



Ch3MS-RF: A Random Forest Model for Chemical Characterization and Improved Quantification of Unidentified Atmospheric Organics 5 Detected by Chromatography-Mass Spectrometry Techniques

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Abstract. The chemical composition of ambient organic aerosols plays a critical role in driving their climate and health relevant properties and holds important clues to the sources and formation mechanisms of secondary aerosol material. In most ambient atmospheric environments, this composition remains incompletely characterized, with the number of identifiable species consistently outnumbered by those that have no mass spectral matches in the literature or NIST/NIH/EPA mass spectral databases, making them nearly impossible to definitively identify. This creates significant challenges in utilizing the full analytical capabilities of techniques which separate and generate spectra for complex environmental samples. In this work, we develop the use of machine learning techniques to quantify and characterize novel, or unidentifiable, organic material. This work introduces Ch3MS-RF (Chemical Characterization by Chromatography-Mass Spec Random Forest Modelling), an open-source R-based software tool for efficient machine-learning enabled characterization of compounds separated in chromatography-mass spec applications but not identifiable by comparison to mass spectral databases. A random forest model is trained and tested on a known 130 component representative external standard to predict the response factors of novel environmental organics based on position in volatility-polarity space and mass spectrum, enabling reproducible, efficient, and optimized quantification of novel environmental species. Quantification accuracy on a reserved 20% test set randomly split from the external standard compound list indicate that random forest modelling significantly outperforms the commonly used methods in both precision and accuracy, with a median response factor % error of -2% for modelled response factors compared to >15% for typically used proxy assignment-based methods. Chemical properties modelling, evaluated on the same reserved 20% test set as well as an extrapolation set of species identified in ambient organic aerosol samples collected in the amazon rainforest, also demonstrates robust performance. Extrapolation set property prediction mean average errors for carbon number, oxygen to carbon ratio (O:C), average carbon oxidation state (\overline{OS}_c), and vapor pressure are 1.8, 0.15, 0.25, and 1.0 (log(atm)), respectively. Extrapolation set Out-of-Sample R^2 for all properties modelled are above 0.75, with the exception of vapor pressure. While predictive performance for vapor pressure is less robust compared to the other chemical properties modelled, random forest-based modelling was significantly more accurate than other commonly used methods of vapor pressure prediction, decreasing mean average vapor pressure prediction error to 0.24 (log(atm)) from 0.55 (log(atm)) (chromatography-based vapor pressure prediction) and 1.2 (log(atm)) (chemical formula-based vapor pressure prediction). The random forest model significantly advances untargeted analysis of the full scope of chemical speciation yielded by GCxGC-MS techniques and can be applied to GC-MS as well. It enables accurate estimation of key chemical properties commonly utilized in the atmospheric chemistry community, which may be used to more efficiently identify important tracers for further individual analysis and to characterize compound populations uniquely formed under specific ambient conditions.

55 1 Introduction

Organic aerosols play a critical role in global radiative forcing and regional aerosol-attributable public health concerns, making up a significant (20-90%) fraction of fine particulate matter around the globe (Jimenez et al., 2009). This organic material is highly complex in terms of chemical composition and constantly changing; Goldstein and



Galbally, 2007 estimates the number of gas and aerosol-phase atmospheric organic constituents to lie in the millions, while Ditto et al., 2018 reports molecular level variability of 60-80% between consecutive samples collected at fixed sites for samples comprised of high thousands of resolvable species. While there has been significant progress towards achieving mass closure of atmospheric reactive carbon using an ensemble of both bulk and speciated measurement techniques over the past two decades, speciated and isomer identified mass closure remains challenging (Heald et al., 2010; Hunter et al., 2017; Isaacman-Vanwertz et al., 2018). A comprehensive review of the challenges and utility of different levels of molecular identification, Nozière et al., 2015, compares the utility of many types of incomplete identification of atmospheric organic compounds, but defines that “An organic compound is fully identified only if its molecular structure is entirely known, including its isomeric and spatial (stereo) configuration.” Important chemical information can be gleaned from formula-based identifications and bulk characterization, but isomer-specific identifications provide critical atmospheric chemistry-relevant insights. As described in Isaacman-Vanwertz and Aumont, 2021, different isomers of the same chemical formula vary over orders of magnitude in volatility and Henry’s constant, and by a factor of 2 in reactivity with the hydroxyl radical, all critical properties for characterization of aerosol formation and properties. Isomer specific identifications also play a crucial role in elucidation of important chemical reaction mechanisms.

Gas chromatography coupled with electron ionization mass spectrometry (GC-MS) is a commonly utilized technique for isomer specific speciation of atmospheric constituents. Observed ambient species may be matched to authentic standards or mass spectral database entries by both retention index (chromatographic elution time relative to that of a series of alkanes) and mass spectrum. Two dimensional gas chromatography (GCxGC-MS), a methodologically similar technique which achieves advanced separation by passing compounds through multiple GC columns configured for different chemical properties, increases the scope of isomer-specific identification by separating species that would coelute in single dimension GC-MS applications (Goldstein et al., 2008; Worton et al., 2011, 2017). However, a significant challenge of fully utilizing the data from these techniques is the novelty and diversity of the atmospheric constituents; most observed organic species have never been synthesized and are not in any mass spectral library and are therefore not directly identifiable from GC-MS or GCxGC-MS techniques. Although the size of mass spectral libraries are rapidly increasing, with ~30,000 new compounds added to the NIST/EPA/NIH mass spectral database between the 2011 and 2014 versions (bringing the number of compounds catalogued in NIST14 EI library to ~250,000), the numbers of identifiable constituents in typical atmospheric samples remain low (Vinaixa et al., 2016). As described in Hamilton et al., 2004, in an urban aerosol sample analyzed by GCxGC-MS, of >10,000 unique observed species, fewer than 2% were identifiable from authentic standard or mass spectral matching. Low numbers of matched relative to novel ambient species persist; Worton et al., 2017 finds that fewer than 35% of ~500 compounds isolated from aerosol samples collected at a forested site match mass spectral database entries, while this work (as later described) finds that fewer than 10% of ~1500 aerosol phase organic species can be matched to published spectra. As described in Worton et al., 2017, species that cannot be identified are often not included in GC-MS and GCxGC-MS-based analyses, meaning that the majority of



95 acquired data is not fully utilized. Note that in accordance with the definition of complete molecular identification
previously quoted from Nozière et al., 2015, “unidentified” compounds are from here on defined as any species that is
not identifiable by comparison (on the basis of retention index and mass spectrum) to either authentic standards or mass
spectral database entries of positively identified species. Pairing GC-EI-MS systems with complementary measurements
such as chemical ionization (described in Bi et al., 2021) or switching to softer electron ionization techniques (specifically
through employing 14 eV vacuum ultraviolet rather than traditional 70 eV EI, intended to preserve sufficient precursor
ion mass for formula identification, as described in Worton et al., 2017) can enable more separated but unidentified
100 compounds to be characterized by formula identification, even where isomer-specific identification remains elusive. That
said, these instrumental configurations are rare, and fragmentation under 14 eV is still sufficiently significant to leave
many species’ formulae not identifiable and therefore still uncharacterized (Worton et al., 2017). Recent efforts to
embrace a larger fraction of the full complexity of chemical information yielded by highly speciated organic aerosol
measurements (on the scale of low to mid 100’s of compounds) have categorized unidentified species by likely source
105 groups and chemical families through time series correlations with known tracer species (Zhang et al., 2018) or by manual
group assignments by individual researcher judgements based on mass spectral features (Liang et al., 2021). These
methods are difficult to standardize and reproduce and become prohibitively inefficient when pushing towards the full
chemical complexity of speciated observations produced from typical atmospheric samples, which extend into the low
to mid thousands of species.

110 Quantification of unidentified compounds faces similar challenges. Where possible, compounds in GC and GCxGC-
MS are directly quantified by calibration curves of authentic standards, but direct quantifications are limited by standard
expense and availability, even for species that can be positively identified. Compounds that cannot be directly quantified,
both in GCxGC-MS and in GC-MS, are most commonly quantified by assigning quantification factors from compounds
resolved closely in chromatographic space, compounds that are identified as sharing chemical structures, or some
115 interpolation of multiple nearby proxies (Hatch et al., 2015; Jen et al., 2019; Liang et al., 2021; Zhang et al., 2018). The
errors associated with these assignments/choices are usually estimated from the range of quantification factors of close
or chemically similar species and are assumed to be high (up to a factor of 2 depending on degree of certainty in assigning
chemical class as described in Jen et al., 2019 and Liang et al., 2021). To our knowledge, this work presents the first
quantitative error analysis of these techniques based on applying proxy quantification techniques to compounds with
120 known quantification factors.

Current manual characterization and quantification proxy-assignments are essentially an exercise of pattern
recognition, as researchers use experience in analysing spectra and position in chromatographic space to categorize or
otherwise characterize unidentifiable species. Given the scale of the novel compound characterization challenge (on the
order of hundreds to thousands of species for a given sampling location using current methods), transitioning to automated
125 characterization methods will be necessary to keep up with data acquisition, and will offer co-benefits in increased
reproducibility and reduced susceptibility to researcher biases. Decision tree-based machine learning methods including



130 gradient boosting and random forests have demonstrated robust performance in pattern recognition-based regression applications including nonlinear features across a wide range of fields (Bentéjac et al., 2021; Rokach, 2016). Random forests, a decision tree-based method which generates predictions based on a combination of diverse trees generated by randomized feature selection and resampling on a training data set (Breiman, 2001), are particularly suited to this application and intended audience. They have demonstrated robust performance across a range of applications, including predictions of chemical properties (Whitmore et al., 2016) and do not require extensive hyperparameter tuning to achieve high performance (Bentéjac et al., 2021). In this work, we develop machine learning models, specifically based on the random forests methodology, that use chromatographic and mass spectral feature inputs to predict a diverse suite of
135 chemical properties, including quantification factor in a TD-GCxGC-MS system, oxygen to carbon ratio (O:C), carbon number, average carbon oxidation state (\overline{OS}_c), and vapor pressure. Coinciding with this manuscript, we have released a repository template including an Rmarkdown notebook (<https://github.com/ebarnesey/Ch3MS-RF>) that enables users with general atmospheric chemistry background, who do not necessarily have special expertise in machine learning data science applications, to tailor our analysis for their specific use cases. As such, robust performance evaluation and ease
140 of applicability to a range of potential use cases are emphasized over extensive application-specific hyperparameter tuning.

