, CIMS | >95% | HCl gas cylinder (10 ppmv) and NaNO ₂ reactor tube |
| (Ren et al., 2010) | (R4) | LOPAP, NO _y -CL, CIMS | n/d | Reverse diffusion tube, high [HCl] (9-12 M), tubing thickness not described |
| (Reed et al., 2016) | (R2) | Differential photolysis NO-CL; NO ₂ CAPS; FTIR | HONO (70.4%; < 50 ppbv); NO (15%); NO ₂ (12.8%); HNO ₃ (1.3%) | Perm tube filled with 37% HCl placed in oven with NaNO ₂ salt; flushed @ 1.5 slpm; <50 ppbv |

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Table 2. Summary of TD-CRDS analyses of the HONO source output. The RH of the diluent gas was in the range of 15% to 35%, except for experiments conducted with cylinder gases which are shown below the dashed line. The range of mixing ratios stated is for output after dilution. The stated errors are from regression analyses of plots of NO₂, NO_x or NO_x+ClNO vs. NO_y and are at the 1σ level. n/d = not determined.

| <u>Date</u> | <u>Setting</u> | <u>Diluent gas</u> | <u>Range (ppbv)</u> | <u>HONO (%)</u> | <u>NO₂ (%)</u> | <u>NO_x (%)</u> | <u>NO_x+ClNO (%)</u> |
|---------------------|----------------|-------------------------------|-------------------------|---------------------|-------------------------------|-------------------------------|------------------------------------|
| <u>Jan 16, 2019</u> | <u>Lab</u> | <u>Scrubbed air</u> | <u>0 - 140</u> | <u>*97.2±0.1</u> | <u>n/d</u> | <u>2.80±0.07</u> | <u>n/d</u> |
| <u>May 3, 2019</u> | <u>Lab</u> | <u>Scrubbed air</u> | <u>0 - 7</u> | <u>96.0±0.2</u> | <u>1.9±0.2</u> | <u>4.0±0.2</u> | <u>(4.0±0.2)**</u> |
| <u>May 6, 2019</u> | <u>Lab</u> | <u>Scrubbed air</u> | <u>0 - 93</u> | <u>96.4±0.4</u> | <u>1.9±0.4</u> | <u>3.6±0.4</u> | <u>(3.6±0.4)**</u> |
| <u>May 10, 2019</u> | <u>Lab</u> | <u>Scrubbed air</u> | <u>0 - 21</u> | <u>*97.6±0.1</u> | <u>1.10±0.03</u> | <u>2.40±0.05</u> | <u>n/d</u> |
| <u>May 17, 2019</u> | <u>Lab</u> | <u>Scrubbed air</u> | <u>0 - 38</u> | <u>98.7±0.1</u> | <u>0.01±0.02</u> | <u>0.92±0.01</u> | <u>1.25±0.01</u> |
| <u>May 22, 2019</u> | <u>Lab</u> | <u>Scrubbed air</u> | <u>0 - 14</u> | <u>97.7±0.1</u> | <u>1.29±0.06</u> | <u>1.54±0.06</u> | <u>2.28±0.04</u> |
| <u>Jan 17, 2020</u> | <u>Field</u> | <u>Scrubbed air</u> | <u>0 - 80</u> | <u>98.5±0.1</u> | <u>1.00±0.05</u> | <u>1.50±0.08</u> | <u>(1.50±0.08)**</u> |
| <u>Mar 19, 2020</u> | <u>Lab</u> | <u>Scrubbed air</u> | <u>0 - 320</u> | <u>*98.72±0.01</u> | <u>n/d</u> | <u>1.28±0.01</u> | <u>n/d</u> |
| <u>Jul 25, 2019</u> | <u>Lab</u> | <u>Cylinder O₂</u> | <u>0 - 95</u> | <u>86.60±0.02</u> | <u>4.60±0.03</u> | <u>n/d</u> | <u>13.4±0.02</u> |
| <u>Jan 30, 2020</u> | <u>Field</u> | <u>Cylinder N₂</u> | <u>0 - 50</u> | <u>81.3±0.1</u> | <u>5.90±0.09</u> | <u>n/d</u> | <u>18.7±0.1</u> |

* Upper limit. ** The TD-CRDS quartz inlet temperature was ramped between 300 °C and 600 °C; the TD-CRDS mixing ratios at an inlet temperature of 300 °C matched the NO_x mixing ratios observed with the room temperature inlet, indicating the absence of ClNO.