In summary, this work aims to provide the GC-MS and GCxGC-MS atmospheric chemistry community with tools to achieve the following objectives:

- 1) Enable accurate chemical characterization of organic constituents separated in gas chromatographic space but not necessarily published (in mass spectral databases)
- 2) Improve the quantification accuracy for species that cannot be directly calibrated using authentic standards

2 Instrumentation and Data

2.1 Calibration Curves Using an External Standard Mixture of Authentic Standards

150 A custom calibration standard mixture (referred to hereafter as “external standard”) was created containing ~130 unique authentic standards selected for maximal coverage of the compounds and compound classes typically observed in atmospheric regions with significant biogenic emissions, as well as influences from anthropogenic activities and biomass burning. The selection of these standard species was informed by previous work targeting similar sample types using the same instrumentation (Worton et al., 2011; Yee et al., 2018; Zhang et al., 2018), and covers species including sugars, PAH’s, and both monoterpene and isoprene oxidation products. In addition to commercially available external standards, 6 sesquiterpene
155 oxidation products were custom synthesized by collaborators (as described in Bé et al., 2019) for expanded coverage of potentially important chemical tracers. The full list of standard components can be found in Table A1, and the standard property distribution in volatility-polarity space is illustrated in Figure 2. The standard was prepared from pure components immediately prior to sample analysis in 1:1 methanol:chloroform solution, replicating the methodology utilized in Zhang et al., 2018.



Standards were introduced to the instrument by injecting onto tissuquartz filter material to maximize consistency between filter
160 samples (organic aerosol also collected on tissuquartz filters) and calibration runs. At 5 points throughout sample analysis, 6-
point calibration curves (5 loaded points and a zero point) were performed to determine the “quantification factors” (internal
standard normalized signal/ng compound) of each external standard species. The internal standard, described in detail in
section 2.3.1, is a solution of ~30 deuterated organics applied identically to all sample and calibration analysis runs to enable
correction for instrument condition and matrix effects. For efficiency, outlier calibration points (significantly deviating from
165 the slope of other points in the quantification factor, which are often caused by coelution with a contaminant) were removed.
A minimum of 3 calibration points above the zero point was maintained to ensure robust quantification factors.

2.2 GoAmazon Field Data

The ambient extrapolation data utilized in this work originates from the Green Ocean Amazon (GoAmazon) field
campaign which was conducted in central Amazonia in 2014. This campaign and the collection of ambient filters for offline
170 analysis are described in detail in Martin et al., 2016, 2017 and Yee et al., 2018. Briefly, the campaign was conducted at a semi
remote site occasionally downwind of the city of Manaus and periodically impacted by smoke from biomass burning. The
campaign spanned two intensive operating periods, one during the Amazonian wet season (February through March) and one
during the dry season (August through early October). Submicron aerosol samples were collected on tissuquartz filters
(Pallflex), stored in pre-baked foil, double contained in sealed mylar bags, and frozen prior to analysis. The samples were
175 analysed by TD-GCxGC-EI-ToF-MS, as described below.

2.3 Instrumentation: TD-GCxGC-EI-ToF-MS

Both external standard species (during calibration runs) and GoAmazon filter samples were analysed by thermal
desorption two-dimensional gas chromatography coupled with electron ionization time-of-flight mass spectrometry (TD-
GCxGC-EI-ToF-MS, hereafter abbreviated GCxGC-MS). This instrumentation is described in detail in Goldstein et al., 2008
180 and Worton et al., 2011, and instrument specifics including sub-component models, column materials, and temperature settings
are described in Franklin et al., 2021. For ambient filter samples, 0.4 cm² aliquots of filter material are directly introduced into
the instrument. Standards are stored in solution and introduced by injection onto pre-baked quartz filter material. An internal
standard (described in section 2.3.1) is applied on top of the sample or external standard filter aliquots immediately prior to
analysis. Briefly, the instrument functions as follows: a thermal desorption oven heats filter material, causing analytes and
185 standards to evaporate into a flow of helium. The desorbed components are focused on a cooled inlet system (Gerstel CIS),
which at the end of the thermal desorption cycle is rapidly heated to simultaneously release all organic species onto the head
of the first column. Compounds are separated by both volatility and polarity by two gas chromatography columns in sequence,
with the transition of compounds from the first to the second column modulated by a cryogenic focus and rapid thermal release
system. Separated analytes are ionized by 70 eV electron ionization (EI) and detected by HR-ToF-MS (TOFWERK, EI-
190 HTOF), with a resolving power of 4000 acquired at 100 Hz. While the mass spectra produced by this technique are high



resolution, these high resolution mass spectra are converted to unit mass resolution spectra to increase the applicability of this technique to unit mass resolution techniques. The vertical (polar) axis of separation is extremely short relative to the horizontal (volatility) axis separation with a vertical stride length of 2.3 seconds compared to a retention time of ~ 1 hr for low volatility organics. As a result, GCxGC-MS deuterated alkane normalized retention indices are directly comparable to retention indices (or, with a linear conversion to non-deuterated retention indices, Kovats indices) in single dimension GC-MS applications. This instrument's volatility range spans approximately C₁₃-C₃₆ *n*-alkane volatility equivalents, covering the atmospherically important transition regime between IVOC (intermediate volatility organic carbon) and LVOC (low volatility organic carbon) species.

During the thermal desorption process, the carrier flow of helium is enriched with the derivatization agent MSTFA (*n*-methyl-*n*-trimethylsilyl-trifluoro-acetamide). This silylating reagent replaces the active hydrogen of polar OH groups with a trimethylsilyl group, -Si(CH₃)₃, a process which significantly enhances the recovery of polar organics. This approach is critical to increase the scope and degree of oxygenation of species recovered by thermal desorption-gas chromatography techniques (Isaacman et al., 2014). However, it poses some challenges for data interpretation for diverse, complex, and novel chemical mixtures, because in the case of many polar species, the compound that is separated and detected by the GCxGC-MS instrumentation has been chemically altered from the species that was collected. This can create challenges in compound identification, as not all species have published derivatized spectra, as well as challenges for mapping chemical properties onto the GCxGC-MS space, as the volatility-polarity distributions of derivatized compounds do not directly reflect their underivatized properties.

2.3.1 Internal Standard Normalization

Both filter samples and external standard impregnated filters (for calibration curves) were doped with a custom 23 component deuterated internal standard covering the full range of volatility sensitivity and a broad variety of functional group types immediately prior to analysis. The internal standard enables normalization for matrix effects, configuration of retention indices relative to the elution times of a deuterated alkane series, and normalization for instrument condition drift for improved consistency and quantification accuracy throughout intensive instrument use. In prior methods, the selection of internal standard involved either 1) assigning each analyte an internal standard nearest in chromatographic space (by retention times) or 2) manual assignment of analytes to their most chemically similar internal standards regardless of proximity in GCxGC space. Analyte signal would then be normalized (divided) by the signal of the selected internal standard obtained during the same chromatographic run. In a new approach employed in this work, in order to maximize the reliability and consistency of normalization across a large number of samples and complex sample media, internal standard signals were each normalized by their own mean signals (throughout the entire analysis period) to yield an indicator of self-normalized instrument sensitivity. Analyte signal was then normalized by the mean self-normalized responses of the three closest internal standard species. This approach has multiple benefits. First, the responses of sample or external standard compounds are not artificially deflated or inflated due to their proximity to internal standard compounds that have higher or lower sensitivities based on their functional



groups and derivatization. Second, this approach enables inclusion and utilization of incomplete data; in previous approaches,
225 if an internal standard cannot be recovered in every sample it cannot be used for normalization, as this would create
inconsistencies for the species that are otherwise assigned to that compound. Compounds at the very high and very low ends
of the volatility space are chemically important but detectable at baseline low levels that can drop below limits of detection
during periods of low sensitivity. Having to discard these species due to a few instances of missing corresponding internal
standard data causes losses of valuable information. Finally, this approach decreases analysis sensitivity to any errors and noise
230 in internal standard identification or isolation, as erroneously high or low individual internal standard responses are moderated
by averaging with the other nearby internal standard species. Volatility-based sensitivity corrections, which can be achieved
by raw internal standard normalization, were achieved in this work through normalization by an external standard-determined
response curve, as described in “Featurization and Target Selection for Quantification Modelling.”

3 Data Preparation and Featurization

235 The analytical pipeline for data preparation through performance evaluation of this random forest modelling work is
illustrated in Figure 1. The processes and decision making around featurization, feature selection, and target selection for both
chemical properties modelling and quantification modelling, as well as the curation of the training, test, and extrapolation data
sets, is described below.

3.1 Featurization, Feature Selection, and Target Selection

240 As the aim of this work is to develop methods that can be applied to novel species not included in mass spectral
databases, features utilized in this analysis rely solely upon the information readily available for unidentifiable species. Given
the size and complexity of the intended use data suites, features must also be automatically generatable from the instrument
data output and not rely upon any visual or manual categorization by researchers. In order to make these models more broadly
useful to the atmospheric community, less common features produced by the GCxGC-MS instrumental setup (e.g. second
245 dimension retention time and high resolution spectra) are not utilized for chemical properties modelling in order to increase
the method’s applicability to single dimension GC-MS systems and instruments with lower resolution mass spectra.

3.1.1 Mass Spectral Featurization

The only chemical information directly produced by GCxGC-MS for unidentified organic species are their locations
in GCxGC volatility-polarity space and mass spectra. These sources of information are therefore exclusively utilized in
250 creating and selecting the features for chemical properties modelling. The retention index of each compound was directly
utilized as a feature, but the mass spectra require interpretation in order to be used.

The unit mass resolution spectra utilized in this analysis include each charged fragment represented by its measured
mass to charge ratio (m/z) and a relative signal score out of 1000 (normalized by the most abundant fragment’s peak signal).



255 EI is a high energy or “hard” ionization technique which typically leaves only a small fraction of molecular ions intact and
creates positively charged ion fragments that are almost all singly charged, with any multiply charged ions at extremely low
abundance. This means that the molecular formulae cannot generally be directly determined from the mass spectrum, even
when the spectra are high resolution, and measured ions can be assumed to have a single charge. That said, the m/z of charged
fragment ions yield useful information into chemical characteristics and functional groupings that can provide critical chemical
information; for example, a peak at $m/z=73$ corresponds to a fragment of $\text{Si}(\text{CH}_3)_3^+$, a derivatization fragment which indicates
260 that the ionized compound contained an OH group which was derivatized (see section 2.3). The mass differences between
charged peaks also represent important pieces of information, as they can indicate losses of uncharged molecular fragments
that similarly point to the structure and characteristics of the original compound. It is important to note that not all neutral mass
differences between charged peaks can be interpreted as direct neutral losses as not all high m/z charged fragments directly
fragment onto lower m/z charged fragments in a manner that can be directly interpreted from neutrally charged fragment losses.
265 However, frequently occurring neutral differences may still hold value in reflecting a common coordination of neutral loss
processes.

The greatest chemical information lies in features that exist in an intermediate range of occurrence frequency in the
data set. A feature which appears in all the training species does not provide any useful information in predicting properties of
the test species. Neither does a feature which is totally unique to a single species, as it does not provide any information on
270 patterns which can be used to adjust prediction of properties for other species. This logic can be applied to mass spectral
featurization; while it would be possible to convert every m/z to a feature and so input the entire raw mass spectrum of each
compound as a series of features for the random forest model, this approach would be inefficient, open to error introduced by
noise, and miss the important information provided by neutral mass differences between charged fragments.

Multiple approaches for mass spectral featurization were tested to optimize the number of features and representation
275 of features. Given the final choice in model structure (random forest, as described in section 4), inclusion of covarying features
or more features than necessary did not introduce significant sources of error. Target-specific feature restriction based on
importance is discussed in section 4. The final mass spectral featurization method selected for this analysis, a simplified
adaptation of methodology described in Eghbaldar et al., 1998, was as follows: the top 5 charged fragments (mass spectral
peaks) from each training set mass spectrum are selected. The mass differences between these 5 peaks (a maximum of 10
280 numbers, if all fragments occur at differently spaced m/z) were then compiled into a list of “neutral losses”. The charged
fragment lists and the neutral loss lists of all training set external standard compounds were next combined in a frequency list,
with each charged fragment or neutral loss quantified by frequency (how many compounds in the external standard test set
exhibited that charged fragment or neutral loss among their top 5 peaks). The top 40 most common charged fragments and top
20 most common neutral losses were converted into features. The identities of these 40 most common fragments and 20 most
285 common neutral mass differences (along with possible identities and notes) can be found in tables A2 and A3, respectively.
The mass spectra of all training, test, and extrapolation set compounds were then simplified using the previously described
method (top 5 peaks extracted and mass differences between those peaks calculated). Each m/z feature was assigned the



normalized signal of that peak in the mass spectrum if the feature m/z was one of the top 5 peaks; otherwise, it was set to zero. Each neutral loss feature was assigned true or false for each compound depending on whether the neutral loss appeared in the mass differences between the 5 most significant peaks. An example mass spectral featurization for the example compound hexadecane can be found in Table A4, and the mass spectral featurization process is included in the open-source R script accompanying this publication.

3.1.2 Target Selection for Chemical Properties Modelling

The goal of chemical properties modelling is to enable inclusion of “unidentified” species in aerosol organic analysis that has previously been restricted to species for which the identity or at least chemical formula is known. One way in which complex organic mixtures are visualized and analyzed is through orientation of observed species in chemical properties spaces that have been developed and broadly utilized in the field of aerosol science. Two such spaces include the Volatility Basis Set (VBS (Donahue et al., 2006)) and the visualization by average carbon oxidation state and carbon number developed in Kroll et al., 2011, hereafter referred to as \overline{OS}_c - n_c space. Compounds can be plotted in VBS space by their O:C or \overline{OS}_c (average carbon oxidation state (Kroll et al., 2011)) against some measure of volatility, either $\log(\text{Vapor Pressure})$ or $\log(C_0)$, where C_0 is the pure component sub-cooled liquid vapor pressure in atm. In \overline{OS}_c - n_c space, compounds are plotted by their average carbon oxidation state (\overline{OS}_c) against carbon number. The ability to map novel or unidentifiable compounds in these spaces would provide critical information about the properties of the individual species, enable identification of groups of chemically distinct novel compounds deserving particular consideration, and more completely visualize the distribution of chemical characteristics for complex mixtures and potential routes of chemical transformation (e.g. oligomerization, functionalization, fragmentation) beyond the identifiable components. With these goals in mind, the properties selected to be the targets of these modelling efforts were number of carbons (n_c), O:C, \overline{OS}_c , and vapor pressure.

Carbon number, O:C, and \overline{OS}_c (based on the equation in Kroll et al., 2011) can all be directly calculated from chemical formula, which was known for each standard and ambient extrapolation compound (see section 3.2). Vapor pressure is not directly calculatable from chemical formula and not all identified compounds in the external standard and extrapolation data sets have reliable experimental vapor pressure measurements available, so structurally-based vapor pressure predictions are utilized instead. Isaacman-Vanwertz and Aumont, 2021 finds that of all structure-based vapor pressure prediction methods available, the average of predictions generated by the EVAPORATION (Compernelle et al., 2011), Nannoolal (Nannoolal et al., 2008), and Simpol (Pankow and Asher, 2008) models yields the most accurate vapor pressure prediction. These methods were therefore utilized to predict the vapor pressures of all standard and extrapolation set compounds, and the average structurally predicted vapor pressures were utilized as the “true” vapor pressures for model training and evaluation. Seven of the external standard test set species and fifteen of the extrapolation set species were incompatible with the prediction capabilities of one or more of the three structural vapor pressure prediction methods and were therefore not utilized in performance analysis. Two additional potential targets, double bond equivalent and H:C ratio, were tested but failed to produce sufficiently robust property predictions.



The final components of the chemical properties random forest models are as follows:

Targets: Carbon number, \overline{OS}_c , O:C, vapor pressure (structurally modelled)

Features: Retention index, 40 feature representation of mass spectral charged fragments, 20 feature representation of neutral mass differences between charged fragments

325 A table listing the entire set of input features for chemical properties modelling of the example compound Hexadecane can be found in table A4, and the instrument-produced mass spectrum for this species can be found in Figure A1.

3.1.3 Featurization and Target Selection for Quantification Modelling

Compound quantification factor is significantly and reliably related to retention index across all compound classes tracked, but this relationship is not linear and changes much more rapidly in some retention index windows than others. This phenomenon, caused by incomplete cold inlet trapping of species in the most volatile sensitivity region and incomplete thermal desorption of species in the least volatile sensitivity region, is illustrated in Figure A2 and is consistent with findings presented in Zhang et al., 2018. A variety of retention index corrections were tested, including the following: a) factorizing the retention indices of each compound (rounded to the nearest 100) and including as a feature in model training and testing, and b) normalizing (dividing) each compound by the raw signal of its nearest deuterated alkane internal standard, the method utilized in Zhang et al., 2018. Both methods however performed poorly in the 1600-1900 RI range, where response increases extremely rapidly with RI (Figure A2). The most reliable normalization method and the method selected for this analysis was normalizing (dividing) all compound quantification factors by the average response curve for alkanes, defined by the combination of 2 best fit exponential curves, which intersect at $RI \approx 1950$ as illustrated in Figure A2, and training on/predicting this normalized response factor rather than the raw quantification factor. The r^2 of the exponential fit of individual calibration period quantification factors around the response curve in the volatile region is .77, while the r^2 of the curve describing the less volatile region is .65. Note that these fits take into account each quantification factor of each calibration window, and are therefore influenced by the variations in the measured quantification factors of the same compounds measured at different points throughout analysis. RI-normalized response factors were translated back to predicted quantification factors for performance evaluation, as other methods of quantification do not utilize this normalization method.

345 Unlike in the case of chemical properties modelling, quantification modelling performance was significantly improved by inclusion of second dimension retention time information, and it was therefore included as a feature in response factor prediction. As a result, this approach in its current form is only usable by GCxGC-MS applications, but could be adapted to single dimension chromatography-mass spec.

350 In this analysis, continuous measurement periods (consecutively collected samples) were analysed in sequences bounded by calibration curve runs. To preserve the quantification continuity in these consecutive measurements and avoid step changes in calculated concentration that might occur due to switching between quantification factors, the two quantification factors bookending an analysis period are averaged to assign the quantification factors for samples run in that interval. To replicate this approach, the compound quantification factors were sequentially averaged to yield 5 quantification periods (the



final calibration curve experienced an instrument failure, and the last calibration period is therefore based solely on the final
355 curve).

The mass spectral featurization is described in “Mass Spectral Featurization” above.

The final components of the quantification model are as follows:

Target: Normalized response factor (RI curve-normalized, calibration period averaged)

360 Features: Retention index, second dimension retention time, calibration period, 40 feature representation of mass
spectral charged fragments, 20 feature representation of neutral mass differences between charged fragments

3.2 Training, Test, and Extrapolation Set Curation

To generate a training and test set from the external standard data, each external standard was assigned to a chemical
group (alkane, sugar, PAH, etc), and the list of external standard compounds was randomly split 80:20 (80% of compounds in
the training set, 20% in the test set) maintaining the ratios of different chemical groups. 200 possible splits were generated,
365 and the split which demonstrated the greatest similarity in median retention index and median second dimension retention time
between the test and training sets was selected to avoid potential extrapolation problems that might occur with a highly skewed
distribution of test and training compounds across the GCxGC space. This process is documented in Supporting Information.

The extrapolation set was curated from the compounds isolated from the GoAmazon samples by comparing the spectra
and retention indices of compounds to the external standard and matches in the NIST14 mass spectral database. Of the ~1500
370 unique compounds identified across 12 template samples, 63 were determined to match external standard compounds and an
additional 71 compounds were identifiable from the NIST library due to high (>800, (Worton et al., 2017)) mass spectral match
factor and retention index agreement with database entries. Based on number of silicon atoms in the assigned formulae from
the NIST identification, each chemical formula was converted to its underivatized form. Only the 71 compounds that were
identifiable from the NIST library but not from external standards were included in the extrapolation set to ensure that
375 performance metrics for the extrapolation set would not be skewed by the inclusion of species that may have been in the
training data, and to ensure that the test set and extrapolation set performance evaluations would be entirely independent. As
illustrated in Figure 2, the distribution of training, test, and extrapolation set species utilized in this work effectively span the
distribution of unknown compounds in GCxGC volatility-polarity space.

4 Model Selection, Training, and Tuning

380 The number and complexity of input features and lack of clear linear relationships between target properties and input
features in this analysis is well suited to a decision tree-based analytical approach (Bentéjac et al., 2021; Rokach, 2016).
Random forest and gradient boosting methods were both preliminarily tested for response factor prediction. Random forests
demonstrated slightly better performance and was selected for this and additional methodological reasons, as follows. Random
Forests are more robust to overfitting than gradient boosting, which is a particular concern in this case given the small number



385 of training compounds (~100) compared to the large numbers of novel environmental organics that are the intended subjects
of unverifiable modelling. Additionally, random forests perform well using the default settings and do not require extensive
tuning to achieve optimal performance (Bentéjac et al., 2021). As the aim of this work is to produce models that the atmospheric
science community, including non-experts in machine learning, can easily implement for novel compound analysis, this
robustness and simplicity is a significant advantage.

390 The training and tuning processes for chemical properties prediction are visualized in Figure 1. For each target property,
the model was trained on the external standard training set data, the curation of which is described above. As previously
referenced, random forests do not require extensive tuning, and for ease of use reasons most parameters were maintained at
their default values. Tuning primarily focused on feature restriction. Feature restriction to enforce tree diversity (mtry) was
optimized by 5-fold cross validation, with the mtry value that minimized mean average error (MAE) selected. Although random
395 forest modelling is comparatively not influenced by the inclusion of features that do not contribute significant predictive
capabilities, the inclusion of unnecessary features can contribute to overfitting of the training data which decreases prediction
performance for the test and extrapolation data sets. To address this problem, the feature importance (a measure of increase in
node purity when this feature is used in a split) of each input feature was extracted from the original predictive model. The
importance metrics were normalized by the total importance of all features to generate a percent importance score for each
400 feature. Importance distributions were highly skewed, with a relatively low number of features contributing the majority of
decrease in node purity. Features that contributed less than 1% to the total importance score were removed, and the model was
re-trained on only the important features. Extrapolation set performance improvements from removal of low importance
features was low, with an improvement in OSR² (out of sample R², defined in detail in section 5.1) on the order of 0-0.03. This
indicates that this step is not crucial for chemical properties or quantification factor prediction. The cross validation-optimized
405 mtry number, number of important features, and identity of important features for the chemical properties models (one
optimized model per property predicted) are summarised in Table 1. For quantification modelling, mtry is optimized at 44
features and 46 features meet an importance criterion of >1 %.

5 Model Performance Evaluation

5.1 Chemical Properties Modelling Performance

410 Three performance metrics are utilized to evaluate target predictions for the four chemical properties models. The first,
out-of-sample r^2 (OSR²), provides a measure of how significantly a model improves upon a baseline assumption that all target
property values are equal to the mean of those values in the training data. It approaches a maximum of 1 for perfect predictions.
The second metric, mean average error (MAE) provides the mean absolute prediction residual in the units of the target property.
This metric is particularly important, as it provides a benchmark for prediction accuracy which can be translated into
415 visualization and utilized to determine which applications are appropriate given prediction errors. The final performance



metric, root mean square error (RMSE), is also a scale dependent error metric and provides the quadratic mean of prediction residuals. The equations for these metrics are provided below:

$$OSR^2 = 1 - \frac{\sum_{i=1}^n (T_i - P_i)^2}{\sum_{i=1}^n (T_i - \bar{R}_T)^2} \quad (1)$$

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$$MAE = \frac{\sum_{i=1}^n |P_i - T_i|}{n} \quad (2)$$

$$RMSE = \sqrt{\frac{\sum_{i=1}^n (P_i - T_i)^2}{n}} \quad (3)$$

In this notation, for each test or extrapolation set compound i summed across a population of n compounds, T_i indicates the true value of the property being tested, P_i indicates the predicted value of that property, and \bar{R}_T indicates the mean of the selected property in the training data set.

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The prediction performance for the tuned and trained chemical properties model are evaluated independently on both the external standard test set (Figure 3, Table 2) and the ambient sample extrapolation set (Figure 4, Table 3). Both of these performance evaluations are important for different reasons. The external standard contains many series of highly chemically similar species (for example alkane and carboxylic acid series), meaning that the test set is likely to be more chemically similar to the training set than a real distribution of ambient organic species would be. Performance evaluation on the extrapolation set therefore provides a more realistic assessment of likely prediction accuracies on the large number of novel ambient organic compounds that are the intended focus of this modelling effort. That said, prediction performance on the external standard test set also yields important information. The external standard is designed to cover the entire space of anticipated chemical features for the environmental samples and is therefore more diverse relative to the number of compounds included compared to the extrapolation set (which is primarily CHO-type compounds). Performance evaluation on the external standard test set therefore yields more information about model performance across a broad suite of compound classes.

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5.1.1 Test Set Performance Evaluation

By all evaluation metrics applied (summarised in Table 2), performance for carbon number, O:C, carbon oxidation state, and log(VP) predictions on the external standard test set are robust. The O:C and carbon number predictions are particularly strong, with OSR^2 of .89 and .88 respectively and average errors of .072 element ratio units and 1.8 carbon number units. For context, given the range in true values from O:C= 0-1 and carbon number = 4-31, both mean average errors are approximately 7% of the range of measured values. For \overline{OS}_c and vapor pressure, the average errors normalized by the measurement range are both approximately 12%. As illustrated Figure 3, this means that the distribution of predicted properties usefully and reliably reflects the distribution of true properties and indicates that the random forest-based model provides

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445 useful information that allows a wide range of compound classes to be reliably characterized based on mass spectrum and retention index.

5.1.2 Extrapolation Set Performance Evaluation

As discussed above, while the external standard test set provides useful information on model performance across a wide range of compound types, its performance is potentially inflated by high degree of chemical similarity between the training and test set compounds. Performance evaluation on the ambient sample extrapolation set is therefore likely a more accurate indicator of prediction performance on novel or uncatalogued species. Of the four properties modelled, the performances for carbon number prediction and carbon oxidation state remain consistent or slightly improve (carbon number OSR² increases to .93), while O:C and log(VP) prediction performances decrease, both in terms of OSR² and MAE.

The weakest extrapolation set performance by far is vapor pressure prediction, which drops to an OSR² of .68. The correlation between predicted and true properties is also the weakest (as illustrated in Figure 4), with particularly large prediction residuals for the highest volatility species. For example, the extrapolation set compound with the highest vapor pressure prediction error is 1,2-Benzenedicarboxylic acid, which has a retention index of <1400 making it more volatile than the most volatile internal standard compound. While this compound does not lie outside of the volatility and polarity boundaries of the external standards in GCxGC space, is significantly more volatile than any diacid compound in the standard mixture, and the influence of double derivitization on its true ambient volatility relative to the chromatographic elution time of its derivatized form may not have been appropriately captured. Unlike the other properties targeted in this analysis, vapor pressure is not directly calculable based on chemical formula and poses challenges for many techniques; as discussed in Isaacman-Vanwertz and Aumont, 2021, molecular structure plays an important role in volatility, which significantly limits the accuracy with which techniques that identify formula but not structure (typically chemical ionization techniques) can predict the true volatility of their measured components. A more complete comparison between the random forest model's performance in vapor pressure prediction compared to other techniques used throughout the field is therefore required to provide context for vapor pressure prediction errors in the ambient sample extrapolation set (further discussed below in section 5.1.3).

For both O:C and \overline{OS}_c (which are highly related properties), extrapolation set prediction performance suffers at the high end of the oxygenation scale, although the performance reduction is far more pronounced for O:C prediction. This is due to the lack of highly oxygenated species in the external standard; random forest models do not extrapolate beyond the range of properties in the training data and therefore cannot predict O:C ratios of higher than 1.5 when that is above the maximum in the training data. The extraneously highly oxidized species for which O:C and \overline{OS}_c prediction accuracy suffers lie almost exclusively in the most volatile region instrument sensitivity, where vapor pressure prediction inaccuracies have been previously described. As a result, extrapolation set property prediction for O:C, \overline{OS}_c , and log(VP) were restricted to compounds above a retention index of 1500. As illustrated in Figure A3 and Figure 2, the significant majority of ambient analytes were above the 1500 retention index threshold, justifying the decision to restrict prediction of these properties to the retention index



window in which their performance is better optimized. In applying these techniques to the larger suite of novel species, maintaining these retention window restrictions is critical to avoid the introduction of significant sources of error.

480 Given the strong and consistent performance of carbon number and \overline{OS}_c predictions across the majority of the retention index space and between both test and extrapolation sets, the most robust visualization of chemical properties based on random forest predictions is likely to be in \overline{OS}_c - n_c space (Kroll et al., 2011). Predicting the carbon numbers and \overline{OS}_c of the known ambient compounds and superimposing the true and predicted property distributions in the \overline{OS}_c - n_c space highlights the strengths and weaknesses of chemical properties modelling. To better represent the prediction capabilities of the full chemical space and the scope of information that would be provided for properties prediction on a complex sample including hundreds
485 of individual species, all identifiable ambient compounds (including those that overlap with the external standard) were included in property prediction and visualization. As illustrated in Figure 5, the real and predicted chemical properties spaces for the ambient data set indicate both strengths and weaknesses for this application of chemically properties modelling. As noted earlier, random forest modelling does not extrapolate and has a tendency to underpredict property extremes. This is apparent in both the high \overline{OS}_c region and the high carbon number regions of the \overline{OS}_c - n_c space, where high carbon oxidation
490 states and high carbon numbers were both underpredicted. These errors could be moderated by adding more oxygenated species and higher carbon number species to the external standard, which would provide the model with more information to predict properties in these regions. In a context of extended continuity of analysis of similar sample media, this suggests an iterative approach in which the addition of new standards to a calibration mixture can be prioritized through analysing the chemical features of poorly predicted compounds in the sample media and adding new standards that replicate those features. Despite
495 the prediction errors visualized in Figure 5, the overall shape of the true chemical properties space was extremely well represented by predictions. While conclusions based on the presence or absence of extremes in predicted properties would not be appropriate, analyses based on the relative distributions of populations of interest provides valuable insight comparable to other parameterizations of compound properties from incomplete knowledge.

5.1.3 Vapor Pressure Modelling: Comparison to Prior Methods

500 Chromatography using a non-polar column is intended to separate compounds by volatility and has been used to directly predict novel compound vapor pressures in previous studies (Isaacman-VanWertz et al., 2016). It is therefore important in this context to evaluate both how significantly random forest modelling improves upon simple linear modelling of volatility based on retention index as well as how this method compares to other parameterizations of vapor pressure. As illustrated in Figure 6 and Table 4, the log(VP) prediction residuals for random forest model predictions indicate that random forest-generated
505 predictions are both more accurate and more precise than predictions by the linearized retention index method or from the Li et al., 2016 chemical formula-based parameterization, as they demonstrate a tighter distribution that is more centered around zero. The mean absolute error for random forest vapor pressure prediction is significantly lower than errors from both predictions based on retention index (t-test p value = .01) and predictions based on chemical formula (t-test p value = 3.1×10^{-5}).



510 5.2 Quantification Modelling Performance

The approach for evaluating performance for quantification modeling requires slight alterations compared to property prediction. Although the random forest model predicts the residuals of quantification factors around the retention index response normalization curve (Figure A2) rather than directly, these residuals are converted back to quantification factors for both the true and predicted properties for performance evaluation. This serves two purposes; first, other quantification methods do not use this retention index-based normalization so conversion to absolute prediction errors is necessary to compare methods, and second, a direct quantification error assessment provides more useful and applicable information about how significantly quantification errors could influence conclusions based on model-quantified data.

The test set compounds were quantified using two alternative quantification methods, *Manual* or *Closest proxy quantification* (described in Liang et al., 2021, which utilizes a combination of both), to benchmark random forest model performance. *Manual proxy quantification* entails manually assigning a compound to a chemically similar external standard based on researcher judgement on what chemical class the unidentified compound would likely belong to based on some combination of location in GCxGC space and mass spectrum. This is the current preferred method for quantification of compounds that are not in the external standard and in theory should provide the most reliable results in cases where an extremely chemically similar standard is available, but it is highly inefficient and relies upon researcher judgement calls which are difficult to standardize. *Closest proxy quantification* assigns each compound to its nearest external standard in GCxGC space, or to an average of the nearest standards within a set radius limitation. In this work, the average of the quantification factors of the 3 nearest standard species was used, as this demonstrated improved performance compared to single closest proxy quantification. This method is efficient, but it introduces potentially significant error by assigning species with different chemical characteristics (and therefore different quantification factors) the same response factor if they are sufficiently close in GCxGC space. Each test set compound was assigned to a proxy quantification factor from the training set based on each of these two methods, and each proxy compound's quantification factor at each time point was substituted as a prediction of the test set compound's quantification factor at that calibration window.

The standard performance metrics for quantification factor prediction using the random forest model, manual proxy quantification and closest proxy quantification, are compared in table 5. The random forest model significantly outperforms both other methods; it has a relatively high OSR^2 of .65 compared to negative OSR^2 values for the two proxy methods (indicating that at least on average, assuming all test compounds have the same quantification factor as the average of all training set compounds would have performed better than proxy quantification). MAE and RMSE also indicate improved performance when using the random forest model over other methods. While these metrics provide useful information on model performance, they do not reveal why the performance (particularly of the proxy methods) is so poor and do not provide useful information to evaluate likely propagation of quantification errors. Unlike for the chemical properties modelling, for quantification modelling % error is a much more important metric than absolute error, because it translates directly to how significant total quantification error across a large suite of compounds is likely to be and provides insights into underlying



biases in different methods. Figure 7 illustrates the quantification factor % error distributions of the three methods and demonstrates the improved performance of random forest modelled quantification predictions on three criteria. First, as
545 illustrated by panel A, random forest modeling produces far fewer and less extreme outlier prediction errors that are orders of magnitude different from the true values. These result when a compound that the instrument is extremely insensitive to (which would have a true extremely low quantification factor) is assigned a moderate or high quantification factor. In practice the influence of these types of quantification inaccuracies is very limited as few ambient species that the instrument is this significantly insensitive to would occur above detection limits, but they could introduce errors nonetheless. Here it is important
550 to keep in mind that each point represents a single quantification from a single calibration period; some outliers therefore indicate compounds that exhibited extremes in quantification factors during a single calibration period. This was most common among standard compounds at the edges of the instrument's sensitivity window, as these species are more significantly impacted by alterations in instrument performance. Second, as illustrated by Figure 7 panel B, the error distribution for the random forest model is significantly more centered around zero compared to either proxy model. Median random forest model
555 quantification error is -2%, compared to 17% for closest proxy quantification and 19% for manual proxy quantification. In practice, this indicates that over a large number of quantified species, random forest modeling is unlikely to introduce biased quantifications that might skew results, while the two proxy methods would likely inflate the apparent mass of novel compounds. Third, also illustrated by Figure 7 panel B (though less directly), random forest modeling produces prediction errors more tightly distributed around the median, meaning that the absolute % error distribution for random forest modelling
560 also outperforms the two proxy methods. Median absolute % error for random forest model predictions is 37%, compared to 57% for the closest proxy method and 41% for the manually assigned proxy method. The average % error improvements from random forest modeling compared to both proxy methods are statistically significant (t-test p values both < .0004), but the median absolute % error distributions of the random forest and manually assigned proxy quantifications are not significantly different based on a Mood's median test. The random forest and closest proxy method absolute % error distribution differences
565 are statistically significant, with a Mood's test p value of .001. While critical for contextualizing the potential impact of quantification errors on mass attribution of complex mixtures, a % error-based analysis of prediction accuracy is necessarily asymmetrical, as a predicted quantification factor can produce a minimum of -100 % error (the case if the predicted value were to be zero) but far more than +100% error if the quantification factor is significantly overpredicted. A symmetrical error analysis of $\log(\text{predicted quantification factor}/\text{true quantification factor})$, illustrated in Figure A4, is required to probe the
570 frequency and dynamics of underprediction in greater depth. Figure A4 demonstrates that the random forest model is more prone to underprediction outliers, but continues to outperform the other methods in achieving a narrow error distribution centered at zero.

A final benefit of random forest modeling-based quantification not captured in the performance metrics is the ability to utilize incomplete data. With proxy quantification, any standard compound that cannot be calibrated for at any point over
575 the course of an analysis cannot be used, as the species that are calibrated by that compound would not be quantifiable during the window with missing calibration data. The random forest-based quantification method relies upon the entire external



standard suite to inform corrections for instrument performance over time and can therefore produce robust quantification factor predictions even when individual standard calibration curves are missing for a particular calibration window. This allows for significantly greater flexibility in analysis, as compounds can be added to the external standard if they are observed in initial samples and still be usable to inform quantifications for periods before they were present.

In summary, random forest quantification factor modelling significantly outperforms both closest proxy and manual proxy quantification methods. It is significantly more efficient than manual proxy modeling, exhibits fewer outliers of multi order of magnitude overestimations, produces an error distribution that is more centered around zero (preventing significant biases in total mass over large numbers of quantified and summed species), and exhibits improvements in absolute percent error of predictions.

6 Conclusions

This work presents a new machine-learning based method for quantifying and predicting chemical properties of novel organic compounds observed in the atmosphere. Based on a relatively small combined training and test set of ~130 known compounds, we are able to predict the carbon numbers, vapor pressures, carbon oxidation states, and O to C ratios of ambient organic compounds with sufficient accuracy to usefully represent compound distributions in chemical property spaces that are important in atmospheric science. That these predictions are generated solely from retention indices and unit mass resolution mass spectra marks a significant step forward in ability to characterize the novel organic components of earth's atmosphere based on measurements generated from a wide range of commonly available atmospheric instrumentation. In GCxGC-MS applications, these methods contribute significant improvements in both accuracy and analytical efficiency for novel compound quantification that enable users to perform untargeted analysis of the rich complexity of data generated by advances in instrumentation. While the untargeted analysis data science techniques described in this work have been developed for and tested on atmospheric applications, they are not structurally limited in scope and could be applied to a wide range of chromatographic-mass spectral data sets to enable characterization of complex organic mixtures. The open-source R script published in supplement to this work is intended to provide a framework for groups throughout the atmospheric chemistry community to efficiently apply and adapt these methods to broadly enhance our ability to take advantage of the increasingly complex information provided by ever accelerating advances in environmental chemistry instrumentation.

Code Availability

Sample code designed for adaptation and use by other users is available in the GitHub repository associated with this manuscript (<https://github.com/ebarnesey/Ch3MS-RE>, <https://doi.org/10.5281/zenodo.6320122>). The knit R markdown including primary analysis is included in "Supporting Information."



Data Availability

Composition and metadata related to the external standard is available in the GitHub repository associated with this manuscript (<https://github.com/ebarnesey/Ch3MS-RF>, <https://doi.org/10.5281/zenodo.6320122>). Mass spectra and unique identifiers of
610 species from the ambient samples collected during the GoAmazon field campaign are available through the Goldstein Library of Biogenic and Environmental Spectra (UCB-GLOBES). All raw data can be provided by the corresponding authors upon request.

Author Contributions

AG and LY planned the measurements. EF prepared and analyzed the standards and samples. LY and RW supervised data
615 collection and preliminary analysis. PG supervised development and selection of data analysis and modeling techniques. BA contributed modeled compound property data. EF wrote the manuscript draft; LY, BA, PG, and AG reviewed and edited the manuscript.

Competing Interests

The authors declare no conflict of interest.

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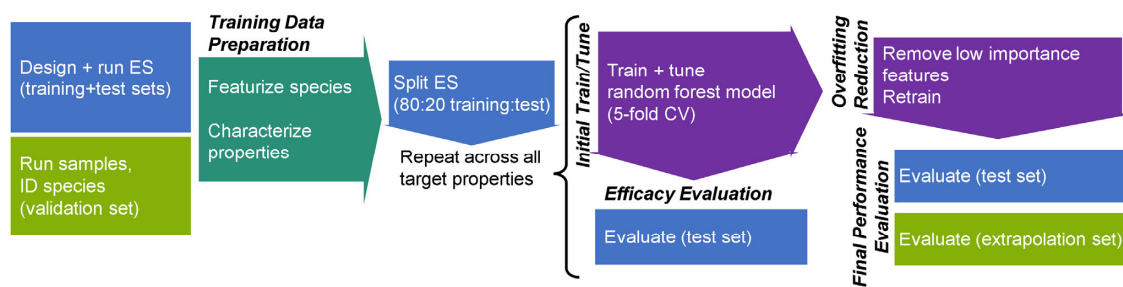
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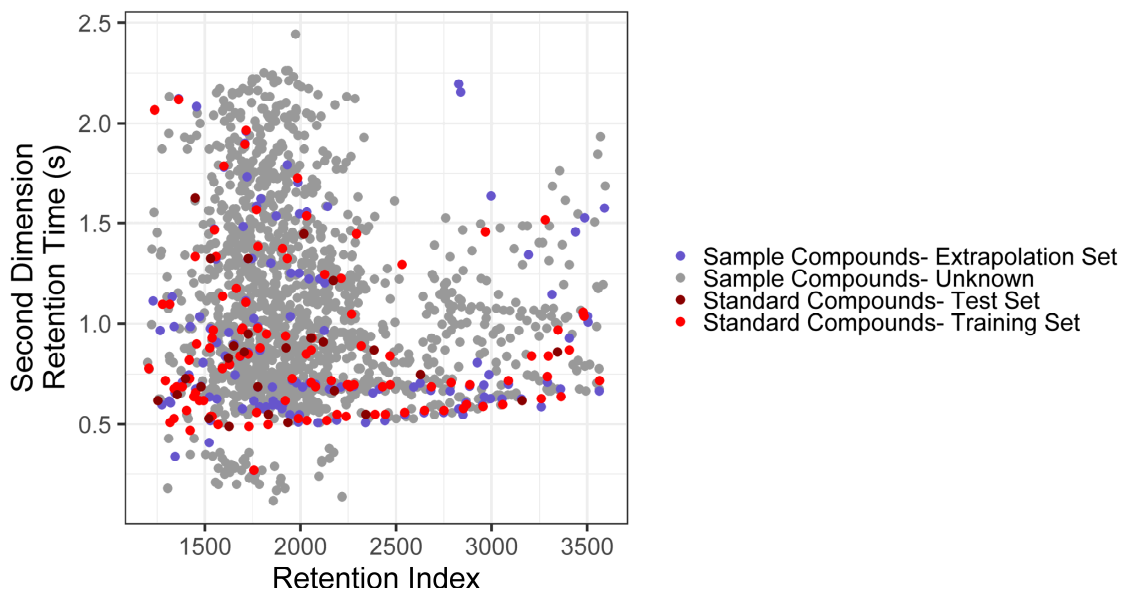
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FIGURES



780 **Figure 1:** Analytical pipeline for chemical properties modelling using a random forest model. ES indicates external standard; CV indicates cross validation



785 **Figure 2:** Distribution of training, test, extrapolation, and unidentified sample compounds in two-dimensional chromatographic chemical properties space

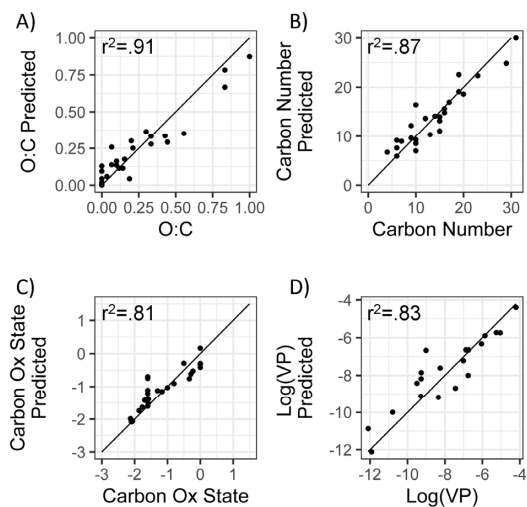


Figure 3: External standard test set true and predicted chemical properties from random forest modelling

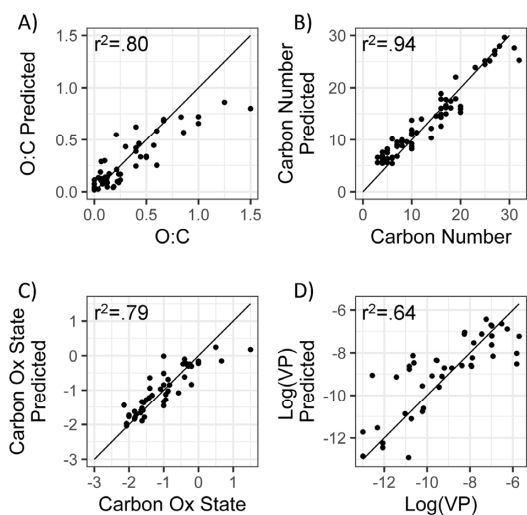


Figure 4: Ambient extrapolation set true and predicted chemical properties from random forest modelling



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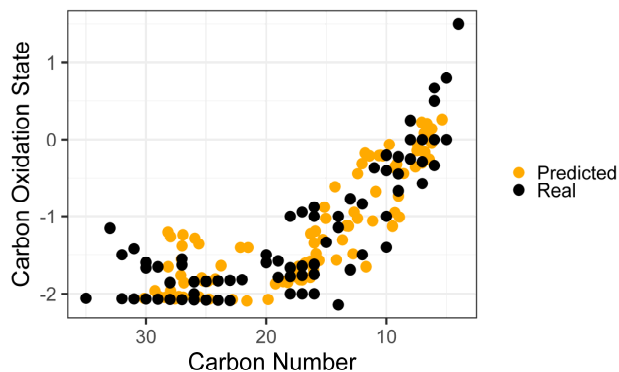
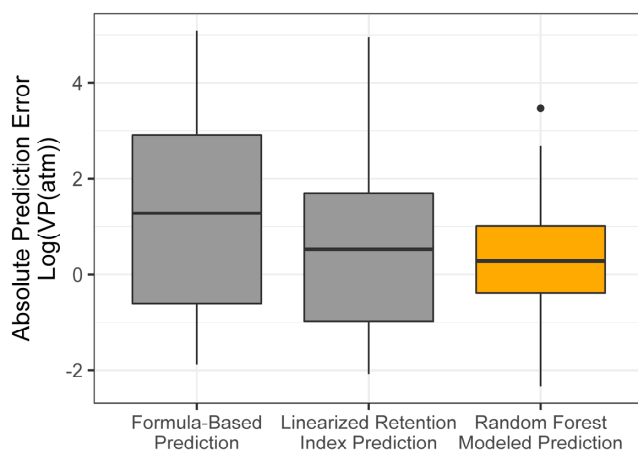
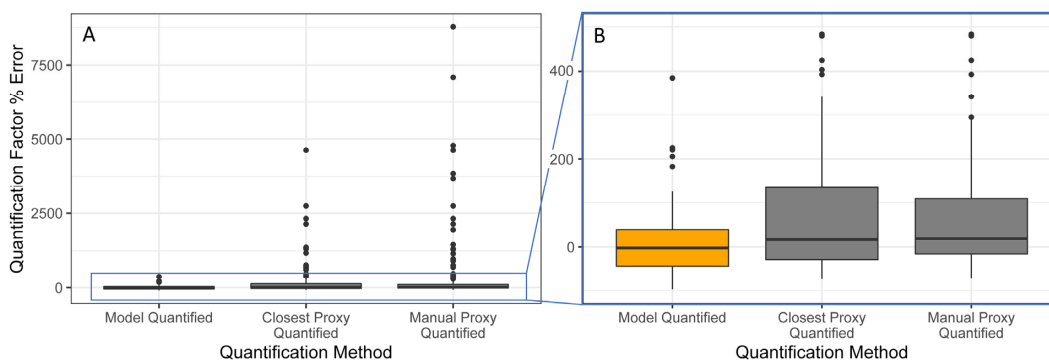


Figure 5: True versus predicted chemical properties distribution of ambient sample organic species within a Carbon Number v. Carbon Oxidation State space



795 Figure 6: Vapor pressure prediction residuals (Log(VP), VP in atm) for vapor pressure predictions of the ambient extrapolation set based on formula-based parameterization (Li et al., 2016), linearized retention index-based modelling, and random forest modelling.



800 **Figure 7: Quantification performance comparison between random forest model (orange) and two previously utilized quantification**
methods, specifically closest proxy quantification and manually assigned proxy quantification. Midline of boxes indicates sample
median, while top and bottom indicate 25th and 75th percentiles. Linear “whiskers” extend to the least extreme values within 1.5 ×
the inner quartile range of the sample. Disconnected dots indicate sample outliers that fall beyond the whisker parameters.

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TABLES

825 **Table 1: Tuning parameters and important features for chemical properties prediction models. m/z indicates charged fragment features and n indicates neutral mass difference features**

Property Model	Optimized mtry	Number of Important Features	Important Features
O:C	4	19	Retention index, m/z 41, m/z 43, m/z 45, m/z 57, m/z 69, m/z 73, m/z 74, m/z 75, m/z 103, m/z 113, m/z 147, m/z 189, m/z 204, m/z 217, n 2, n 15, n 28, n 30
Carbon Number	6	9	Retention index, m/z 41, m/z 45, m/z 55, m/z 57, m/z 73, m/z 99, n 14
Average Carbon Oxidation State	4	17	Retention index, m/z 41, m/z 43, m/z 45, m/z 55, m/z 57, m/z 69, m/z 73, m/z 75, m/z 91, m/z 93, m/z 117, m/z 119, m/z 147, n 1, n 2, n 30
Log(Vapor Pressure)	9	9	Retention index, m/z 55, m/z 73, m/z 75, m/z 129, m/z 145, m/z 147, n 3, n 30

Table 2: Performance metrics for random forest-based modelling of chemical properties of the external standard test set. “Range of true properties” units in units of property: O:C in unitless atom#/atom#, Carbon Number in atom#, average carbon oxidation state in mean charge, and Log(Vapor Pressure) in Log(atm).

Property	Out of Sample R ²	Mean Average Error	Root Mean Square Error	Range of True Properties
O:C	.89	.072	.094	0-1
Carbon Number	.88	1.8	2.4	4-31
Average Carbon Oxidation State	.79	.24	.33	(-2.1)- 0
Log(Vapor Pressure)	.82	.72	.93	(-12)-(-4.2)

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Table 3: Performance metrics for random forest-based modelling of chemical properties of the ambient aerosol sample extrapolation set. “Range of true properties” units in units of property: O:C in unitless atom#/atom#, Carbon Number in atom#, average carbon oxidation state in mean charge, and Log(Vapor Pressure) in Log(atm).

Property	Out of Sample R ²	Mean Average Error	Root Mean Square Error	Range of True Properties
O:C*	.78	.11	.17	0-1.5
Carbon Number	.93	1.8	2.2	3-32
Average Carbon Oxidation State*	.80	.25	.37	(-2.1)- (1.5)
Log(Vapor Pressure)*	.68	1.0	1.4	(-13)- (-5.7)

*Restricted to retention index > 1500

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Table 4. Error distribution metrics random forest model, retention index linear model, and formula-based predictions of vapor pressure. All reported errors in units of log(VP(atm)).

Vapor Pressure Prediction Method	Mean Error	Median Error	Mean Absolute Error	Median Absolute Error
Random Forest Model	.24	.21	1.1	.76
Retention Index Linear Model	.55	.52	1.5	1.1
Formula-Based Parameterization	1.2	1.3	2.0	1.3

840 **Table 5. Performance metrics for quantification factor prediction for three methods of unidentified compound quantification: random forest modelling, manually assigned proxy quantified, and closest proxy quantified.**

Quantification Method	Out of Sample R ²	Mean Average Error	Root Mean Square Error
Random Forest Model	.65	.00085	.0021
Manually Assigned Proxy Quantified	-4.1	.0036	.0080
Closest Proxy Quantified	-1.8	.0026	.0059

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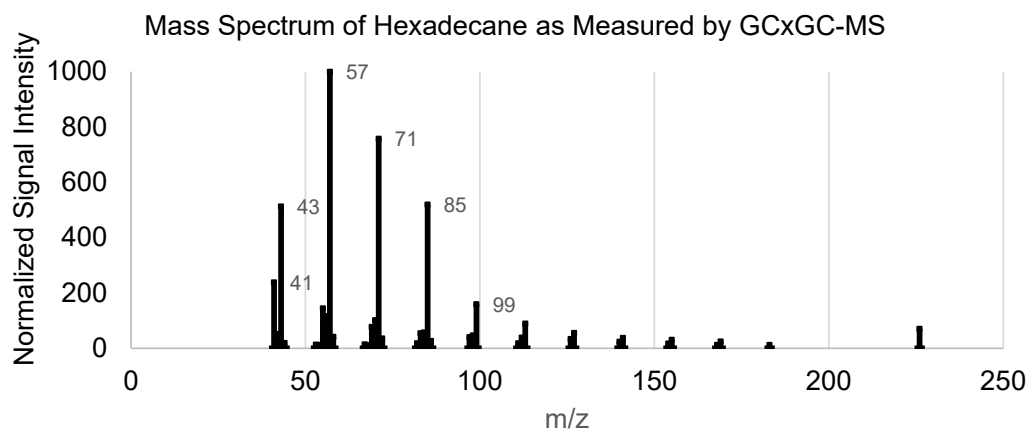
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Appendix A: Supplementary Tables and Figures

FIGURES



865 Figure A1: Mass Spectrum of Hexadecane as measured by GCxGC-MS and featured in table A2.

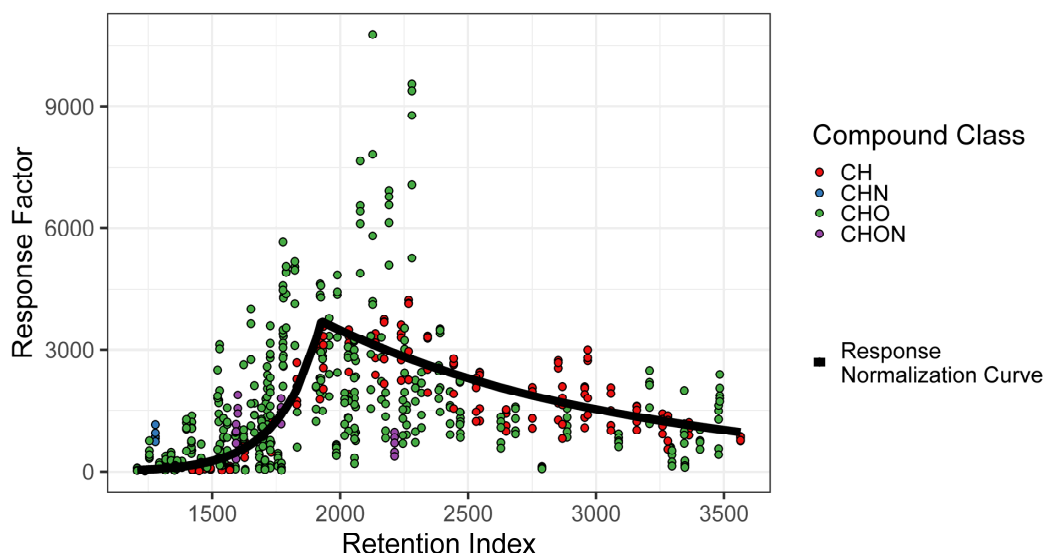
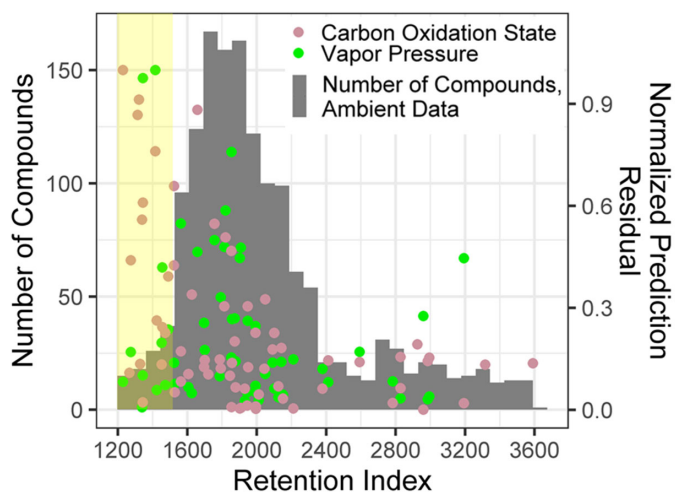


Figure A2: Quantification factor normalization curve based on average response factors of alkanes



870 **Figure A3:** Normalized prediction residuals of carbon oxidation state and vapor pressure v. retention index for ambient data compound property predictions set, overlaid with compound number distribution over the retention index for ambient data set. The yellow highlighted region indicates compounds below a retention index of 1500.

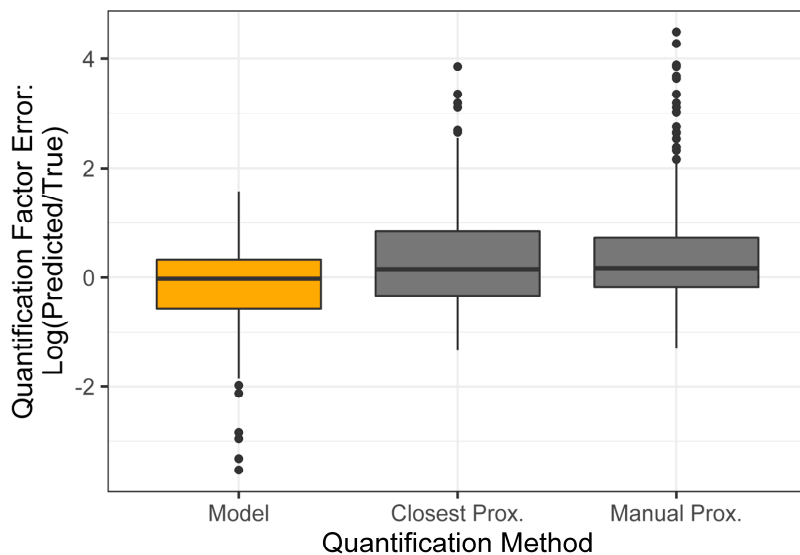


Figure A4: Quantification factor prediction errors expressed in $\text{Log}(\text{predicted quantification factor}/\text{true quantification factor})$ for test set quantification factors predicted by random forest model (orange), closest proxy, and manual proxy methods.

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TABLES

Table A1: External standard names, formulae (underivatized), retention indexes, split (training set versus test set), and manually assigned quantification proxies.

Name	Chemical Formula	Retention Index*	Split	Manual Proxy
12-OH C18 acid	C18H36O3	2470	Train	
16-OH C16 acid	C16H32O3	2429	Train	
2-ketoglutaric acid	C5H6O5	1629	Train	
3-5-dimethoxyphenol	C8H10O3	1525	Train	
4, 4 dimethoxy-benzophenone	C15H14O3	2293	Train	
4-hydroxybenzoic acid	C7H6O3	1651	Test	2-ketoglutaric acid
4-nitrocatechol	C6H5NO4	1769	Train	
4-terpineol	C10H18O	1206	Train	
9H-florenone	C13H8O	1778	Train	
a-amyrin	C30H50O	3479	Train	
abietic acid	C20H30O2	2468	Train	
anthraquinone	C14H8O2	2017	Test	xanthone
benzophenone	C13H10O	1664	Train	
beta-caryophyllene aldehyde	C15H24O2	1715	Train	
beta-caryophyllinic acid	C14H22O4	2060	Train	
beta-caryophyllonic acid	C15H24O3	1931	Train	
beta-nocaryophyllinic acid	C13H20O5	2127	Train	
beta-nocaryophyllone aldehyde	C14H22O3	1757	Train	
beta-nocaryophyllonic acid	C14H22O4	1985	Train	
beta-sitosterol	C29H50O	3406	Train	
bisabolol	C15H26O	1770	Train	
borneol	C10H18O	1254	Test	nonanol
C10 carboxylic acid	C10H20O2	1479	Test	dimethyl glutaric acid
C10 diacid (sebacic acid)	C10H18O4	1922	Train	
C12 diacid	C12H22O4	2120	Test	beta-caryophyllinic acid
C13 acid	C13H26O2	1776	Test	vanillic acid
C14 alkane	C14H30	1422	Train	
C14 diacid	C14H26O4	2317	Train	
C16 alkane	C16H34	1626	Train	
C16 acid	C16H32O2	2078	Train	
C17 alkane	C17H36	1730	Test	C17 alkane
C17 acid	C17H34O2	2177	Test	linoleic acid
C18 alkane	C18H38	1830	Train	
C18 acid	C18H36O2	2280	Train	



C19 alkane	C19H40	1934	Test	C20 alkane
C20 alkane	C20H42	2034	Train	
C21 alkane	C21H44	2137	Train	
C22 alkane	C22H46	2238	Train	
C22 acid	C22H44O2	2684	Train	
C23 alkane	C23H48	2341	Test	C24 alkane
C24 alkane	C24H50	2443	Train	
C24 acid	C24H48O2	2886	Train	
C25 alkane	C25H52	2545	Train	
C26 alkane	C26H54	2649	Train	
C26 acid	C26H52O2	3088	Train	
C27 alkane	C27H56	2750	Train	
C28 alkane	C28H58	2852	Train	
C28 acid	C28H56O2	3291	Train	
C29 alkane	C29H60	2955	Train	
C30 alkane	C30H62	3058	Train	
C31 alkane	C31H64	3159	Test	C30 alkane
C32 alkane	C32H66	3259	Train	
C33 alkane	C33H68	3363	Train	
C35 alkane	C35H72	3564	Train	
C7 acid	C7H14O2	< 1400	Train	
C8 acid	C8H16O2	1293	Train	
C9 acid	C9H18O2	1381	Train	
C9 diacid (azelaic acid)	C9H16O4	1822	Train	
cholesterol	C27H46O	3209	Train	
chrysene	C18H12	2531	Train	
cis-vaccenic acid	C18H34O2	2259	Train	
citronellol	C10H20O	1338	Train	
cycloisolongifolene	C15H24	1355	Test	pyrocatechol
DEET	C12H17NO	1600	Train	
deoxycholic acid	C24H40O4	3347	Train	
dibenz(ah)anthracene	C22H14	3280	Train	
dimethyl glutaric acid	C7H12O4	1456	Train	
dodecyl benzene	C18H30	1920	Train	
eicosanol	C20H42O	2390	Train	
ergosterol	C28H44O	3296	Train	
erythritol	C4H10O4	1528	Train	
FAME16 (methyl palmitate)	C17H34O2	1957	Train	
FAME18 (methyl stearate)	C19H38O2	2161	Train	



farnesol	C15H26O	1832	Test	bisabolol
galactosan	C6H10O5	1684	Train	
gamma dodecalactone	C12H22O2	1709	Train	
glyceric acid	C3H6O4	1352	Train	
hexadecanamide	C16H33NO	2212	Train	
hexadecanol	C16H34O	1989	Train	
homosalate	C16H22O3	2054	Test	beta-caryophyllinic acid
hydroquinone	C6H6O2	1420	Train	
ionone	C13H20O	1449	Train	
isoeugenol	C10H12O2	1591	Train	
isopimaric acid	C20H30O2	2385	Test	C14 Diacid
ketopinic acid	C10H14O3	1530	Test	pinonic acid
levoglucosan	C6H10O5	1726	Train	
linoleic acid	C18H32O2	2245	Train	
lupeol	C30H50O	3483	Train	
maltol	C6H6O3	1316	Train	
mannosan	C6H10O5	1706	Test	galactosan
MBTCA	C8H12O6	1776	Train	
Me-OH-glutaric acid	C6H10O5	1623	Test	2-ketoglutaric acid
monopalmitin	C19H38O4	2628	Test	monostearin
monostearin	C21H42O4	2788	Train	
nonanol	C9H20O	1318	Train	
octadecanal	C18H36O	2056	Train	
octadecanol	C18H38O	2191	Train	
octadecanone	C18H36O	2031	Train	
oleic acid	C18H34O2	2251	Train	
palmitoleic acid	C16H30O2	2056	Train	
p-anisic acid (4-methoxybenzoic acid)	C8H8O3	1544	Train	
pentadecanone	C15H30O	1726	Test	pinic acid, isomer 1
perylene	C20H12	2967	Train	
phthalic acid	C8H6O4	1714	Train	
phthalimide	C8H5NO2	1593	Train	
pinic acid, isomer 1	C9H14O4	1692	Train	
pinic acid, isomer 2	C9H14O4	1697	Train	
pinonic acid	C10H16O3	1550	Test	hexadecanamide
pyrene	C10H16	2171	Train	
pyrocatechol	C6H6O2	1339	Train	
quinoline	C9H7N	1278	Train	



resorcinol	C6H6O2	1399	Test	hydroquinone
retene	C18H18	2267	Train	
Sesquiterpene 1 [†]	C15H24	1404	Train	
Sesquiterpene 2 [†]	C15H24	1442	Test	Sesquiterpene 3
Sesquiterpene 3 [†]	C15H24	1449	Train	
Sesquiterpene 4 [†]	C15H24	1451	Train	
Sesquiterpene 5 [†]	C15H24	1471	Train	
Sesquiterpene 6 [†]	C15H24	1493	Train	
Sesquiterpene 7 [†]	C15H24	1537	Train	
Sesquiterpene 8 [†]	C15H24	1569	Train	
Sesquiterpene 9 [†]	C15H24	1610	Train	
sinapinaldehyde	C11H12O4	2032	Train	
squalene	C30H50	2868	Train	
stigmaterol	C29H48O	3344	Test	ergosterol
syringaldehyde	C9H10O4	1726	Test	9H-florenone
syringic acid	C9H10O5	1924	Test	C10 diacid (sebacic acid)
syringol	C8H10O3	1418	Train	
threitol	C4H10O4	1521	Test	erythreitol
triacetin	C9H14O6	1362	Train	
tridecanal	C13H26O	1537	Train	
vanillic acid	C8H8O4	1789	Train	
vanillin	C8H8O3	1558	Train	
verbenone (-)	C10H14O	1237	Train	
xanthone	C13H8O2	1906	Train	

*Normalized by deuterated alkane standard series

880 [†]Isomer identity undetermined, only quantification factor and properties related to chemical formula included in modelling

Table A2: 40 most common charged fragments featurized for mass spectral featurization, with possible formulae and implications of published peaks.

Fragment m/z	Possible Formulae	Notes
41	C3H5+	
43	C3H7+, C2H3O+	Propyl group, ketone indicator
45	CHO2	Carboxyl indicator, underivatized
55		
56		
57	C4H9+, C3H5O+	Signature alkane fragment, ketone/ester
67		
69		
71	C4H7O+	Ketone/ester
73	Si(CH ₃) ₃ +	Indicates derivatization and therefore presence of OH group



74		
75		
77	C6H5+	phenyl
79		
81		
83		
85		
91		
92		
93	C6H5O+	Oxygenated aromatics
95		
99		
103		
105		
107		
109		
111		
113		
117		
119		
121		
129		
131		
132		
135		
145		
147		
189		
204	Si2C8H20O2+	Indicative of sugars
217	Si2C9H21O2+	Indicative of sugars

885 **Table A3: 20 most common neutral mass differences between charged peaks, selected for mass spectral featurization, with possible formulae and implications of commonly reported neutral losses.**

Neutral Loss/Mass Difference (amu)	Probable Formulae/ Interpretation	Notes
1	Loss of H	
2		
3		
4		
6		
8		
10		
11		
12		
13		
14		
15	CH3	Methyl
16	O	Alcohol- derivatization agent loss



18		
20		
26		
27		
28	CO	Carbonyl
30		
42		

Table A4. Full chemical properties modelling features for Hexadecane

Feature	Feature Class	Feature Input
Retention Index (d-alkane normalized)	Chromatography	1627
m/z 41	Mass Spectrum Common Fragment	238
m/z 43	Mass Spectrum Common Fragment	512
m/z 45	Mass Spectrum Common Fragment	0
m/z 55	Mass Spectrum Common Fragment	144
m/z 56	Mass Spectrum Common Fragment	116
m/z 57	Mass Spectrum Common Fragment	999
m/z 67	Mass Spectrum Common Fragment	0
m/z 69	Mass Spectrum Common Fragment	0
m/z 71	Mass Spectrum Common Fragment	757
m/z 73	Mass Spectrum Common Fragment	0
m/z 74	Mass Spectrum Common Fragment	0
m/z 75	Mass Spectrum Common Fragment	0
m/z 77	Mass Spectrum Common Fragment	0
m/z 79	Mass Spectrum Common Fragment	0
m/z 81	Mass Spectrum Common Fragment	0
m/z 83	Mass Spectrum Common Fragment	0
m/z 85	Mass Spectrum Common Fragment	519
m/z 91	Mass Spectrum Common Fragment	0
m/z 92	Mass Spectrum Common Fragment	0
m/z 93	Mass Spectrum Common Fragment	0
m/z 95	Mass Spectrum Common Fragment	0
m/z 99	Mass Spectrum Common Fragment	0
m/z 103	Mass Spectrum Common Fragment	0
m/z 105	Mass Spectrum Common Fragment	0
m/z 107	Mass Spectrum Common Fragment	0
m/z 109	Mass Spectrum Common Fragment	0
m/z 111	Mass Spectrum Common Fragment	0
m/z 113	Mass Spectrum Common Fragment	89
m/z 117	Mass Spectrum Common Fragment	0
m/z 119	Mass Spectrum Common Fragment	0
m/z 121	Mass Spectrum Common Fragment	0
m/z 129	Mass Spectrum Common Fragment	0
m/z 131	Mass Spectrum Common Fragment	0
m/z 132	Mass Spectrum Common Fragment	0
m/z 135	Mass Spectrum Common Fragment	0



m/z 145	Mass Spectrum Common Fragment	0
m/z 189	Mass Spectrum Common Fragment	0
m/z 204	Mass Spectrum Common Fragment	0
m/z 217	Mass Spectrum Common Fragment	0
loss of 1	Mass Spectrum Neutral Loss/Mass Diff.	FALSE
loss of 2	Mass Spectrum Neutral Loss/Mass Diff.	TRUE
loss of 3	Mass Spectrum Neutral Loss/Mass Diff.	FALSE
loss of 4	Mass Spectrum Neutral Loss/Mass Diff.	FALSE
loss of 6	Mass Spectrum Neutral Loss/Mass Diff.	FALSE
loss of 8	Mass Spectrum Neutral Loss/Mass Diff.	FALSE
loss of 10	Mass Spectrum Neutral Loss/Mass Diff.	FALSE
loss of 11	Mass Spectrum Neutral Loss/Mass Diff.	FALSE
loss of 12	Mass Spectrum Neutral Loss/Mass Diff.	FALSE
loss of 13	Mass Spectrum Neutral Loss/Mass Diff.	FALSE
loss of 14	Mass Spectrum Neutral Loss/Mass Diff.	TRUE
loss of 15	Mass Spectrum Neutral Loss/Mass Diff.	FALSE
loss of 16	Mass Spectrum Neutral Loss/Mass Diff.	TRUE
loss of 18	Mass Spectrum Neutral Loss/Mass Diff.	FALSE
loss of 20	Mass Spectrum Neutral Loss/Mass Diff.	FALSE
loss of 26	Mass Spectrum Neutral Loss/Mass Diff.	FALSE
loss of 27	Mass Spectrum Neutral Loss/Mass Diff.	FALSE
loss of 28	Mass Spectrum Neutral Loss/Mass Diff.	TRUE
loss of 30	Mass Spectrum Neutral Loss/Mass Diff.	TRUE
loss of 42	Mass Spectrum Neutral Loss/Mass Diff.	FALSE